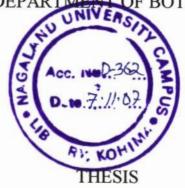
# MELISSOPALYNOLOGICAL STUDIES IN AND AROUND MOKOKCHUNG DISTRICT

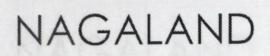


BY
MISS TEMSUNUNGLA
DEPARTMENT OF BOTANY



1)

SUBMITTED
IN PARTIAL FULFILMENT OF THE REQUIREMENT
OF
THE DEGREE OF DOCTOR OF PHILOSOPHY
IN
BOTANY
OF NAGALAND UNIVERSITY, Hqrs:LUMAMI
2005





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#### **CERTIFICATE**

This is to certify that the accompanying thesis entitled "Melissopalynological Studies In And Around Mokokchung District" submitted in fulfillment of requirements for the award of the Degree of Doctor of Philosophy in Botany, by Miss. Temsunungla contains a bonafide research work carried out by her at Nagaland University, Lumami.

Further, certified that no part of this thesis has been submitted anywhere for any other degree or diploma. The assistance and help received during the course of study by the candidate and source of literature have duly been acknowledged. This thesis embodies the work of candidate herself.

(S.K.Chaturvedi)



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### **CERTIFICATE**

This is to certify that the present Thesis entitled "Melissopalynological studies in and around Mokokchung District" submitted as the partial fulfillment for obtaining the degree of Doctor of Philosophy in Botany of Nagaland University, is an original piece of work carried out by me for the first time. None of the portion or portions of this work i.e., photographs, figures or tables has been copied or reproduced from anywhere else.

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To all of them I convey my Good Wishes.

Temsunungla

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## INTRODUCTION

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#### -1-

#### INTRODUCTION

"Melissopalynology is the branch of palynology deals with the study of pollen content in a particular honey sample". Over hundred years the literature pertaining to the study of pollen in honey has been termed or spelled several ways including melissopalynology, Mellittopalynology and melittopalynology. According to panton's Botanical dictionary (1868) both "Melissa" and "melitta" means a 'Bee". The word "melliferous" comes from the Latin word "melifer" means honey and the suffix-ous meaning "having full off" or characterized by (Bryant, 2001). The International Commission for Bee Research (ICBR) uses the term "Melissopalynology" for the study of pollen and honey, which is the term used throughout this thesis.

#### The Honey

For survival, honey bee require four natural resources, viz., Water, resin, nectar and pollen (Seedley, 1985). Water is used to cool the hive and to dilute the honey fed to the young broods. Resin is utilized to repair the hive e.g., sealing of decaying wood and to plug up holes. Nectar is the major source of carbohydrates from which honey bee obtain their energy. According to Winston (1987) it can be fed directly to the brood or to the adults. The nectar is collected by the workers bees in their honey stomachs and it is usually transferred to hive workers for processing into honey. These workers add an enzyme, to this nectar, which is secreted by their hypopharyngeal glands. This enzyme breaks down the

nectar into simple forms of sugars, which are easier for the bees to digest. These enzymes protect the stored honey from bacteria. The water in the nectar is then evaporated off of the worker's tongue. The nectar is placed into cells and fanned to further reduce water in it. According to Winston (1987), this process reduce the moisture content of nectar less than 18%. Once the evaporation process is complete, the nectar is considered "ripenedquot" and is called honey. This honey is stored in the cells of the honey comb and is capped with wax.

It has been estimated that to make one pound of honey, honey bees must visit about two million flowers, fly a total of about 50,000 miles and carry about 73,000 loads of nectar back to the hive According to Teale (1942), during the main blooming periods it is common for the bees from a single hive to visit as many as 2,50,000 flowers during the course of a single day. Winston's (1987) studies reveal that each worker bee is able to carry a load of nectar equal to one - half its total weight and during its lifetime one worker will collect enough nectar to produce about 1/12 of a teaspoon of honey. During foraging, a honey bee consumes 0.5 mg of ripe honey per kilometer of flight. Feeding a bee larva from the egg to maturity requires about 142 mg of honey.

#### SOURCE OF POLLENS IN HONEY

According to Gary (1975) pollen is the major source of proteins, fatty substances, minerals and vitamins for honey bees. Pollens are essential for the growth of larvae and young adult bees (Dietz, 1975). The pollen

grains are collected by honey bees from the anthers of flowers by using their tongue and mandibles. At the time of nectar and pollen collection, body parts of honey bee are sprayed with pollen grains. These pollen grains are combed and mixed with pollen from her mouth and transferred it to the corbicula, or "Pollen - basket" on her posterior pair of legs. At hive, worker bees pack the pollen into the comb. To prevent bacterial growth and pollen germination a phytocidal acid is added to the pollen as it is packed into the comb. Other enzymes produced by worker bees are also added which prevent anaerobic metabolism and fermentation, thereby, enhancing the longevity of the stored pollen. The completely processed pollen grains for storage kept in comb is referred to as "bee bread" and is ready for later consumption by the bees. According to Alfonsus (1933), the protein source needed for rearing one worker bee from larval to adult stage require approximately 120 to 145 mg of pollen. An average size bee colony will collect about 20 to 57 kg of pollen a year (Armbruster 1921, Eckert, 1942).

The pollen grains are collected from entomophilous as well as anemophilous plants also. However, according to Bryant (2001) wind pollinated species of Salix (Willow), Quercus (Oab), Celtis (hach berry) and many species of grasses (Poaceae) as well as some of the wind – pollinated type of composites (Asteraceae) are considered important pollen sources for foraging by honey bees. Airborne pollen is another potential source of pollen in honey. Many types of airborne pollen produced mostly by wind – pollinated plants that are not usually visited by honey bees can enter a hive through wind currents. But the number

of airborne pollen grains are usually few in number. Mostly the airborne pollen can settle out in areas where open comb cells are being filled with nectar.

#### Importance of Melissopalynological Studies

Pollen is an essential tool in the analyses of honey. The floral nectar source utilized by honey bees for honey production can be determined by the identification of pollen taxa (Lieux, 1975, 1978; Moar, 1985; Louveaux et al 1970; Sawyer, 1988; Van Der Ham et al, 1999). Thus, the relative pollen frequency is often used to verify and label a honey sample as to the major or minor nectar sources. Identifying and quantifying the pollen in honey samples is one of the best ways to determine the range of nectar types used to produce a honey and therefore label it correctly based on actual foraging resources. Another most important reason that pollen analysis of honey are often required is to identify the geographical source of origin as well as assessment of phytogeography of the region. The combination of anemophiles and entomophiles found in a honey sample will often produce a pollen spectrum that is unique for the specific geographical region where it produced. Trade agreements, import tariffs and legal trade restrictions require accurate labeling of honey before it can be sold in the market of leading honey producing nations of the world. However, in India no such rules and regulations are strictly followed for the honey trade and lead to the adulteration of the honey with sugar syrup.

### Unifloral and Multifloral honey

After experimental investigation on European honey for thirteen years Demianowicz (1961, 1964) realized that the relative pollen count in honey did not always reflect the primary floral and nectar Demianowicz (1964) attempted to identify the pollen characteristics of 45 different types of unifloral honey that are common to various regions of Europe. For her experiments she used small bee-hive with one queen and 300-400 workers. These bees were allowed to feed on the flowers of only one species of plant. She extracted the honey of each hive and considered it for absolute pollen concentration (APC) for the flower type being tested. On the basis of her experiments she developed 18 different categories of plants with respect of their APC values in honey are under or over represented. Thus, for each category she assigned an "average number" that she called her "Pollen Co-efficient Classes". According to this pollen co-efficient value could be used as a guide for determining the true unifloral nature of a honey sample, irrespective of the data represented by the relative pollen concentrations.

Demianowicz's Class 1 unifloral type should contain not more than 740 pollen grains per 10 gm of honey. In this class she kept Asclepias (milkweed) with pollen co-efficient value 32. Each additional class should have APC values that are upto twice as high as the previous category. In her Class 2 of unifloral honey types the APC value should be between 750-1500 per 10 gm of honey. In this category

she kept Robinia pseudoacacia (white acacia, locust), Cucumis (cucumber) and Epilobium (Fireweed). In Class 18 with prolific pollen types she kept Myosotis (Forget -me-not) which produced nearly 200 million pollen grains per 10 gm of honey.

A number of scientists have produced tables and charts noting what they believe should be the "expected" percentages of relative pollen in unifloral types. In New Zealand, Moar (1985) points out that since 45% of a single pollen type is the universal "minimal" amount needed for a honey to be classified as unifloral. However, according to Sawyer (1981,1988) if the pollen grains of a particular plant species are 45% of the total pollen count in honey sample, the honey sample should be called "unifloral" and if none of the plant species, represented through the pollen in a particular sample, reaches upto 45%, the honey should be called as "multifloral".

#### Review of Literature

First of all the pollen contents of various Swiss, French and other Europeans honey was examined by Pfister (1895). He tried to demonstrate the possibility of determining the geographical origin of honey from the pollen within it. The first published report on the pollen content of honey from United States of America is by W.J.Young at the beginning of twentieth century. He published a paper in 1908 to determine if pollen studies could be used in the future to "Judge the adulteration of a sample" (Young, 9108). Based on his work, Young determined that the range of pollen concentration values varied from a

low of 123 pollen grains / g to a high of 5,410 grains commonly found in U.S. honey and discussed the importance of protecting honey samples from airborne contaminants. He also reported the presence of insect body parts, fragments of the comb, Fungal spores, dust, pollen etc. in honey. The first European, who published his work on Melissopalynology was Fehlman. Fehlman (1911) published report on the pollen spectra found in various examples of Swiss honey (Maurizio, 1951; Maurizio and Louveaux, 1965; Lieux, (1969). He was the first European to use pollen as a way to identify and differentiate honeydew from nectar honeys and to demonstrate that pollen contents were the key to determine the nectar sources in honey samples. In United States it was Parker (1923) who conducted a study of bees and the honey they collect. He described 28 different kinds of pollen grains collected by honeybees and included photographs of the 12 most important ones. He was convinced that the pollen content in honey was a valuable tool for identifying the foraging sources used to make it.

Betts (1923, 1925) worked on English honey and published sketches of 15 different kinds of pollen sources found in it. She suggested that flowers from herbarium specimens could be used as a source of pollen to make comparative reference samples. A few years later, Allen (1928a) noted that same pollen grains remain on the surface of the honey, instead of becoming mixed with the honey like other types of pollen. He was also the first to report that pollen found mixed with nectar could come from sources other than the nectar plant's own anther and pollen (Allen, 1928b). He had also reported

floating pollen grains on the surface of honey and reasoned that due to their lesser density than honey some pollen grains float on its surface. Allen (1928, a, b) also noted that air borne pollen grains could easily contaminate honey when combs were being removed from hives and also during the subsequent honey extraction process. But according to him contamination of honey through air borne pollen is a minor problem and pollen in honey mostly reflects the actual floral sources used to make the honey. However, Allen (1929) was the first researcher to focus some of the problems of conducting accurate melissopalynology analyses. He also emphasized the difficulties of pollen identification in melissopolynology (Allen, 1928 a, b; 1929). His most valuable contribution is proposed classification system for English honey. However, his remark regarding English honey which states that "One should doubt the origin of a honey sample as being English if the sample contains six-grooved pollen grains" has been disapproved by Bryant (2001) on the bases that many Lamiaceous taxa such as Mentha (mint), Thymus (thyme) and Salvia (Sage) are now grow in English garden and foraged by the honey bees. The foundation of melissopalynological research in Europe was laid down by the five volume work of Zander (1935, 1937, 1941, 1949, 1951). In these volumes he has described descriptions, drawings and photographs of pollen that he found in various types of European honey. He also described fungal spores and hyphae from honey samples. His voluminous work on English melissopalynology has brought him the honours to be called as "leader in melissopalynology research in Europe" by Maurizio and Louveaux (1965).

In United States, Pellett (1930) and Pammel et al (1930) published "American honey plants and "Honey plants of Iowa" respectively. However, there is no other report of melissopalynological research in United States during this period i.e.1930s. Later, Oertel (1939) published the results of his seven—year study on the sources and blooming periods of plants though to be principal honey bee nectar sources in various regions of the United States (Bryant, 2001).

Whitcomb and Wilson (1929) studied the cause of dysentery in honey bees and reported that many of the pollen grains sucked into a bee's honey stomach along with nectar were quickly removed through a process of filtering. These authors reported that once nectar reaches a bee's honey stomach it is filtered within 10 minutes and only pure nectar is left in honey stomach. These authors concluded that the filtering of nectar in their honey stomach is a device to remove unwanted pollen and fungal spores so as to prevent future honey from pollen and spore germination. But, Snodgrass and Erickson (1992) have described the process of nectar filteration in detail. However, due to filteration of nectar, lots of pollen grains and fungal spores and other unwanted plant products e.g. hairs etc., are also defecated by these bees in bulk which is called "Yellow Rain" and this phenomenon comes in the form of small yellow spots on cars, buildings, leaves of plant etc.

Todd and Vansell (1942) published their work on the relationship and importance of pollen in honey at United States

Department of Agriculture (USDA) California. These workers found that honey bees do not reproduce or lay eggs if feed on sugar syrup only

but if the pollen were added to the feeding syrup, egg laying in the hive started within twelve hours. These workers also made qualitative and quantitative melissopalynological studies of over 2,600 samples of nectar collected from various apiaries in California.

Recently, hundreds of papers have been published on the melissopalynological studies in Europe and USA for maintaining the quality of honey. Although, in India the uses of honey are multifarious as it is used in various rituals by Hindu communities right from the birth of the child up to the death, yet, the measures taken to control the quality of honey are not sufficient. The literature on melissopalynological studies in India is also meager.

In India during 1950's onward Deodikar et al. (1958) and Chaubal and Deodikar. (1965) have reported melissopalynological data from Mahabaleshwar and Western ghats respectively. These workers have reported major honey yielding plants from the invetigatd regions. However, Chaubal (1980) reported that most of the pollen grains in honey comes from member of Acanthaceae and Compositae and emphasized a need of preparing a critical palynological data for most of the members of Acanthaceae (Chaubal Deodikar, 1965; Chaubal 1966) and Compositae (Chaubal and Deodikar. 1966 – 1967; Chaubal, 1976), to facilitate the identification of pollen from loads and honey samples especially from Western ghats of India. However, during their studies Chaubal and Deodikar, 1963; Deodibar et al. 1958 B, 1958 C) have also reported occurrence of poisonous pollen grains in honey samples collected from plains of Sayhadri, but such poisonous pollen grains did

not occur in the samples of Sagarmal region of Kolhapur district of Maharashtra, India (Chaubal, 1980)

During 1970's the melissopalynological studies have geared up and at NBRI, Lucknow, Nair, 1964; Sharma, 1970; Chaturvedi, 1973, 1983, 1989; Garg and Nair, 1994; Garg, 1996), have also published a few reports of melissopalynological studies in that region of India.

At the Department of Botany Allahabad University, Chaturvedi (1983) have enlisted several plant species visited by honey bees for pollens only, for nectar only and for pollen and nectar both. His work is based on the field survey and observations in and around Allahabad district (Chaturvedi, 1993; Chaturvedi and Chaturvedi, 2001). In India, other centers for melissopalynological studies include North Bengal University (Department of Botany), Allahabad University (Department of Botany), Bangalore University (Department of Botany), Dr. B.R.Ambedkar University, Agra (Department of Botany), Haryana Agriculture University, Hisar, Manipur Agriculture University, Imphal and certain NGO's like Century foundation, Bangalore, Bioved Research And Communication Society, Allahabad are engaged in similar type of investigations.

In Nagaland, Chaturvedi and Temsunungla (2004) have published first hand report of melissopalynological studies and concluded that seasonal honey samples, collected during winter and autumn, from the bee hive of *Apis indica* (The native Indian honey bee) kept in the CTC campus of Ungma village of Mokokchung district, varies in the frequency of pollen grains and the colour of the honey. Winter samples

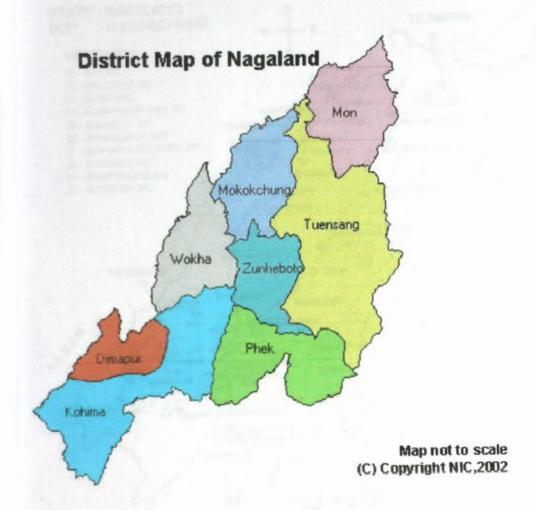
of honey exhibits 50.81 % of Asteraceous pollen, whereas Autumn samples show maximum number of poaceous pollen grains i,e. 23.34 %.

The present investigation deals with the melissopalynological studies of seasonal honey samples collected from three villages viz., Khensa, Kubza and Ungma, of the Mokokchung district in Nagaland State, (Maps: 1-3).



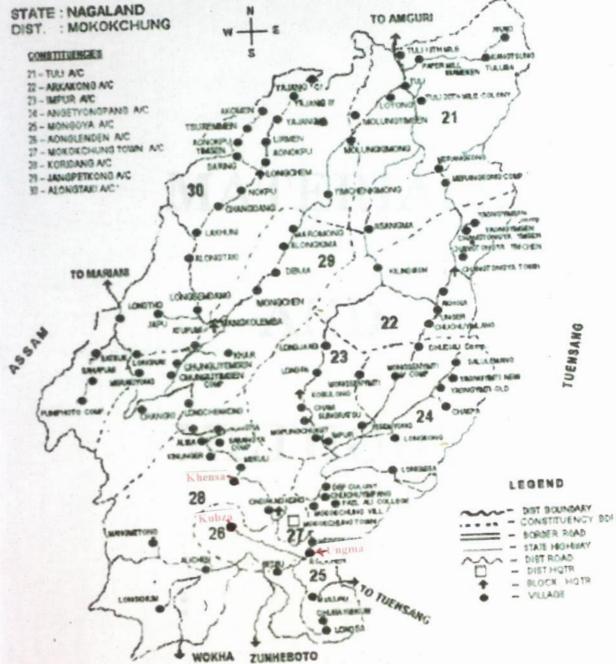
MAP-1

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MAP-2

# MAP OF MOKOKCHUNG TO AMGU



MAP-3

## **MATERIALS**

**AND** 

## **METHODS**

#### MATERIALS AND METHODS

Melissopalynological studies involves various methodology such as collection of honey samples, collection of plant species for preparation of reference slides. Storage of honey samples, extraction of pollen grains, preparation of slides of pollen-grains, photomicrography and identification of pollen grain.

These methodologies are described as follows -

#### Collection of honey samples

Honey samples for the present investigation have been collected from three different localities, viz., CTC campus of Ungma village, Kubza village and Khensa village of Mokokchung District, Nagaland. For the collection of honey samples two seasons were selected, winter (January – February) and Autumn (August– September).

The samples were collected from the honey comb of the Apiary directly into the PVC 250 milliliters bottles which are temperature resistant and autoclavable. These samples were marked for the localities and date of collection and kept at room temperature in the laboratory at Botany Department, Nagaland University, Lumami for further investigation.

#### Storage of honey samples

The collected samples were stored in laboratory conditions at an ambient temperature ranges from 7°C - 16°C during winter months with RH 20% - 60% and 16°C - 30°C during summer months with RH 60% - 90%.

# Extraction of pollen grains and preparation of slides from honey samples

There are various techniques for pollen extraction from honey samples as given by Sawyer (1981,1988), Ricciardelli D'Albore(1998), Lieux (1969), Bryant and Jones(2001), Barth et al (2004), Jones et al (1995), Ohe et al (2004), Louveaux et al (1978), Herrero et al (2002), Lutier and Vaissiere

(1993) and others and thus. However, for the present investigation, Sawyer's (1981) method has been modified and used for the pollen extraction from the honey samples. However, the methodology used for the present investigation is as follows –

- (1). 10 grams of honey is mixed with 20 milliliters of hot water. After thorough mixing, the Solution is placed in two tubes and centrifuged for 10 minutes at 2500 rpm.
- (2). The supernatant solution is decanted and the residue is transferred to one tube. Both are filled with water (to balance the centrifuge) and recentrifuged for 5 minutes.
- (3). The liquid is decanted and the residue is transferred to a microscope slide using a micro-pipette and spread over two third of microscopic glass slides and dried. For Qualitative and quantitative melissopalynological analysis following methods which were used by Ohe et al (2004), were modified and used for the present investigation –

#### Processing of honey for the preparation of pollen slides

10 grams of honey by weight have been taken in a 50 milliliters capacity pointed Glass centrifuge tube. 20 milliliters of distilled hot water (20°-40°k) was mixed so as to dissolve the honey. This solution was centrifuged for 10 minutes at 1000 rpm. The supernatant liquid was decanted. Again 20 milliliters of distilled water was added to the residue so as to dissolve the remaining crystals. Again this mixture was centrifuged for 5 minutes at 1000 rpm. The supernatant liquid was decanted and removed by placing the tube upside down at 45° angle. The excess liquid is allowed to be taken up by an absorbent paper.

The glycerin jelly (mounting medium) was liquified by heating it at 40° C on a heating plate. The glycerin jelly which has been taken for the mounting of pollen grains was prepared by adding some drops of 0.1%(w/v) sapanin ethanol solution (0.5-1 ml of this solution was added to 10 ml of fluid glycerin jelly). A water proof marker was used to draw a 22 x 22 mm square on the microscope slide and the slide put on the heating plate. The entire residue was mixed thoroughly with a Pasteur pipette and the entire residue was transferred on the slide. The residue was spread evenly with a thin glass rod over the marked 22 x 22 mm area. Now the slide is left on the heating plate only for the time strictly necessary to dry the residue. Some of the cover – slips (22 x 22 mm), Were warm up on the heating plate. One drop of glycerin jelly was taken and applied it on to the cover slip to form a large cross diagonally. Now the cover slip was placed on the slide very slowly to avoid air bubbles. The preparation was left on the heating plate for 5 minutes so that an even dispersion of glycerin jelly and uniform swelling of pollen grains take place. It should be kept in mind as precautionary measure that the drop of glycerin jelly should never be applied directly on the dried residue of pollen grains. During the whole procedure, great care has been taken to prevent contamination from foreign pollen, coming from either previous honey preparations or from air borne pollen grains.

# Identification and counting of the pollen grains qualitative analusis

The examination under the microscope has been carried out at the magnification 400 x 1000x. First of all the types and density of pollen grains were determined and then relative frequencies of each pollen type are determined as follows –

Pollen grains were identified and counted in groups of 100 from 5 parallel equidistant lines uniformly distributed from one edge of the cover slip to the other. Abortive, irregular or broken pollen grains were also counted. Non identifiable or non-identified pollen grains were noted separately. Further, the fungal spores, hyphae and other microscopic elements were also noted down separately.

#### Calculation and reporting the result

As far as possible the pollen grains were identified upto family, genus and species level. For each type of pollen grain, relative frequency with respect to the total number of pollen grains was calculated as the respective percentage. For the determination of botanical origin of honey, the relative frequency of the pollens were recalculated by excluding the number of pollens of nectarless plants.

#### Interpretation of the results

For the identification of pollen types and the interpretation of pollen spectra a collection of reference pollen slides and photographic atlas were used as suggested by Maurizio and Louveaux (1965), Sawyer (1988), Ricciardelli d' Albore (1997,1998).

#### Determination of Botanical origin

The determination of the botanical origin of the honey has been categorized as unifloral (if the relative frequency of the pollen of the taxon exceeds 45%), otherwise it has been kept under the multifloral category. As ar as possible, pollens were identified upto spacing level but a few are left which could not be identified upto species level.

#### Methods of Quantitative Melissopalynological Analysis

The slides were prepared by using the same methodology as used for qualitative analysis.

In order to examine the surface uniformly, ten equidistant parallel lines in the field of objective were observed from an edge of the cover slip to the other.

#### Calculation, Expression And Interpretation of Results

The absolute number of pollen-grains in 10 gram of honey (PG/10 g) were calculated as follows –

#### PG / 10 g = S x npg x 10

sxaxp

Where 'S' is the surface area (mm<sup>2</sup>) of a part of the glass slide containing the sediments residue. 's' the area of one microscopic field at the magnification used (mm<sup>2</sup>), npg is the total number of pollen grains (PG) counted.

#### Acetolysis Method For The Preparation Of Reference Slides

This method has been given by Erdtman (1960) and used for the preparation of reference slides of the identified taxa of the region for comparison. This method comprises with the following steps –

- (i). Put the pollen material into a heat resistant centrifuge tube and cover with 5 ml mixture of acetic anhydride and sulfuric acid (this mixture can be prepared by adding the acid, drop by drop, to nine times the volume of acetic anhydride).
- (ii). Insert a glass rod to each centrifuge tube and transfer the tubes to a water bath at 70° C if possible in a fuming chamber).
- (iii). Heat the water bath to boiling point, then immediately stop heating and stir the liquid in the tubes and transfer these to the centrifuge.
- (iv). Centrifuge the mixture at 2500 rpm for 5 minutes, decant the reaction mixture into a reserve receptacle.
- (v). Add 10 ml of water alcohol mixture to the residue and shake the tube thoroughly.

After acetolysis and washing transfer about one third of the suspension from the centrifuge tube to another tube. Centrifuge and decant again, then add to the residue about 2 ml of glacial acetic acid (GAA), 1 or 2 drops of saturated sodium chlorate solution and finally 2

or 3 drops of concentrated hydrochloric acid. Stir the liquid with glass rod. By this reaction chlorine is produced immediately, and bleaching is usually obtained in a few seconds.

Again centrifuge the mixture, decant and wash it twice with distilled water. Then mix the suspensions of acetolysed pollen grains and of acetolysed and chlorinated pollen grains. After centrifuging and decanting once again suspend the residue in a few drops of a mixture of glycerin and water (1:1). Leave for at least 10 minutes, centrifuge, decant, then invest the centrifuge tubes on glass slides. Fix a drop of glycerin jelly on a needle, and carefully dip it into the pollen bearing sediment. Then transfer to a slide the jelly and the pollen material adhering to it. Cover with a carefully cleaned, very thin circular cover—glass (diameter, about 10 mm) or a square or rectangular cover—glass.

The margin of the cover slip should be sealed with melted paraffin. Then turn the slide up side down to allow small pollen fragments to settle close to the cover glass.

#### Stability of Preparations of Non-Acetolysed Pollen Grains

Pollen grains in reference preparations alter in the course of time. If the fatty oil is not removed from them, they become pale and the exine also losses colour. In all types of preparation the pollen grains increase in size, because of swelling. Old preparations are also useful as they show particular characteristics better than fresh ones, but they should not be used for comparing diameters.

WINTER SAMPLES

Kubza Village

Mokokehung district reveal the presence of 1 lb and grains belonging to various members of respect to be collected by the bees of April Indian The market

OBSERVATIONS

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Myrtus communed )

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Matricaria sp. Michenia micrantha, Screen

Nanthium strumarum, Ebenaceae - Diocygron
eteclsior, Boraginaceae - Cynoglessum

Convolvalaceos - Convolvalas, arvensis, Polemante

### OBSERVATIONS

#### WINTER SAMPLES

#### Kubza Village

Winter samples of honey obtained from Kubza Village of Mokokchung district reveal the presence of following types of pollen grains belonging to various members of respective families in the honey collected by the bees of *Apis indica*. The number of pollen grains is given per ml of honey sample.

Ranunculaceae - Helleborous niger, Nigella, Clematis vitalba, Magnoliaceae - Magnolia grandiflora, Brassicaceae - Brassica napus, Violaceae - Viola tricolor, Caryophyllaceae - Stellaria media, Spergularia rupicola, Cistaceae - Helianthemum chamacistis, Malvaceae -Hibiscus rosa sinensis, Abutilon indicum, Convovulaceae - Convolvulus cantribrica, Geraniaceae - Geranium rotundifolium, Balsaminaceae -Impatiens glandulifera, Rutaceae - Citrus limon, Vitaceae - Parthenocissus tricuspidata, Vitis vinifera, Hippocastanaceae - Aesculus hippocastanum, Anarcardiaceae - Pistacia lentiscus, Platanaceae - Platanus orientalis, Fabaceae - Erythrina indica, Desmodium sp, Lupinus albus, Melilotus alba, Ononis pubescens, Ulex europacus, Ulex galli, Caesalpinaceae - Cassia didimobotrya, Mimosoideae - Acacia dealbata, Rosaceae - Prunus domestica, Prunus dulcis, Pyrus communis, Crassulaceae - Sedum acre, Myrtaceae -Myrtus communis, Eucalyptus gunii, Passifloraceae - Passiflora, edulis, Cucurbitaceae - Ecballium elaterium, Citrullus lanatus, Apiaceae -Bupleurum fruticosum, Araliaceae - Hedera helix, Cornaceae - Cornus sanguinea, Asteraceae - Ageratum conizoides, Arctium sp, Carduus sp, sp, Cirsium sp, Eupatorium japonica, Helianthus annuus, Carthamus Matricaria sp, Michenia micrantha, Senecia sp, Solidago canadensis, Xanthium strumarium, Ebenaceae - Diospyros kaki, Oleaceae - Fraxinus excelsior, Boraginaceae - Cynoglossum creticum, Symphytum, Convolvulaceae - Convolvulus arvensis, Poleminiaceae - Phlox drummondii, Scrophulariaceae - Antirrhinum majus, Linaria vulgaris, Verbascum

thapsus, Verbenaceae - Verbena officinalis, Lamiaceae - Thymus sp,
Lavandula angustifolia, Rosmarinus sp, Teucrium sp, Plantaginaceae Plantago lanceolata, Polygonaceae - Fagopyrum esculentum, Lauraceae Laurus sp, Thymelaceae - Daphne gnidium, Elaeagnaceae - Hippophae
rhamnoide, Loranthaceae - Loranthus sp, Euphorbiaceae - Chrozophora sp,
Urticaceae - Urtica dioica, Ulmaceae - Ulmus procera, Betulaceae - Alnus
glutinosa, Fagaceae - Querus robur, Musaceae - Musa paradisiaca, Iridaceae
- Iris unguicularis, Liliaceae - Allium cepa , Asparagus acutifolius,
Smilaceae - Smilax aspera, Poaceae - Alopecurus pratensis, Pinaceae Pinus insularis, Cupressaceae - Cupressus sempervirens.

Plate - 1

Figure - 1

Genus - Helleborous

Species - niger

Shape – Irregularly round

Size  $-35\mu$ Pore / colpi -Furrows

Number of pollen grains - 1

Genus – Nigella

Species -

Shape – Oblate spheroidal

 $\begin{array}{ccc} Size & & - & 37.6 \ \mu \\ Pore \ / \ colpi & & - \ Tricolpate \end{array}$ 

Number of pollen grains - 15

Plate -1

Figure - 2

Genus - Clematis

Species - vitalba

Shape – Suboblate Size – 20 μ

Pore / colpi – Tricolpate

Plate - 1

Figure - 4

Genus

- Magnolia

Species - grandiflora

Shape

- Oblate

Size

 $-51.5 \mu$ 

Pore / colpi

- Monocolporate

Number of pollen grains - 90

Plate - 2

Figure - 6

Genus

- Brassica

Species - napus

Shape

Oblate Spheroidal

Size

 $-24.4 \mu$ 

Pore / colpi

- Tricolporate

Number of pollen grains - 63

Plate - 2

Figure - 8

Genus

- Viola

Species – tricolor

Shape

Multisided or Irrigular

Size

 $-80.4 \mu$ 

Pore / colpi

- Furrows with bores

Number of pollen grains - 579

Plate - 3

Figure - 10

Genus

- Spergularia

Species - rupicola

Shape

- Oval Flattened

Size

 $-23.7 \mu$ 

Pore / colpi

- Furrows only

Plate - 3

Figure - 11

Genus - Stellaria

Species - media

Size  $-24.4 \mu$ 

- Spheric

Pore / colpi

Shape

Periporate

Number of pollen grains - 9

Plate - 4

Figure - 13

Genus - Helianthemum

Species - chamaecistis

Shape

- Oblate spheroidal – 45.4 μ

Size

Pore / colpi – Tricolporate

Number of pollen grains - 1

Plate - 4

Figure - 14

Genus - Hibiscus

Species – rosa - sinensis

Shape Size

- Spheric - 147.5 μ

Pore / colpi - Tricolporate

Number of pollen grains - 17

Genus - Abutilon

Species - indicum

Shape Size

- Suboblate

Pore / colpi

- 58.3 - Tricolporate

- Convolvulus Genus

- Oblate spheroid

Shape

Size Pore / colpi

- 53.6 µ - Tricolporate

Number of pollen grains - 4

Plate - 6

Figure - 17

Genus

- Geranium

Species - rotundifolium

Species - arvensis

- Oblate spheroid Shape - 47.2 μ

Size

Pore / colpi - Tricolporate

Number of pollen grains - 11

Plate - 7

Figure - 20

Genus

- Citrus

Species - limon

Shape

- Oblate spheroidal - 40.2 µ

Size

Pore / colpi

- Tetracolporate

Number of pollen grains - 23

Plate - 7

Figure - 19

Genus

- Impatiens

Species – glandulifera

- Long Shape Size - 47.2 µ

Pore / colpi

- Furrows only

Plate - 7

Figure - 22

Genus

- Vitis

Species - vinifera

Shape

- Oblate spheroidal

Size

- 23.1 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 16

Plate - 7

Figure - 21

Genus

-Parthenocissus

Species – tricuspidata

Shape

- Prolate spheroidal

Size

- 38.3 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 8

Plate - 8

Figure - 23

Genus – Aesculus

Species - hippocastanum

Shape - Prolate spheroidal Size

- 18.5 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 2

Plate - 8

Figure - 25

Genus

- Pistacia

Species – lentiscus

Shape

Size - 28.3 μ

Pore / colpi

- Stephanoporate

- Oblate spheroidal

Figure - 26

Genus

- Platanus

Species - orientalis

Shape

- Subprolate

Size

- 24.2 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 3

Genus - Desmodium

Species -

Shape

Size

Pore / colpi

Number of pollen grains -

Genus

- Erythrina

Species - indica

Shape

Size

Pore / colpi

Number of pollen grains -

Plate - 11

Figure - 31

Genus

- Lupinus

Species - albus

Shape

- Prolate spheroidal

Size

- 29.5 µ

Pore / colpi

- Tricolporate/tricolporoidate

Figure - 33

Genus

- Melilotus

Species – alba

Shape

- Prolate spheroidal

Size

- 23.7 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 6

Plate - 12

Figure - 35

Genus

- Ononis

Species - pubescens

Shape - Prolate Size - 38.6 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 4

Genus - Ulex Species - europacus

Shape

- Oval flattened

Size

- 43.5 µ

Pore / colpi

- Furrows

Number of pollen grains - 6

Plate - 13

Figure - 38

Genus

- Ulex

Species – galli

Shape

Size

- Oval flattened

- 43.5 µ

Pore / colpi

Figure - 40

Genus

- Cassia

Species - didimobotrye

Shape

- Oblate spheroidal

Size

- 34.2 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 10

Plate - 14

Figure - 41

Genus

- Acacia

species- dealbata

Shape

Size

- 54.1 µ

Pore/colpi

- Inaperturarte

Number of pollengrains- 31

Plate - 15

Figure - 43 - 44

Genus

- Malus

Species – domestica

Shape

- Suboblate

Size

- 27.2 µ

Pore / colpi

- Tricolporoidate

Number of pollen grains - 7

Plate - 16

Figure -45

Genus

- Prunus

Species - dulcis

Shape

Size

- Suboblate  $-43.4 \mu$ 

Pore/Colpi

- Tricolporoidate

Figure - 46

Genus

- Pyrus

Species – cummunis

Shape

- Triangular

Size

 $-46.7 \, \mu$ 

Pore/Colpi

- Furrows with pores

Number of pollen grains - 80

Plate - 17

Figure - 47

Genus

- Sedum

Species - acre

Shape Size

- Suboblate  $-24.4 \mu$ 

Pore / colpi

- Tricolporoidate

Number of pollen grains - 2

Plate - 17

Figure - 48

Genus

- Eucalyptus

Species - gunii

Shape

- Oblate - 13.5 µ

Size Pore / colpi

- Tricolporate/Syncolpate

Number of pollen grains - 39

Plate - 17

Figure - 49

Genus

- Myrtus

Species - communis

Shape

- Oblate

Size

 $-10.7 \mu$ 

Pore / colpi

- Tricolporate/Syncolpate

Figure - 50

Genus

- Passiflora

Species - edulis

Shape

- Spheric

Size

 $-64.8 \mu$ 

Pore / colpi

- Spiraperture syncolpate

Number of pollen grains - 5

Plate - 18

Figure - 52

Genus

- Citrullus

Species - lanatus

Shape Size

- Oblate spheroidal - 52.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 15

Plate - 19

Figure - 53

Genus

- Ecballium

Species – elaterium

Shape Size

- Oblate spheroidal - 46.9 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 14

Plate - 19

Figure - 55

Genus

- Bupleurum

Species - fruticosum

Shape

Size

- Prolate - 18.4 µ

Pore / colpi

- Tricolporate

Figure - 57

Genus

- Hedera

Species - helix

Shape

- Oblate spheroidal

Size

- 32.3 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 6

Plate - 20

Figure - 58

Genus

- Cornus

Species – sanguinea

Shape

- Prolate spheroidal

Size

- 54.3 μ

Pore / colpi

- Tricolporate

Number of pollen grains - 45

Plate - 20

Figure - 59

Genus

- Ageratum

Species - conizoides

Shape

Size

Pore / colpi

Number of pollen grains - 340

Plate - 20

Figure - 60

Genus

- Arctium

Species -

Shape

Prolate

Size

 $-45.7 \mu$ 

Pore / colpi

- Tricolporate

Figure - 62

Genus

- Carduus

Species -

Shape

- Prolate spheroidal

Size

 $-54.3 \mu$ 

Pore / colpi

- Tricolporate

Number of pollen grains - 100

Plate - 21

Figure - 63

Genus

Shape

- Carthamus

Species -

Size

 Subprolate  $-60.6 \,\mu$ 

Pore / colpi

- Tricolporate

Number of pollen grains - 7

Plate - 21

Figure - 64

Genus

- Cirsium

Species -

Shape

- Oblate spheroidal

Size

- 42.1 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 24

Genus

- Eupatorium

Species - japonica

Shape

- Oblate spheroidal

Size

- 22.5 μ

Pore / colpi - Tricolporate

Figure - 65

Genus

- Helianthus

Species - Annuus

Shape

- Oblate spheroidal

Size

 $-38.4 \mu$ 

Pore / colpi

- Tricolporate

Number of pollen grains - 599

Plate - 22

Figure - 67

Genus – Matricaria

Species -

Shape

Oblate spheroidal

Size

 $-23.6 \mu$ 

Pore / colpi — Tricolporate

Number of pollen grains - 17

Plate - 22

Figure - 68

Genus - Michenia

Species - micrantha

Shape

Size

Pore / colpi

Number of pollen grains - 517

Genus

- Senecio

Species -

Shape

- 28.2 u

Size

Pore / colpi - Tricolporate

- Oblate spheroidal

Figure - 69

Genus

- Solidago

Species - canadensis

Shape

Oblate spheroidal

Size

- 22.4 µ

Pore / colpi

- Tricolporate

Spherical

- 26.8 µ

Number of pollen grains - 51

Plate - 23

Figure - 71

Genus

- Xanthium

Species - strumarium

Shape Size

- Tricolporate Pore / colpi

Number of pollen grains - 57

Plate - 24

Figure - 74

Genus – Diospyros

Species - kaki

Shape

- Prolate spheroidal

Size

- 45.2 µ

Pore / colpi

- Tricolpate

Number of pollen grains - 2

Plate - 24

Figure - 75

Genus

- Fraxinus

Species – excelsior

Shape

- Oblate spheroidal

Size

- 19.5 µ

Pore / colpi

- Tricolporoidate

Figure - 77

Genus

- Cynoglossum

Species - creticum

Shape Size

- Prolate - 18.5 µ

Pore / colpi

- Stephanocolporate

Number of pollen grains - 13

Plate -25

Figure - 79

Genus

- Symphylum

Species - officinale

Shape Size

- Prolate - 27.3 µ

Pore / colpi

- Stephonocolporate

Number of pollen grains - 4

Plate - 25

Figure - 80

Genus - Convolvulus

Species - arvensis

Shape

- Oblate spheroidal

Size

- 53.6 µ - Tricolpate

Pore / colpi Number of pollen grains - 4

-PhloxGenus

Species – drummondii

Shape

- Irregularly round

Size Pore / colpi - Pores only

- 57.6 µ

Figure - 81

Genus

- Antirrhinum

Species - majus

Shape

- Oblate spheroidal

Size

- 12.5 µ

Pore / colpi

- Tricolporate/Syncolporate

Number of pollen grains - 16

Genus

- Linaria

Species - vulgaris

Shape

- Irregularly round

Size

- 14.2 µ

Pore / colpi

- Furrows with pores

Number of pollen grains - 3

Plate - 26

Figure - 82

Genus

- Verbascum

Species - thapsus

Shape

- Oblate spheroidal

Size

- 23.5 µ

Pore / colpi

- Tricolpate

Number of pollen grains - 10

Plate - 27

Figure - 83

Genus

- Verbena

Species - officinalis

Shape

- Oblate spheroidal

Size

- 25.4 µ

Pore / colpi

- Heterocolpate

Figure - 84

Genus

- Lavandula

Species - angustifolia

Shape

- Oblate

Size - 25.5

Pore / colpi

- Stephanocolpate

Number of pollen grains - 2

Plate - 29

Figure - 87 - 88

Genus

- Rosmarinus

Species -

Shape Size - Suboblate

- 33.2 µ Pore / colpi - Stephanocolpate

Number of pollen grains - 1

Plate - 29

Figure - 89

Genus

- Teucrium

Species -

Shape

- Subprolate

Size

- 52.2 µ

Pore / colpi

- Tricolpate

Number of pollen grains - 1

Plate - 29

Figure - 90

Genus

- Thymus

Species -

Shape

Size

- 30.4 µ

Pore / colpi

- Stephanocolpate

- Oblate spheroidal

Genus - Plantago

Number of pollen grains - 1

Species - lanceolata

Plate - 30

Genus - Fagopyrum

Shape - Subprolate

Size - 40.4

Pore / colpi - Tricolporate

Number of pollen grains - 37

Figure - 92

Species – esculentum

Plate - 31

Genus - Laurus

 $\begin{array}{ccc} Shape & & - Spheric \\ Size & & - 55.4\,\mu \end{array}$ 

Pore / colpi - Inaperturate

Number of pollen grains - 3

Figure - 93 - 94

Species -

Plate -32

Genus - Daphne

Shape - Spheric
Size - 21.3 μ
Pore / colpi - Periporate

Number of pollen grains - 2

Figure - 95

Species - gnidium

- Hippophae Genus

- Irregularly round

Shape Size

- 23.4 µ

Pore / colpi

- Furrows with pores

Number of pollen grains - 1

Plate -32

Figure - 97

Species - rhamnoides

Genus

- Loranthus

Species -

Shape Size

 Oblate - 15.4 µ

Pore / colpi

- Tricolporate/Syncolpate

Number of pollen grains - 3

Plate - 32

Figure - 98

Genus

- Chrozophora

Species -

Shape

 Suboblate - 54.6 µ

Size Pore / colpi

- Stephanocolporate

Number of pollen grains - 11

Plate - 33

Figure - 99

Genus

- Urtica

Species - dioica

Shape

- Irregularly round

Size

- 13.8 µ

Pore / colpi - pores only

Figure - 100

Genus

- Ulmus

Species - procera

Shape

- Suboblate

Size

- 25.5 µ

Pore / colpi

- Stephanocolporate

Number of pollen grains - 16

Plate - 33

Figure - 101

Genus

- Alnus

Species - glutinosa

Shape

- Irregular oval

Size

- 24.5 µ

Pore / colpi

- pores only

Number of pollen grains - 5

Plate - 34

Figure - 104

Genus

- Quercus

Species - robur

Shape

- Oblate spheroidal

Size

- 28.4 µ

Pore / colpi

- Tricolporoidate

Number of pollen grains - 43

Plate - 34

Figure - 106

Genus

- Musa

Species - paradisiaca

Shape

Size

- Circular - 120.6 µ

Pore / colpi - Inaperturate

Figure - 108

Genus Shape - Iris

Species - unguicularis

Size

– Round - 78.4 μ

Pore / colpi

- Furrows only

Number of pollen grains - 3

Plate - 35

Figure - 109

Genus

- Allium

Species - cepa

Shape – Oblate Size – 18.5 μ

Pore / colpi

- Monocolpate

Number of pollen grains - 32

Genus – Asparagus

Species - acutifolius

Shape Size Oblate

Pore / colpi

- 16.4 μ - Monocolpate

Number of pollen grains - 23

Genus - Smilax

Species – aspera

Shape Size

– Spheric - 20.5 μ

Pore / colpi

- Inaperturate

Figure - 111

Genus - Alopecurus

Species - pratensis

Shape Size Round
 47.5 μ

Pore / colpi

- Pores only

Number of pollen grains - 2

Plate - 38

Figure - 114 - 116

Genus

- Pinus

Species - msularis

Shape – Multisided or irregular Size - 86.3 µ

Daniel Carlo

Pore / colpi

Number of pollen grains - 3

Plate - 39

Figure - 117

Genus

- Cupressus

Species - sempervirens

Shape - Spheroid Size - 24.1 μ

Pore / colpi - Monoporate

Number of pollen grains - 5

## Ungma Village

Winter samples of honey obtained from Ungma Village of Mokokchung district reveal the presence of following types of pollen grains belonging to various members of respective families in the honey collected by the bees of Apis indica. The number of pollen grains is given per ml of honey sample.

Ranunculaceae - Helleborus niger, Nigella sp, Clematis vitalba, Magnoliaceae - Liriondendron tulipifera, Magnolia grandiflora, Berberidaceae - Berberis darwinii, Papaveraceae - Hypecoum procumben, Brassicaceae - Brassica napus, B. oleifera Cardamine pratensis, Violaceae - Viola tricolor, Caryophyllaceae - Lychnis floscuculi, Tamaricaceae - Tamarix gallica, Malvaceae - Hibiscus rosa - sinesis, Malva sylvestris, Tiliaceae - Tilia peteolaris, Geraniaceae - Geranium pretense, G. rotundifolium, Oxalidaceae - Oxalis articulata, Rutaceae - Citrus limon, Vitaceae - Parthenocissus tricuspidata, Vitis vinifera, Aceraceae - Acer pseudoplatanus, Anacardiaceae - Pistacia lentiscus, Platanaceae - Platanus orientalis, Fabaceae - Calycotome spinosa, Ceratonia siliqua, Coronilla emerus, Colutea arborescens, Desmodium sp, Erythrina indica, Gleditsia triacanthos, Lupinus albus, Medicago sativa, Ononis pubescens, O. spinosa, Onobrychis vicifolia, Sophora japonica, Trifolium alexandrium, Vicia faba, Caesalpiniaceae - Cassia didimobotrya, Mimosoideae - Acacia dealbata, Rosaceae - Crataegus monogyna, Filipendula ulmaria, Malus domestica, Prunus dulcis, Pyrus Communis, Crassulaceae - Sedum acre, Myrtaceae Eucalyptus gunnii, Myrtus communis, Passifloraceae - Passiflora edulis, Cucurbitaceae - Bryonia dioica, Citrullus lanatus, Cucurbita pepo, Apiaceae Bupleurum fruticosum, Araliaceae - Hedera helix, Cornaceae - Cornus sanguinea, Asteraceae - Ageratum conizoides, Aster sp, Carduus sp, Carthamus sp, Eupatorium japonica, Helianthus annuus, Inula sp, Solidago canadensis, Taraxacum officinale, Xanthium strumarium, Ericaceae -Arbutus unedo, Erica sp, Plumbaginaceae - Armeria maritima, Ebenaceae -Diospyrus kaki, Oleaceae - Fraxinus excelsior, Buddlejaceae Buddleja officinalis, Symphytum officinate, Convolvulaceae Convolvulus arvensis, Polemoniaceae - Phlox drummondii, Scrophulariaceae - Linaria vulgaris, Verbascum thapsus, Verbenaceae - Verbena officinalis, Lamiaceae -Lavandula stoechas, Mentha pulegium, Teucrium sp, Thymus sp, Plantaginaceae - Plantago lanceolata, Chenopodiaceae - Chenopodium, Polygonaceae - Fagopyrum esculentum, Lauraceae - Laurus nobilis,

Loranthaceae - Loranthus, Euphorbiaceae - Chrozophora, Urticaceae - Urtica dioica, Ulmaceae - Ulmus procera, Betulaceae - Alnus glutinoso, Betula pendula, Castanea sativa, Fagaceae - Quercus robur, Salicaceae - Populus tricocarpa, Salix caprea, Liliaceae - Asparagus acutifolius, Asphodelus microcarpus, Allium cepa, Poaceae - Zea mays, Pinaceae - Pinus insularis.

Plate - 1

Figure -1

Genus

- Helleborus

Species - niger

Shape Size

Pore / colpi

- 35 μ - Furrows

- Irregularly round

Number of pollen grains - 7

Genus

- Nigella

Species -

Shape

Oblate spheroidal

Size

- 37.6 μ

Pore / colpi

- Tricolpate

Number of pollen grains - 10

Plate - 1

Figure - 2

Genus

- Clematis

Species -vitalba

Shape

Size

Pore / colpi

Figure - 3

Genus

- Liriodendron

Species – tulipifera

Shape

- Suboblate

Size

 $-51.6 \mu$ 

Pore / colpi

- Monocolpate

Number of pollen grains - 2

Plate - 1

Figure - 4

Genus

- Magnolia

Species – grandiflora

Shape Size - Oblate  $-51.5 \mu$ 

Pore / colpi

- Monocolpate

Number of pollen grains - 26

Plate -

Figure -

Genus

- Berberis

Species – darwinii

Shape

- Irregularly round

Size

 $-40.1 \mu$ 

Pore / colpi

- United or irregular furriws may occur

Number of pollen grains - 70

Plate - 2

Figure - 5

Genus

- Hypecoum

Species – procumben

Shape

- Prolate spheroidal

Size

 $-24.5 \mu$ 

Pore / colpi - Dicolbate

Figure - 6

Genus

- Brassica

Species - napus

Shape

Oblate spheroidal

Size

 $-24.7~\mu$ 

Pore / colpi

- Tricolpate

Number of pollen grains - 58

Genus

- Brassica

Species - oleifera

Shape

- Oblate spheroidal

Size

 $-24.7 \mu$ 

Pore / colpi

- Tricolpate

Number of pollen grains - 68

Plate - 2

Figure - 7

Genus

- Cardamine

Species – pratensis

Shape

Irregularly round

Size

-24.6

Pore / colpi

- Furrows only

Number of pollen grains - 50

Plate - 2

Figure - 8

Genus

- Viola

Species -tricolor

Shape

Multisided or irregular

Size

 $-80.4 \mu$ 

Pore / colpi

Furrows with pores

Figure – 9

Genus

- Lychnis

Species - flos -cuculis

Shape Size

- Spheric

Pore / colpi

- 45.5 µ - Periporate

Number of pollen grains - 4

Plate - 3

Figure - 12

Genus Shape

- Tamarix

Species – gallica

Size

- Oblate spheroidal - 18.8 µ

Pore / colpi

- Tricolpate

Number of pollen grains - 5

Plate - 4

Figure - 14

Genus

- Hibiscus

Species – rosa-sinesis

Shape Size

- Spheric  $-147.5 \,\mu$ 

Pore / colpi

- Periporate

Number of pollen grains - 2

Genus

-Malva

Species – sylvestris

Shape

- Spheric

Size

 $-105.4 \mu$ 

Pore / colpi

- Periporate

Figure - 15

Genus

- Tilia

Species – petiolaris

Shape

- Triagular/irregular

Size

 $-35.6 \mu$ 

Pore / colpi

- Furrows with pores

Number of pollen grains - 23

Plate - 5

Figure - 16

Genus

- Geranium

Species - pratense

Shape - Oblate spheroidal Size -47.2

Pore / colpi

- Tricolporate

Number of pollen grains – 10

Plate - 6

Figure - 17

Species – rotundifolium

Genus

- Geranium

- Oblate spheroidal

Shape

 $-45.3 \mu$ 

Size Pore / colpi

- Tricolporate

Number of pollen grains - 11

Plate - 6

Figure - 18

Genus

- Oxalis

Species – articulata

Shape

 $-32.5 \mu$ Size

Pore / colpi

- Tricolporate

- Oblate spheroidal

Figure - 20

Genus

- Citrus

Species – limon

Shape

- Oblate spheroidal

Size

 $-40.2 \mu$ 

Pore / colpi

- Tetracolporate

Number of pollen grains - 5

Plate - 7

Figure - 21

Genus

- Parthenocissus

Species – tricuspidata

Shape

- Prolate spheroidal

Size

 $-38.3~\mu$ 

Pore / colpi

- Tricolporate

Number of pollen grains - 95

Plate - 7

Figure - 22

Genus

- Vitis

Species - vinifera

Shape

Size

-38.3

Pore / colpi

- Tricolporate

- Oblate spheroidal

Number of pollen grains – 84

Plate - 8

Figure - 24

Genus – Acer

Species - pseudoplatanus

Shape - Oval flattened

Size

- 43.8 μ

Pore / colpi

- Furrows only

Figure - 25

Genus

- Pistacia

Species – lentiscus

Shape

- Oblate spheroidal

Size

- 28.3 µ

Pore / colpi

- Stephanoporate

Number of pollen grains - 3

Plate - 8

Figure - 26

Genus

- Platanus

Species - orientalis

Shape Size - Subprolate - 24.2 μ

Pore / colpi

- Tricolpate

Number of pollen grains - 25

Plate - 10

Figure - 28

Genus

- Calycotome

Species -spinosa

Shape

- oblate spheroidal

Size Pore / colpi - 25.2 μ - Tricolpate

Number of pollen grains - 22

Genus

- Ceratonia

Species - siliqua

Shape

- Oblate spheroidal

Size

- 23.5

Pore / colpi

- Tetracolporate

Figure -27

Genus

- Coronilla

Species – emerus

Shape

Size

- Oblate spheroidal - 27.2 µ

Pore / colpi

- Tricolporoidate

Number of pollen grains - 26

Genus

- Colutea

Species – arborescens

Shape

- Oblate spheroidal

Size

- 35.2 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 70

Genus – Desmodium

Species -

Shape

Size

Pore / colpi

Number of pollen grains -

Genus -ErythrinaShape

Size

Pore / colpi

Number of pollen grains -

Species – indica

Figure - 29

Genus

- Gleditsia

Species – triacanthos

Shape - Oblate spheroidal Size - 32.4 μ

Pore / colpi

- Tricolporate

Number of pollen grains - 27

Plate - 11

Figure - 31

Genus

- Lupinus

Species – albus

Shape

- Prolate spheroidal

Size

- 29.5 µ

Pore/colpi

Number of pollen grains - 32

Plate - 11

Figure - 32

Genus

- Medicago

Species – sativa

Shape

- Oblate spheroidal

Size - 32.9 μ

Pore / colpi - Tricolporoidate

Number of pollen grains - 9

Plate - 12

Figure - 35

Genus

Species – pubescens

Shape

- Ononis

Size

- Prolate - 38.6 µ

Pore / colpi - Tricolporate

Genus - Ononis

Shape - Prolate Size - 22.3 µ

Pore / colpi - Tricolporate

Number of pollen grains - 42

Plate - 12

Figure - 34

Species – vicifolia

Species -spinosa

Genus – Onobrychis

Shape - Prolate Size - 27.9 µ - Tricolpate Pore / colpi

Number of pollen grains - 29

Plate - 13

Figure - 37

Genus - Sophora

Species - japonica

Shape Size

- Prolate spheroidal - 17.2 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 143

Genus - Trifolium

Species - alexandrinum

Shape

- Prolate spheroidal

Size

- 42.4 μ

Pore / colpi

- Tricolporate

Figure - 39

Genus

- Vicia

Species – faba

Shape Size

- Prolate - 51.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 2

Plate - 13

Figure - 40

Genus

- Cassia

Species - didimobotrya

Shape - Oblate spheroidal Size - 34.2 µ

- Tricolporate

Pore / colpi

Number of pollen grains - 39

Plate - 14

Figure - 41

Genus

- Acacia

Species - dealbata

Shape

Size

- 54.1 µ

Pore / colpi

- Inaperture

Number of pollen grains -

Plate - 14

Figure - 42

Genus

- Crataegus

Species - monogyna

Shape

- Oblate spheroidal

Size - 37.5 µ

Pore / colpi

- Tricolporoidate

Figure - 43 - 44

Genus

- Malus

Species – domestica

Shape Size

- Suboblate - 27.2 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 90

Plate - 16

Figure - 45

Genus

- Prunus

Species – dulcis

Shape - Subolblate Size - 43.4 µ

Pore / colpi

- Tricolporoidate

Number of pollen grains - 81

Plate - 16

Figure - 46

Genus

- Pyrus

Species – *cummunis* 

Shape

- Triangular - 46.7 µ

Size Pore/Colpi

- Furrows with pores

Number of pollen grains - 118

Plate - 17

Figure - 47

Genus

- Sedum

Species -acre

Shape

- Oblate spheroidal Size

- 24.5 µ

Pore / colpi - Tricolporate

Figure - 48

Genus

- Eucalyptus

Species – gunii

Shape

- Oblate

Size Pore / colpi - 13.5 μ - Tricolporate/syncolpate

Number of pollen grains - 198

Plate - 17

Figure - 49

Genus

- Mrytus

Species – communis

Shape Size - Oblate - 10.7 μ

Pore / colpi

- Tricolporate/syncolpate

Number of pollen grains - 141

Plate - 17

Figure - 50

Genus

- Passiflora

Species – edulis

Shape Size - Spheric - 64.8 μ

Pore / colpi

- Spiraperture Syncolpate

Number of pollen grains - 10

Plate - 18

Figure -51

Genus

- Bryonia

Species -dioica

Shape

- Irregularly round

Size

- 38.4 μ

Pore / colpi

- Furrows with pores

Figure - 52

Genus

- Citrullus

Species -lanatus

Shape

- Oblate spheroidal - 52.4 μ

Pore / colpi

- Tricolporate

Number of pollen grains - 12

Plate - 19

Figure - 54

Genus

- Cucurbita

Species – pepo

Shape Size Pore / colpi

- Periporate

- Spheric

- 115.7 µ

Number of pollen grains - 1

Plate - 19

Figure - 55

Genus

- Bupleurum

Species – fruticosum

Shape

- Prolate

Size Pore / colpi - 18.4 μ - Tricolporate

Number of pollen grains - 22

Plate - 20

Figure - 57

Genus

- Hedera

Species -helix

Shape

- Oblate spheroidal

Size

- 32.3 µ

Pore / colpi

- Tricolporate

Figure - 58

Genus

- Cornus

Species – sanguinea

Shape

- Prolate spheroidal

Size

- 54.3 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 57

Plate - 20

Figure - 59

Genus - Ageratum

Species – conizoides

Shape

Size Pore / colpi

Number of pollen grains - 609

Plate - 21

Figure - 61

Genus

- Aster

Species -

Shape

- Triangular - 23.5 µ

Size Pore / colpi

- Furrows with pores

Number of pollen grains - 203

Plate - 21

Figure - 62

Genus

- Carduus

Species –

Shape

- Oblate spheroidal

Size

- 42.4 µ

Pore / colpi

- Tricolporate

Figure -63

Genus

- Carthamus

Species -

Shape

- Subprolate

Size

- 60.6 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 148

Genus - Eupatorium

Species – japonica

Shape

- Oblate spheroidal

Size

- 22.5 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 242

Plate - 22

Figure - 65

Genus - Helianthus

Species - annuus

Shape

- Oblate spheroidal

Size

- 38.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 1041

Plate - 22

Figure - 66

Genus

-Inula

Species -

Shape

- Oblate spheroidal

Size

- 31.2 µ

Pore / colpi

- Tricolporate

Figure - 69

Genus

- Solidago

Species – canadensis

Shape

- Oblate spheroidal

Size

- 22.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 88

Plate - 23

Figure - 70

Genus

- Taraxacum

Species – officinale

Shape

- Oblate spheroidal

Size

- 34.3

Pore / colpi

- Tricolporate/Tetracolporate

Number of pollen grains - 98

Plate - 23

Figure - 71

Genus - Xanthium

Species - strumarium

Shape Size

- Spherical - 26.8 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 249

Plate - 23

Figure -72

Genus

Species – unedo

Shape

- Arbutus

Size

- 52.5 µ

Pore / colpi - Tricolporate

Figure - 73

Genus

- Erica

Species -

Shape

Size - 32.4 μ

Pore / colpi

- Tricolporate

Number of pollen grains - 41

Genus - Armeria

Species – maritima

Shape

Size

- 76.8 µ

Pore / colpi

- Furrows only

Number of pollen grains -

Plate - 24

Figure - 74

Genus

Diospyrus

Species - kaki

Shape

- Prolate spheroidal

Size

- 45.2 µ

Pore / colpi

- Tricolpate

Number of pollen grains - 5

Plate - 24

Figure - 75

Genus

- Fraxinus

Species- excelsior

Shape

- 19.5 µ Size

Pore / colpi

- Oblate spheroidal

- Tricolporoidate

Genus - Buddleja

Species-davidii

Shape

Size

- 18.5 µ

Pore / colpi

- Furrows with pores

Number of pollen grains - 13

Plate - 24

Figure - 76

Genus - Alkanna

Species – tinctoria

- Prolate Shape - 12.5 µ Size

Pore / colpi

- Tricolporate

Number of pollen grains - 16

Genus

- Borago

Species – officinalis

Shape

- Oblate spheroidal - 38.5 µ

Size Pore / colpi

- Stephanocolporate

Number of pollen grains - 14

Plate - 25

Figure - 79

Genus

- Symphytum

Species - officinale

- Prolate Shape Size - 27.3 µ

Pore / colpi - Stephanocolporate

Figure - 80

Genus - Convolvulus

Species – arvensis

Shape

- Oblate spheroidal

Size

- 53.6 µ

Pore / colpi

- Tricolpate

Number of pollen grains -18

Genus

- Phlox

Species - drummondi

Shape

- Irregularly round

Size

- 57.6 µ

Pore / colpi

- Pores only

Number of pollen grains - 14

Genus

- Linaria

Species -vulgaris

Shape

- Irregularly round

Size

- 14.2 µ

Pore / colpi

- Furrows with pores

Number of pollen grains - 4

Plate - 26

Figure - 82

Genus

-Verbascum

Species - thapsas

Shape

- Oblate spheroidal Size - 23.5 µ

Pore / colpi

- Tricolporate

Figure - 83

Genus

- Verbena

Species - officinalis

Shape

- Oblate spheroidal

Size

- 25.4 µ

Pore / colpi

- Heterocolpate

Number of pollen grains - 4

Genus

Lavandula

Species – stoechas

Shape Size - Oblate - 25.5 µ

Pore / colpi - Stephanocolpate

Number of pollen grains - 2

Plate - 28

Figure - 85 - 86

Genus

- Mentha

Species – pulegium

Shape

- Oblate spheroidal

Size

- 18.5 µ

Pore / colpi - Stephanocolpate

Number of pollen grains - 22

Plate - 29

Figure - 89

Genus

- Teucrium

Species -

Shape

- Subprolate

Size

- 52.2 μ

Pore / colpi - Tricolpate

Figure - 90

Genus

- Thymus

Species -

Shape

- Oblate spheroidal

Size

- 30.4 µ

Pore / colpi

- Stephanocolpate

Number of pollen grains - 22

Genus - Plantago

Species – lanceolata

Shape Size

- Spheric - 24.3 µ

Pore / colpi

- Periporate

Number of pollen grains - 6

Plate - 30

Figure - 91

Genus

- Chenopodium

Species -

Shape Size

- Spheric - 26.5 µ

Pore / colpi

- Periporate

Number of pollen grains - 16

Plate - 30

Figure - 92

Genus

- Fagopyrum

Species – esculentum

Shape

- Subprolate

Size

- 40.4

Pore / colpi

- Tricolporate

Figure - 93 - 94

Genus

- Laurus

Species – nobilis

Shape

- Spheric - 55.4 µ

Size Pore / colpi

- Inaperturate

Number of pollen grains - 81

Plate - 32

Figure - 97

Genus - Loranthus

Species -

Shape Size

- Oblate - 15.4 µ

Pore / colpi - Tricolporate/Syncolpate

Number of pollen grains – 2

Plate - 32

Figure - 98

Genus - Chrozophora

Species -

Shape Size - Suboblate - 54.6 µ

Pore / colpi - Stephanocolporae

Number of pollen grains - 10

Plate - 33

Figure – 99

Genus

- Urtica

Species – dioica

Shape

- Irregularly round

Size

- 13.8 µ

Pore / colpi - Pores only

Figure - 100

Genus

- Ulmus

Species - procera

Shape

- Suboblate

Size

- 25.5 µ

Pore / colpi

- Stephanocolporate

Number of pollen grains - 4

Plate - 33

Figure – 101

Genus

- Alnus

Species – glutinosa

Shape

- Irregular oval

Size Pore / colpi - 24.5 µ - Pores only

Number of pollen grains - 1

Plate - 33

Figure - 102

Genus

- Betula

Species – pendula

Shape

- 26.2 µ Size

Pore / colpi

- Pores only

Number of pollen grains - 7

Plate - 34

Figure - 103

Genus

- Castanea

Species – sativa

Shape

- Prolate

- 15.7 µ Size

Pore / colpi

- Tricolporate

Figure - 104

Genus

- Quercus

Species - robur

Shape

- Oblate Spheroidal

Size

- 28.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 11

Plate - 34

Figure - 105

Genus

- Populus

Species – tricocarpa

Shape Size Pore / colpi

- 26.2 μ - Periporate

- Spheric

Number of pollen grains -

Genus

- Salix

Species - caprea

Shape Size Subprolate18.4 μ

Pore / colpi

- Tricolporate

Number of pollen grains -

Genus

- Asparagus

Species - acutifolis

Shape Size - Oblate

Pore / colpi

- 16.4 μ - Monocolpate

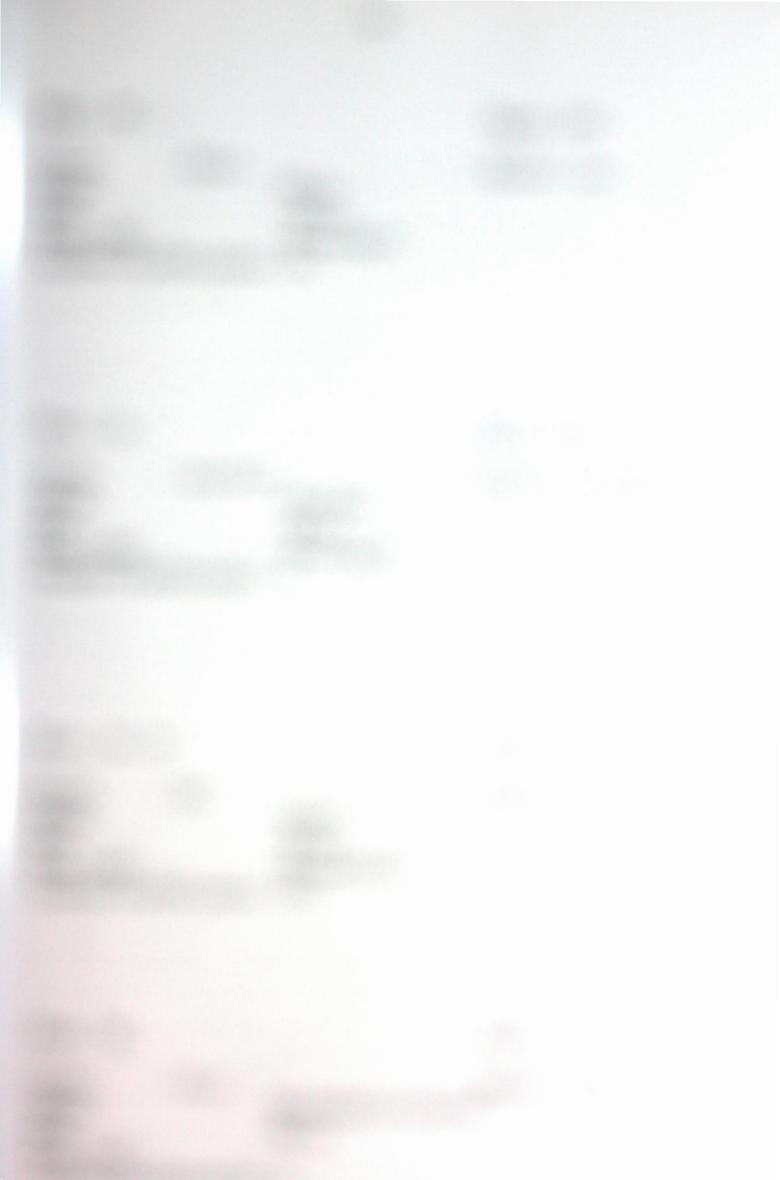


Figure - 109

Genus

- Allium

Species – cepa

Shape Size

- Oblate - 18.5 µ

Pore / colpi - Monocolpate

Number of pollen grains - 44

Plate - 35

Genus - Asphodelus

Size

- Suboblate Shape - 40.2 µ

Pore / colpi

Monocolpate

Number of pollen grains - 3

Figure - 110

Species - microcarpus

Plate - 36 - 37

Figure - 112 - 113

Species - mays

Genus – Zea Shape - Round Size - 70.4 µ

Pore / colpi

Monoulcerate

Number of pollen grains - 231

Plate - 38

Figure - 114 - 116

Genus - Pinus

Species - insularis

Shape - Multisided or irregular

Size

- 86.3 µ

Pore / colpi Number of pollen grains - 2

## Khensa Village

Winter samples of honey obtained from Khensa Village of Mokokchung district reveal the presence of following types of pollen grains belonging to various members of respective families in the honey collected by the bees of *Apis indica*. The number of pollen grains is given per ml of honey sample.

Ranunculaceae - Helleborus niger, Magnoliaceae -Magnolia grandifolia, Berberidaceae - Berberis darwinii, Brassicaceae -Brassica napus, Cardamine pratensis, Violaceae - Viola tricolor, Caryophyllaceae - Lychnis flos cuculi, Malvaceae - Hibiscus rosa - sinensis, Abutilon indicum, rosa sinensis, Tiliaceae - Tilia petiolaris, Geraniaceae pratense, G. rotundifolium, Oxalidaceae - Oxalis articulata. Balsaminaceae - Impatiens glandulifera, Rutaceae - Citrus limon, Vitaceae -Parthenocissus tricuspidata, Anacardiaceae - Pistacea lentiscus. Plantanaceae - Platanus orientalis, Fabaceae - Colutea arborescens Gleditsa triancanthos, Onobrychis vicifolia, Ononis pubescens, Sophora japonica, Ulex gali, Caesalpiniaceae - Cassia didimobotrya, Mimosoideae - Acacia dealbata, Rosaceae - Crataegus monogyna, Eriobotrya japonica, Malus domestica, Prunus dulcis, Myrtaceae - Myrtus communis, Passifloraceae -Passiflora edulis, Cucurbitaceae - Bryonia dioica, Citrullus Apiaceae - Bupleureum fruticosm, Smyrnium olusatrum, Araliaceae - Hedera helix, Cornaceae - Cornus sanguinea, Asteraceae - Ageratum conizoides, Aster sp, Carduus sp, Carthamus sp, Cirsium, Eupatorium japonica, Helianthus annuus, Inula sp, Matricaria sp, Michenia micratha, Senecio sp, Solidago canadensis, Taraxacum sp, Xanthium strumarium, Ericaceae -Erica sp, Oleaceae - Fraxinus excelsior, Boraginaceae - Echium italicum, officinale, Convolvulaceae - Convolvulus cantabrica, Symphytum Polemoniaceae - Phlox drummondi, Scrophulariaceae - Antirrhinum majus, Linaria vulgaris, Verbanaceae - Verbena officinalis, Laminceae - Lavandula agustifolia, L. stoechas, Mentha pulegium, Rosmarinus, Plantaginaceae -Plantago lanceolata,

Chenopodiaceae - Chenopodium sp, Polygonaceae - Fagopyrum esculentum,
Lauraceae - Laurus nobilis, Elaeagnaceae - Hippophae rhamnoides,
Loranthaceae - Loranthus sp, Euphorbiaceae - Chrozophora sp, Urticaceae Urtica dioica, Ulmaceae - Ulmus procera, Fagaceae - Quercus robur,
Salicaceae - Populus tricocarpa, Musaceae - Musa paradisiaca, Iridaceae Iris unguicularis, Liliaceae - Asparagus acutifolis, Smilacaceae - Smilax
aspera, Poaceae - Zea mays, Alopecurus pratensis, Cupressaceae - Cupressus
sempevirens.

Plate - 1

Figure - 1

Genus - Helleborus

Species - niger

Shape - Irregularly round

 $\begin{array}{ccc} \text{Size} & & -35\,\mu \\ \text{Pore / colpi} & & -\text{Furrows} \end{array}$ 

Number of pollen grains - 15

Plate - 1

Figure - 4

Genus - Magnolia

Species - grandifolia

Shape - Oblate Size - 51.5 μ

Pore / colpi - Monocolporate

Number of pollen grains - 28

Plate -

Figure -

Genus - Berberis

Species - darwinii

Shape - Irregularly round

Size - 40.1 μ

Pore / colpi - United or irregular furrows may occur

Figure - 6

Genus

- Brassica

Species - napus

Shape

- Oblate spheroid

Size - 24.7 μ

Pore / colpi

- Tricolporate

Number of pollen grains - 22

Plate - 2

Figure - 7

Genus

- Cardamine

Species - pratensis

Shape

Iregularly round

Size

- 24.6 µ

Pore / colpi - Furrows only

Number of pollen grains - 29

Plate - 2

Figure - 8

Genus

- Viola

Species - tricolor

Shape

- Multisided or irregular

Size

- 80.4 µ

Pore / colpi

- Furrows with pores

Number of pollen grains - 80

Plate - 3

Figure - 9

Genus

- Lychnis

Species - flos-cuculi

Shape

- Spheric

Size

- 45.5 µ

Pore / colpi

- Periporate

- Abutilon Genus

Shape - Suboblate Size - 58.3 µ Pore / colpi - Tricolporate

Number of pollen grains - 1

Species - indicum

Plate - 4

Genus

- Hibiscus

Shape - Spheric Size - 147.5 µ Pore / colpi - Periporate

Number of pollen grains - 32

Figure - 14

Species – rosa-sinesis

Plate - 5

Figure - 15

Genus - Tilia Species – petiolaris

- Triangular/irregular Shape

Size - 35.6 µ

Pore / colpi - Furrows with pores

Number of pollen grains - 13

Plate - 5

Figure - 16

Species - pratense Genus - Geranium

- Oblate spheroidal Shape

- 47.2 µ Size - Tricolporate Pore / colpi

Figure - 17

Genus

- Geranium

Species - rotundifolium

Shape

- Oblate spheroidal

Size

- 45.3 µ

Pore / colpi

- Tricolporate

Number of pollen grains -20

Plate - 6

Figure - 18

Genus

- Oxalis

Species - articulata

Shape

Oblate spheroidal

Size

- 32.5 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 13

Plate - 7

Figure - 19

Genus

- Impatiens

Species – glandulifera

Shape

Long

Size

- 42.7 µ

Pore / colpi

- Furrows only

Number of pollen grains -13

Plate - 7

Figure - 20

Genus

- Citrus

Species - hmon

Shape

- Oblate spheroidal

Size

- 40.2 µ

Pore / colpi

- Tetracolporate

Figure - 21

Genus

- Parthenocissus

Species - tricuspidata

Shape

- Prolate spheroidal

Size

- 38.3 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 39

Plate - 8

Figure - 25

Genus

- Pistacia

Species - kentiscus

Shape

 Oblate spheroidal - 28.3 µ

Size Pore / colpi

- Stephanoporate

Number of pollen grains - 1

Plate - 8

Figure - 26

Genus - Platanus

Species - orientalis

Shape

- Subprolate

Size - 24.3 μ

Pore / colpi

- Tricolporate

Number of pollen grains - 21

Genus

- Colutea

Species - arborescens

Shape

- Oblate spheroidal

Size

- 35.2 µ

Pore / colpi - Tricolporate

Figure - 29

Genus

- Gleditsa

Species - triancanthos

Shape

- Oblate spheroidal

Size

- 32.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 35

Genus – Onobrychis

Species - vicifolia

Shape Size

- Prolate - 27.9 u

Pore / colpi

- Tricolporate

Number of pollen grains - 10

Plate - 12

Figure - 35

Genus

- Ononis

Species – pubescens

Shape Size

 Prolate - 38.6 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 37

Plate - 13

Figure - 37

Genus

- Sophora

Species - japonica

Shape

- Prolate spheroidal

Size

- 17.2 µ

Pore / colpi

- Tricolporate

Figure - 38

Genus

- Ulex

Species - galli

Shape

- Oval flattened

- 43.5 µ

Pore / colpi

- Furrows only

Number of pollen grains - 7

Plate - 13

Figure - 40

Genus

- Cassia

Species – didimobtrya

Shape

Oblate spheroidal

Size

- 34.2 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 37

Plate - 14

Figure – 41

Genus

- Acacia

Species - dealbata

Shape Size

- 54.1 μ

Pore/Colpi

Inaperturate

Number of pollen grains - 46

Plate - 14

Figure - 42

Genus

- Crataegus

Species - monogyna

Shape

- 37.5 µ Size

Pore / colpi

- Tricolporoidate

Oblate spheroidal

Genus - Eriobotrya

Shape – Suboblate – 24.4 μ

Pore / colpi - Tricolporoidate

Number of pollen grains - 43

Plate - 15

Figure - 43-44

Species - domestica

Species - japonica

Genus - Malus

 $\begin{array}{ccc} Shape & & - Suboblate \\ Size & & - 27.2 \ \mu \\ Pore \ / \ colpi & & - Tricolporate \end{array}$ 

Number of pollen grains - 52

Plate - 16

Figure - 45

Species - dulcis

Genus - Prunus

Shape – Suboblate Size - 43.4 μ

Pore / colpi - Tricolporoidate

Number of pollen grains - 64

Plate - 16

Figure – 46

Genus - Pyrus

Species - cummunis

Shape - Triangular Size - 46.7 μ

Pore/Colpi - Furrows with pores

Figure - 49

Genus

- Myrtus

Species - communis

Shape Size

Oblate
 10.7 μ

Pore / colpi

- Tricolporate/Syncolpate

Number of pollen grains - 100

Plate - 17

Figure - 50

Genus

- Passiflora

Species - edulis

Shape Size – Spheric - 64.8 μ

Pore / colpi

- Spiraperture Syncolpate

Number of pollen grains - 3

Plate - 18

Figure - 51

Genus

- Bryonia

Species - dioica

Shape

Irregularly round

Size

- 38.4 µ

Pore / colpi

- Furrows with pores

Number of pollen grains - 15

Plate - 18

Figure - 52

Genus

- Citrullus

Species - kanatus

Shape

Oblate spheroidal

Size

- 115.7 μ

Pore / colpi

- Periporate

Figure - 55

Genus

- Bupleureum

Species - fruticosum

Shape Size

- Prolate - 18.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 31

Plate - 19

Figure - 56

Genus

-Symrnium

Species - olustrum

Shape Size

- Prolate - 30.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains -2

Plate - 20

Figure - 57

Genus

- Hedera

Species - helix

Shape

- 32.3 µ

Size Pore / colpi - Tricolporate

Oblate spheroidal

Number of pollen grains - 2

Plate - 20

Figure - 58

Genus

- Cornus

Species - sanguina

Shape

- Prolate spheroidal

Size

- 54.3 µ

Pore / colpi

- Tricolporate

Figure - 59

Genus

- Ageratum

Species - conizoides

Shape

Size

Pore / colpi -

Number of pollen grains - 508

Plate - 21

Figure - 61

Genus

- Aster

Species -

Shape

- Triagular Size - 23.5 μ

Pore / colpi

- Furrows with pores

Number of pollen grains - 333

Plate - 21

Figure - 62

Genus

- Carduus

Species -

Shape

Oblate spheroidal

Size

- 42.4 µ

Pore / colpi - Tricolporate

Number of pollen grains - 150

Plate - 21

Figure - 63

Genus

- Carthamus

Species -

Shape

- Subprolate

Size

- 60.6 µ

Pore / colpi - Tricolporate

Figure - 64

Genus

- Cirsium

Species -

Shape

Oblate spheroidal

Size

- 42.1 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 12

Genus

- Eupatorium

Species – japonica

Shape

Oblate spheroidal

Size

- 22.5 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 103

Plate - 22

Figure - 65

Genus

- Halianthus

Species - annuus

Shape Size

- 38.4 µ

Pore / colpi

- Tricolporate

Oblate spheroidal

Number of pollen grains - 275

Plate - 22

Figure - 66

Genus

- Inula

Species -

Shape

Oblate spheroidal

Size

- 31.2 µ

Pore / colpi

- Tricolporate

Figure - 67

Genus

- Matricaria

Species -

Shape

- Oblate spheroidal

Size

- 23.6 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 15

Plate - 22

Figure- 68

Genus

- Michenia

Species - micrantha

Shape

Size

Pore / colpi

Number of pollen grains - 820

Genus

- Senecio

Species -

Shape Size

Oblate spheroidal28.2 μ

Pore / colpi

- Tricolporate

Number of pollen grains - 10

Plate - 23

Figure - 69

Genus

- Solidago

Species – canadensis

Shape

Oblate spheroidal

Size

- 22.4 µ

Pore / colpi

- Tricolporate

Figure - 70

Genus

- Taraxacum

Species -

Shape

- Oblate spheroidal

Size

- 34.3 µ

Pore / colpi

Tricolporate/Tetracolporate

Number of pollen grains - 92

Plate - 23

Figure - 71

Genus

- Xanthium

Species - strumarium

Shape

– Spherical - 26.8 μ

Size Pore / colpi

- Tricolporate

Number of pollen grains - 197

Plate - 24

Figure – 73

Genus

- Erica

Species -

Shape

паре

Size

 $-32.4~\mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 29

Plate - 24

Figure - 75

Genus

- Fraxinus

Species – excelsior

Shape

Oblate spheroidal

Size

- 19.5 µ

Pore / colpi

- Tricolporoidate

Figure - 78

Genus

- Echium

Species - italicum

Shape

- Prolate

Size

- 18.7 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 16

Plate - 25

Figure - 79

Genus

- Symphytum

Species - officinale

Shape Size - Prolate - 27.3 µ

Pore / colpi - Stephanocolporate

Number of pollen grains - 32

Plate - 25

Figure - 80

Genus

- Convolvulus

Species - arvensis

Shape Size - Oblate spheroidal - 53.6 µ

Pore / colpi - Tricolporate

Number of pollen grains - 11

Genus

- Pholx

Species – drummondi

Shape

Size - 57.5 µ

Pore / colpi

- Pores only

Irregularly round

Figure - 81

Genus

- Antirrhinum

Species - indicum

Shape

Oblate spheroidal

Size - 12.5 μ

Pore / colpi

- Tricolporate/Syncolporate

Number of pollen grains - 7

Genus

- Linaria

Species - vulgaris

Shape

Irregularly round

Size

- 14.2 µ

Pore / colpi

- Furrows with pores

Number of pollen grains - 14

Plate - 27

Figure - 83

Genus

- Verbena

Species - officinalis

Shape

Oblate spheroidal

Size

- 25.4 µ

Pore / colpi

Heterocolpate

Number of pollen grains - 4

Plate - 27

Figure - 84

Genus

- Lavandula

Species – agustifolia

Shape

Oblate

Size - 25.5 μ

Pore / colpi

- Stephanocolpate

Genus - Lavandula

Species - stoechas

Shape – Oblate Size – 26.2 μ

Pore / colpi - Stephanocolpate

Number of pollen grains - 9

Plate - 28

Figure - 85 - 86

Genus - Mentha

Species – pulegium

Shape – Oblate spheroidal

Size - 18.5 μ

Pore / colpi - Stephanocolpate

Number of pollen grains - 22

Plate - 29

Figure – 87 - 88

Genus - Rosmarinus

Species -

Shape – Suboblate Size – 33.2 μ

Pore / colpi - Stephanocolpate

Number of pollen grains - 5

Genus - Plantago

Species – lanceolata

Shape – Spheric
Size – 24.3 μ
Pore / colpi – Periporate

Figure - 91

Genus

- Chenopodium

Species -

Shape Size

- Spheric - 26.5 µ

Pore / colpi

- Periporate

Number of pollen grains - 11

Plate - 30

Figure – 92

Genus - Fagopyrum

Species - esculentum

Shape Size

 Subprolate - 40.4 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 4

Plate - 31

Figure - 93 - 94

Genus

- Laurus

Species - nobilis

Shape Size

- Spheric

Pore / colpi

- 55.4 µ - Inaperturate

Number of pollen grains - 63

Plate - 32

Figure - 96

Genus

- Hippophae

Species – rhamnoides

Shape

- Irregularly round

Size

- 23.4 µ

Pore / colpi

- Furrows with pores

Figure - 97

Genus

- Loranthus

Species -

Shape

Oblate

Size

- 15.4 µ

Pore / colpi

- Tricolporate/Syncolpate

Number of pollen grains - 1

Plate - 32

Figure - 98

Genus

- Chrozophora

Species -

Shape

- Suboblate - 54.6 µ

Size Pore / colpi

- Stephanocolporate

Number of pollen grains - 4

Plate - 33

Figure - 99

Genus

Urtica

Species – dioica

Shape

Irregularly round

Size

- 13.8 μ

Pore / colpi

- Pores only

Number of pollen grains - 1

Plate - 33

Figure - 100

Genus

- Ulmus

Species - procera

Shape

Size

- Suboblate - 25.5 µ

Pore / colpi

- Stephanocolporate

Figure - 104

Genus

- Quercus

Species - robur

Shape

- Oblate spheroidal

Size - 28.4 μ

Pore / colpi

- Tricolporoidate

Number of pollen grains - 33

Plate - 34

Figure - 105

Genus - Populus

Species – tricocarpa

Shape

Size

Pore / colpi

Number of pollen grains - 8

Plate - 34

Figure - 106

Genus

- Musa

Species – paradisiaca

Shape Size

Pore / colpi - Inaperturate

- Circular

- 120.6 μ

Number of pollen grains - 1

Plate - 35

Figure - 108

Genus

- Iris

Species – unguicularis

- Round Shape - 78.4 µ Size

- Furrows only Pore / colpi

Genus - Asparagus

Shape – Oblate Size – 16.4 μ

Pore / colpi - Monocolpate

Number of pollen grains - 27

Genus - Smilax

Shape - Spheric - 20.5

Pore / colpi - Inaperturate

Number of pollen grains - 4

Plate - 36

Genus - Alopecurus

 Shape
 - Round

 Size
 - 14.5 μ

 Pore / colpi
 - Pores only

Number of pollen grains - 44

Species – acutifolis

Species – aspera

Figure - 111

Species - pratensis

Plate - 36 - 37

Genus - Zea

Shape – Round – 70.4 μ

Pore / colpi - Monoulcerate

Number of pollen grains - 277

Figure – 112 - 113

Species - mays

Figure - 117

Genus - Cupressus

Species - sempevirens

 $\begin{array}{ccc} Shape & & - Spheroidal \\ Size & & - 24.1 \ \mu \end{array}$ 

Pore / colpi - Monoporate

Number of pollen grains - 1

## SUMMER SAMPLES

## Kubza Village

Summer samples of honey obtained from Kubza Village of Mokokchung district reveals the presence of following types of pollen grains belonging to various members of respective families in the honey collected by bees. The number of pollen grains is given per ml of honey sample.

Ranunculaceae - Clematis vitalba, Papaveraceae -Papaver rhoes, Brassicaceae - Mathiola sp, Malvaceae - Gossypium, Tiliaceae - Tilia petiolaris, Geraniaceae - Geranium pratense, G. rotundifolium, Aceraceae - Acer sp, Platanaceae - Platanus orientalis, Fabaceae - Coronilla, Glycine max, Melilotus alba, Sophora sp, Caesapinasceae - Cassia didimobotrya, Rossaceae - Filipendula ulmaria, Myrtaceae - Myrtus communis, Cucurbetaceae - Citrullus lanatus, Cucumis sativus, Cucurbita pepo, Diplotaxis erucoides, Apiaceae - Bupleurum fruticosum, Cornaceae -Cornus sanguinea, Caprifoliaceae - Sambucus nigera, Asteraceae - Ambrosia maritime, Chrysanthemum sp, Matricaria sp, Michenia micrantha, Boraginaceae - Cynoglossum creticum, Loranthaceae - Loranthus europaeus, Santalaceae - Thesium humile, Corylaceae - Corylus avellana, Carpinus betulus, Fagaceae - Fagus sylvatica, Salicaceae - Populus Musaceae - Musa paradisiaca, Amaryllidaceae - Galanthus nivalis, Poaceae - Zea mays.

Figure - 1

Genus - Clematis

Species – vitalba

Shape

Suboblate

Size

- 20.2 µ

Pore / colpi

- Tricolpate

Number of pollen grains - 2

Plate - 40

Figure - 3

Genus - Papaver

Species - rhoes

Shape

- Oblate spheroidal

Size Pore / colpi - 23.6 µ - Tricolpate

Number of pollen grains - 3

Genus - Mathiola

Species-

Shape Size

Spheric - 23.5 μ

Pore / colpi

- Inaperturate

Number of pollen grains - 19

Genus - Gossypium

Species-

Shape Size Spheric

Pore / colpi

- 123.5 µ - Periporate

Figure - 8

Genus - Tilia

Species - petiolaris

Shape

- Triangular/irregular

Size

- 35.6 µ

Pore / colpi

- Furows with pores

Number of pollen grains - 2

Plate - 43

Figure - 9

Genus - Geranium

Species -pratense

Shape

Oblate spheroidal

Size

- 45.3 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 25

Plate - 43

Figure - 10

Species - rotundifolium

Genus - Geranium

Oblate spheroidal

Shape Size

- 47.2 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 6

Plate - 44

Figure - 12

Genus - Acer

Species -

Shape

Oval flattened

Size

- 42.8 µ

Pore / colpi

- Furrows with pores

Figure - 14

Genus - Platanus

Species - orientalis

Shape – Subprolate Size – 24.2 μ

Pore / colpi - Tricolporate

Number of pollen grains - 3

Plate - 45

Figure - 15

Genus - Coronilla

Species -

Shape – Oblate spheroidal - 27.2 μ

Pore / colpi - Tricolporoidate

Number of pollen grains -

Plate - 45

Figure - 16

Genus - Glycine

Species - max

Shape – Oblate spheroidal

- 23.6 μ

Pore / colpi - Tricolporate

Number of pollen grains - 10

Plate - 46

Figure - 19

Genus - Melilotus

Species - alba

Shape - Prolate spheroidal

 $\begin{array}{ccc} \text{Size} & & -23.7 \, \mu \\ \text{Pore / colpi} & & -\text{Tricolporate} \end{array}$ 

Figure - 20

Genus - Sophora

Species-japonica

Shape – Prolate Size – 17.2 μ

Pore / colpi - Tricolporate

Number of pollen grains - 2

Plate - 46

Figure - 21

Genus - Cassia

Species- didimobotrya

Shape –Oblate spheroidal

Size - 34.2 μ

Pore / colpi - Tricolporate

Number of pollen grains - 7

Plate - 46

Figure - 22

Genus - Filipendula

species-ulmaria

Shape – Prolate spheroidal

Size - 19.5 μ

Pore / colpi - Tricolporate

Number of pollen grains - 8

Plate - 47

Figure - 23

Genus - Myrtus

Species-communis

Shape – Oblate Size – 10.7 μ

Pore / colpi - Tricolporate/Syncolpate

Figure - 26

Genus - Citrullus

Species-lanatus

Shape

- Oblate spheroidal

Size

- 52.4 μ

Pore / colpi

- Tricolporate

Number of pollen grains -

Plate - 47

Figure - 25

Species-sativus

Genus - Cucumis

Oblate spheroidal

Shape Size

Oblate spriero
 45.5 μ

Pore / colpi

- Triporate

Number of pollen grains -

Plate -47

Figure - 24

Genus - Cucurbita

species- pepo

 $\begin{array}{lll} \text{Shape} & & -\text{Spheric} \\ \text{Size} & & -115.7 \ \mu \\ \text{Pore / colpi} & & -\text{Periporate} \end{array}$ 

Number of pollen grains - 6

Genus - Diplotaxis

Species-erucoides

 $\begin{array}{ccc} Shape & & - Subprolate \\ Size & & - 26.4 \, \mu \\ Pore \, / \, colpi & & - Tricolpate \end{array}$ 

Figure - 27

Genus - Bupleurum

Species – fruticosum

 $\begin{array}{ccc} Shape & & - Prolate \\ Size & & - 18.4 \, \mu \end{array}$ 

Pore / colpi - Tricolporate

Number of pollen grains - 14

Plate - 48

Figure - 28

Genus - Cornus

Species -sanguinea

Shape – Prolate spheroidal

 $\begin{array}{ccc} \textbf{Size} & & \textbf{-} \ \, \textbf{54.3} \ \mu \\ \textbf{Pore} \ / \ \, \textbf{colpi} & & \textbf{-} \ \, \textbf{Tricolporate} \end{array}$ 

Number of pollen grains - 1

Genus - Sambucus

Species - nigera

Shape – Suboblate – 17.9 μ

Pore / colpi - Tricolporoidate

Number of pollen grains - 2

Plate - 48

Figure - 29

Genus - Ambrosia

Species - maritima

Shape – Suboblate
Size – 18.3 µ
Pore / colpi – Tricolporate

Figure - 31

Genus - Chrysanthemum

Species -

Shape

Size

Pore / colpi

Number of pollen grains - 28

Plate - 49

Figure - 33 - 34

Genus - Matricaria

Species -Oblate spheroidal

Shape Size

- 23.6 µ

Pore / colpi

- Tricolporate

Number of pollen grains - 20

Plate - 50

Figure - 35

Genus - Michenia Shape

Species - micrantha

Size

Pore / colpi

Number of pollen grains - 12

Plate - 50

Figure - 36

Species -creticum

Genus - Cynoglossum

- Prolate

Shape Size

- 18.5 µ

Pore / colpi

- Stephanocolporate

Figure - 38

Genus - Loranthus

Species - europaeus

Shape – Oblate Size – 15.4 μ

Pore / colpi - Tricolporate/Syncolporate

Number of pollen grains - 1

Genus - Thesium

Species – humile

Shape – Oblate spheroidal

Size  $-22.6 \mu$ 

Pore / colpi - Trichotomonocolpate

Number of pollen grains - 2

Genus - Corylus

Species - avellana

 $\begin{array}{ccc} Shape & & - Triangular \\ Size & & - 29.3 \, \mu \\ Pore \, / \, colpi & & - pores \, only \end{array}$ 

Number of pollen grains - 12

Plate - 51

Figure - 42

Genus - Carpinus

Species – betulus

Shape - Irregularly round

Size - 32.6 μ
Pore / colpi - pores only
Number of pollen grains - 5

Figure - 43

Genus - Fagus

Species - sylvatica

Shape - Irregularly round

Size  $-43.4 \,\mu$ 

Pore / colpi - Furrows with pores

Number of pollen grains - 11

Plate - 52

Figure - 44

Genus - Populus

Species - nigra

 $\begin{array}{ccc} Shape & & - Spheric \\ Size & & - 26.2 \ \mu \\ Pore \ / \ colpi & & - Periporate \end{array}$ 

Number of pollen grains - 11

Plate - 52

Figure - 46

Genus - Musa

Species – paradisiaca

Shape – Circular Size – 120.6 μ Pore / colpi – Inaperturate

Number of pollen grains - 2

Plate - 53

Figure - 48

Genus - Galanthus

Boat shaped

Shape – Boat sh Size – 27.3 μ

Pore / colpi - Furrows only

Number of pollen grains - 1

Species - nivalis

Figure - 51

Genus - Zea

Species - mays

Shape – Round - 70.4 μ

Pore / colpi - Monoulcerate

Number of pollen grains - 4

## Khensa Village

Summer samples of honey obtained from Khensa Village of Mokokchung district reveal the presence of following types of pollen grains belonging to various members of respective families in the honey collected by the bees of *Apis indica*.

Ranunculaceae - Ranunculus bulbosus, Brassicaceae Diplotaxis erocoides, Malvaceae - Hibiscus rosa - sinensis, Tiliaceae - Tilia
petiolaris, Geraniaceae - Geranium rotundifolium, Hippocastanaceae Aesculus hippocastanum, Platanaceae - Platanus orientalis, Fabaceae Glycine max, Phaseolus coccineus, Robinia pseudoacacia, Caesalpiniaceae Cassia didimobotrya, Rosaceae - Filipendula ulmaria, Myrtaceae Eucalyptus gunnii, Cucurbitaceae - Cucumis sativus, Apiaceae - Bupleurum
fruticosum, Asteraceae - Artemesia vulgaris, Chrysanthemum sp, Michenia
micrantha, Polyganaceae - Fagopyrum esculentum, Santalaceae - Thesium
humile, Euphorbiaceae - Mercurialis perennis, Cotoneaster horizontalis,
Corylaceae - Corylus avellena, Fagaceae - Facus sylvatica, Thypaceae - Thypa
angustifolia, Musaceae - Musa paradisiaca, Iridaceae - Iris pseudoacorus,
Liliaceae - Allium cepa, Poaceae - Alopecurus pratensis, Zea mays

Genus - Ranunculus

Species - bulbosus

Shape – Suboblate

 $\begin{array}{ll} \text{Size} & -28.5 \ \mu \\ \text{Pore/Colpi} & -\text{Tricolpate} \end{array}$ 

Figure - 5

Genus

- Diplotaxis

Species – erocoides

Shape

- Subprolate

Size

 $-26.4 \,\mu$ 

Pore/Colpi

- Tricolpate

Number of pollen grains - 2

Plate - 41 - 42

Figure - 6 - 7

Shape

Genus - Hibiscus

Species – rosa-sinensis

Size

 Spheric  $-147.5 \,\mu$ 

Pore/Colpi

-24

Number of pollen grains -

Plate - 42

Figure - 8

Genus - Tilia

Species - petiolaris

Shape

- Triangular/irregular

Size

 $-35.6 \mu$ 

Pore/Colpi

- Furrows with pores

Number of pollen grains - 5

Plate - 43

Figure – 10

Genus - Geranium

Species - rotundifolium

Shape

Oblate spheroidal

Size

 $-45.3 \mu$ 

Pore/Colpi

- Tricolporate

Figure – 11

Species - hippocastanum

Genus - Aesculus

Shape

Prolate spheroidal

Size

 $-18.5 \,\mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 3

Plate -44

Figure - 14

Genus - Platanus

Species – orientalis

Shape

- Subprolate

Size

 $-24.2 \mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 7

Plate - 45

Figure - 16

Species -max

Genus - Glycine

- Oblate spheroidal

Shape

Size

 $-23.6 \mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 1

Plate -45

Figure – 17

Species – coccineus

Shape

Genus - Phaseolus

- Irregularly round

Size

 $-32.5 \mu$ 

Pore/Colpi

- Pores only

Figure – 18

Genus

- Robinia

Species – pseudoacacia

Shape

- Suboblate

Size

 $-24.5 \,\mu$ 

Pore/Colpi

- Tricolporate/tricolporoidate

Number of pollen grains - 3

Plate - 46

Figure - 21

Genus - Cassia

Species – didimobotrya

 Oblate spheroidal Shape

Size

 $-34.2 \mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 2

Plate -46

Figure - 22

Genus - Filipendula

Species - ulmaria

Shape

- Prolate spheroidal

Size

 $-19.5 \mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 7

Genus

- Eucalyptus

Species – gunnii

Shape Size

- Oblate

 $-13.5 \,\mu$ 

Pore/Colpi

- Tricolporate/Syncolpate

Figure - 25

Genus - Cucumis

Species – sativus

Shape

 Oblate spheroidal Size  $-45.5 \,\mu$ 

Pore/Colpi - Triporate

Number of pollen grains - 3

Plate -48

Figure – 27

- Bupleurum

Species - fruticosum

- Prolate Shape  $-26.4 \mu$ Size Pore/Colpi - Tricolpate

Number of pollen grains - 4

Plate - 48

Figure – 30

Genus - Artemesia

Species - vulgaris

Shape Oblate spheroidal

Size – 19.6 μ Pore/Colpi - Tricolpate

Number of pollen grains -

Plate - 49

Figure – 31

Genus - Chrysanthemum

Species -

Shape Size

Pore/Colpi

Figure – 35

- Michenia Genus

Species – micrantha

Shape

Size -

Pore/Colpi

Number of pollen grains -

Plate -50

Figure – 37

Genus - Fagopyrum

Species – esculentum

Shape Subprolate

Size  $-40.4 \mu$ Pore/Colpi - Tricolporate

Number of pollen grains - 6

Genus - Thesium

Species - humile

Shape Oblate spheroidal

 $-22.6 \mu$ Size

- Trichotomonocolpate Pore/Colpi

Number of pollen grains - 8

Plate -51

Figure – 39

Genus - Mercurialis

Species – perennis

- Prolate spheroidal Shape

 $-24.3 \,\mu$ Size

- Tricolporate Pore/Colpi

Genus - Corylus

Shape - Triangular

 $\begin{array}{ccc} \text{Size} & & -29.3 \, \mu \\ \text{Pore/Colpi} & & -\text{Pores only} \end{array}$ 

Number of pollen grains - 7

Species – avellena

Plate -52

Figure - 43

Genus - Fagus

Species - sylvatica

Shape - Irregularly round

Size  $-43.4 \mu$ 

Pore/Colpi - Furrows with pores

Number of pollen grains - 3

Genus - Thypha

Species - angustifolia

Shape – Oblate Size – 18.3

Pore/Colpi – Monoporate

Number of pollen grains - 2

Plate - 52

Figure-46

Genus - Musa

Species - paradisiaca

 $\begin{array}{ccc} Shape & & - Circular \\ Size & & -120.6 \ \mu \end{array}$ 

Pore/Colpi - Inaperturate

Genus - Iris

- Round

Size

- 78.4 μ

Pore/Colpi

Shape

- Furrows only

Number of pollen grains - 4

Plate - 53

Figure - 49

Species - pseudoacorus

Genus - Allium

Species – cepa

Shape – Oblate Size – 18.5 μ

Pore/Colpi – Monocolpate

Number of pollen grains - 26

Plate -54

Figure - 50

Genus - Alopecurus

Species - pratensis

 $\begin{array}{ccc} \textbf{Shape} & & - \; \textbf{Round} \\ \textbf{Size} & & - \; 47.5 \; \mu \\ \textbf{Pore/Colpi} & & - \; \textbf{Pores only} \end{array}$ 

Number of pollen grains - 2

Plate -54

Figure – 51

Genus - Zea

Species – mays Round

Shape – Round Size – 70.4 μ

Pore/Colpi – Monoulcerate

## Ungma Village

Summer samples of honey obtained from Ungma Village of Mokokchung district reveal the presence of following types of pollen grains belonging to various members of respective families in the honey collected by the bees of *Apis indica*. The number of pollen grains is given per ml of honey sample.

Magnoliaceae - Liriodendron tulipefera, Papaveraceae -Papaver rhoes, Brassicaceae - Brassica napus, Malvaceae - Abutilon indicum, Hibiscus rosa-sinensis, Geraniaceae - Geranium rotundifolium, Hippocastanaceae - Aesculus hippocastanum, Aceraceae – Acer sp, Anacardiaceae - Cotinus, Fabaceae - Ceratonia siliqua, Phaseolus coccineus, Robinia pseudoacacia, Rosaceae - Eriobotrya japonica, Filipendula ulmaria, Myrtaceae - Eucalyptus gunnii, Cucurbitaceae - Cucumis sativus, Apiaceae -Bupleurum fruticosum, Asteraceae - Artemesia vulgaris, Chrysanthemum sp, Helianthus annuus, Michenia micrantha, Boraginaceae - Cynoglossum creticum, Scrophulariaceae - Antirrhinum majus, Polygonaceae - Fagopyrum esculentum, Euphorbiaceae - Mercurialis sp, Cotoneaster horizontalis, Urticaceae - Urtica dioica, Juglandaceae - Juglans, Salicaceae - Salix triandra, Populus tricocarpa, Thyphaceae - Thypha Agustifolia, Musaceae -Musa paradisiaca, Iridaceae - Crocosmia, Liliaceae - Allium cepa.

Plate -40

Figure – 2

Genus - Liriodendron

Species – tulipifera

Shape – Suboblate – S1.6 μ

Pore/Colpi – Monocolpate

Figure – 3

Genus

- Papaver

Species – rhoes

Shape

Oblate spheroidal

Size

 $-23.6 \mu$ 

Pore/Colpi

- Tricolpate

Number of pollen grains - 12

Plate

-40

Figure - 4

Species -napus

Genus

- Brassica

- Oblate spheroidal

Shape Size

 $-24.7 \,\mu$ 

Pore/Colpi

- Tricolpate

Number of pollen grains - 11

Genus

- Abutilon

Species - indicum

Shape

Suboblate58.3 μ

Size Pore/Colpi

- Tricolporate

Number of pollen grains - 3

Plate

-41 - 42

Figure - 6 - 7

Genus

- Hibiscus

Species -rosa-sinensis

Shape Size - Spheric

Pore/Colpi

147.5 μPeriporate

Figure - 10

Genus

- Geranium

Species - rotundifolium

Shape

Oblate spheroidal

Size

 $-45.3\,\mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 9

Plate -44 Figure – 11

Genus

- Aesculus

Species -hippocastanum

- Prolate spheroidal Shape

Size

 $-18.5 \mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 15

- 44 Plate

Figure -12

Species -

Genus

- Acer

Oval flattened

Shape Size

 $-42.8 \mu$ 

Pore/Colpi

- Furrows with pores

Number of pollen grains - 8

Plate

-44

Figure - 13

Genus

- Cotinus

Species -

Shape

Prolate spheroidal

Size

 $-24.2 \,\mu$ 

Pore/Colpi

- Tricolporate

Genus - Ceratonia

Species -

Shape

- Oblate spheroidal

Size

 $-23.5 \mu$ 

Pore/Colpi

- Tetracolporate

Number of pollen grains - 4

Plate -45 Figure –17

Genus

- Phaseolus

Species – coccineus

Shape

- Irregularly round

Size Pore/Colpi  $-32.5 \,\mu$ 

- Pores only Number of pollen grains - 9

Plate

-45

Figure - 18

Genus

- Robinia

Species – pseudacacia

Shape

- Suboblate

Size

 $-24.5 \,\mu$ 

Pore/Colpi

- Tricolporate/tricolporoidate

Number of pollen grains - 3

Genus

- Eriobotrya

Species – japonica

Shape

- Suboblate

Size

 $-24.4 \,\mu$ 

Pore/Colpi

- Tricolporoidate

Figure - 22

Genus

- Filipendula

Species - ulmaria

Shape

- Prolate spheroidal

Size

 $-19.5 \mu$ 

Pore/Colpi

- Tricolporate

Number of pollen grains - 6

Genus – Eucalyptus

Species – gunnii

Shape

- Oblate -13.5

Size Pore/Colpi

- Tricolporate/syncolpate

Number of pollen grains - 2

Plate

-47

Figure – 25

Genus

- Cucumis

Species – sativus

Shape

Oblate spheroidal

Size Pore/Colpi

 $-45.5 \mu$ - Triporate

Number of pollen grains - 4

Plate

-48

Figure – 27

Genus

- Bupleurum

Species - fruticosum

Shape

Size

- Prolate  $-18.4 \,\mu$ 

Pore/Colpi

- Tricolporate

Figure – 30

Genus

- Artemesia

 ${\bf Species}-vulgaris$ 

Shape

- Oblate spheroidal

Size

- 19.6 µ

Pore/Colpi

- Tricolporate

Number of pollen grains – 10

Plate - 49

Figure - 31

Genus

- Chrysanthemum

Species -

Shape Size

\_

Pore/Colpi

Pl.

Number of pollen grains - 25

Plate -49

Figure-32

Species – annuus

Genus

- Helianthus

- Oblate spheroidal

Shape Size

-38.

Pore/Colpi

- Tricolporate

Number of pollen grains - 31

Plate -50

Figure – 35

Genus

- Michenia

Species - micrantha

Shape

vicnenia -

Size

- 11

Pore/Colpi

\_\_\_

Figure – 36

Genus - Cynoglossum

Species – creticum

- Prolate Shape - 18.5 u Size

Pore/Colpi - Stephanocolporate

Number of pollen grains - 54

Genus - Antirrhinum

Species – majus

- Oblate spheroidal Shape

Size  $-12.5 \,\mu$ 

- Tricolporate/syncolpate Pore/Colpi

Number of pollen grains - 3

Plate - 50

Figure – 37

Genus - Fagopyrum Species – esculentum

 Subprolate Shape Size  $-40.4 \,\mu$ 

- Tricolporate Pore/Colpi

Number of pollen grains - 19

Plate -51 Figure – 39

Genus - Mercurialis Species -

 Prolate spheroidal Shape

 $-24.3 \,\mu$ Size

Pore/Colpi Tricolporate

Figure – 40

Genus - Urtica

Species – dioica

Shape - Irregularly round

Size  $-13.8 \mu$ Pore/Colpi - Pores only

Number of pollen grains - 8

Plate -51

Figure – 41

Genus - Juglan Species -

Shape Size

Pore/Colpi

Number of pollen grains - 4

Plate -52 Figure – 45

Genus - Salix

- Subprolate

Shape Size

 $-18.4 \,\mu$ 

Pore/Colpi - Tricolporate

Number of pollen grains -

Species -triandra

Plate -52 Figure - 44

Genus - Populus Species -tricocarpa

- Spheric Shape  $-26.2 \,\mu$ Size Pore/Colpi – Periporate

Genus - Thypha

Shape – Oblate Size – 18.3

Pore/Colpi – Monoporate

Number of pollen grains - 6

Species – angustifolia

Plate - 52

Figure - 46

Species – paradisiaca

Genus - Musa

Shape – Circular Size – 120.6 μ

Pore/Colpi – Inaperturate

Number of pollen grains - 5

Plate -53

Figure - 47

Genus - Crocosmia

Species –

Shape – Oval flattened/semi circular Size – 66.8 μ

Pore/Colpi – Furrows only

Number of pollen grains - 11

Plate -53

Figure – 49

Genus - Allium

Species - cepa

Pore/Colpi – Monopolate

## MARKET SAMPLES

Market samples of honey obtained from Mokokchung district reveal the presence of following types of pollen grains belonging to various members of respective families in the honey collected by the bees of *Apis indica*. The number of pollen grains is given per ml of honey sample.

Brassicaceae - Brassica napus, Capparidaceae - Capparis spinosa, Violaceae - Viola tricolor, Balsaminaceae - Impatiens glandulifera, Meliaceae - Melia azedarach, Fabaceae - Glycine max, Melilotus alba, Ononis pubescens, Trifolium pretense, Rosaceae - Prunus domestica, Cucurbitaceae - Cucumis sativus, Apiaceae - Bupleurum fruticosum, Boraginaceae - Alkanna tinctoria, Echium italicum, Convolvulaceae - Convolvulus arvensis, Lauraceae - Laurus nobilis, Fagaceae - Castanea sativa, Liliaceae - Asparagus acutifolius.

Plate - 55

Figure - 1

Genus - Brassica

Species - napus

Shape - Oblate spheroidal Size - 24.7 μ

Pore/Colpi - Tricolporate

Number of pollen grains - 1

Genus - Capparis

Species - spinosa

Shape - Prolate spheroidal

Size - 29.5 μ
Pore/Colpi - Tricolpate

Genus - Viola

Species - tricolor

Shape - Multisided or irregular

- 80.4 µ

Size

Pore/Colpi – Furrows with pores

Number of pollen grains - 2

Plate - 55

Figure - 2

Genus - Impatiens

Species – glandulifera

 $\begin{array}{ccc} \text{Shape} & & \text{- Long} \\ \text{Size} & & \text{- }42.7~\mu \end{array}$ 

Pore/Colpi – Furrows only

Number of pollen grains - 1

Genus - Melia

Species – azedarach

Shape - Oblate spheroidal

Size  $-39.5 \mu$ 

Pore/Colpi - Tetracolporate

Number of pollen grains - 1

Plate -55

Figure – 3

Genus - Glycine

Species -max

Shape - Oblate spheroidal

- 23.6 μ

Pore/Colpi - Tricolporate

Figure – 4

Genus

- Melilotus

Species - alba

Shape

- Prolate spheroidal

Size - 23.7 μ

Pore/Colpi

- Tricolporate

Number of pollen grains - 3

Genus

- Ononis

Species – pubescens

Shape Size

- Prolate - 38.6 µ

Pore/Colpi

- Tricolporate

Number of pollen grains - 1

Genus

- Trifolium

Species – pratense

Shape

- Prolate spheroidal

Size

- 42.4 µ

Pore/Colpi

- Tricolporate

Number of pollen grains - 3

Plate

-56

Figure - 5

Genus

- Prunus

Species – domestica

Shape

- Suboblate

Size

- 43.4 u

Pore/Colpi

- Tricolporoidate

Genus - Cucumis

Shape - Oblate spheroidal

 $\begin{array}{ccc} \text{Size} & -45.5 \; \mu \\ \text{Pore/Colpi} & -\text{Triporate} \end{array}$ 

Number of pollen grains - 1

Plate - 56

Genus - Bupleurum

Shape - Prolate - 18.4 μ

Pore/Colpi - Tricolporate

Number of pollen grains - 7

Plate - 56

Genus - Alkanna

 $\begin{array}{ccc} Shape & & - Prolate \\ Size & & - 12.5 \ \mu \end{array}$ 

Pore/Colpi - Tricolporate

Number of pollen grains - 1

Genus - Echium

Shape - Prolate Size - 18.7 μ

Pore/Colpi - Tricolporate

Number of pollen grains - 2

Species – sativus

Figure - 6

Species - fruticosum

Figure – 7

Species – tinctoria

Species - italicum

Figure - 8

Genus

- Convolvulus

Species - cantabrica

Shape

- Oblate spheroidal

Size

- 53.6 µ

Pore/Colpi

- Tricolpate

Number of pollen grains - 2

Genus

- Laurus

Species – nobilis

Shape Size

- Spheric - 55.4 µ

Pore/Colpi

- Inaperturate

Number of pollen grains - 2

Genus - Castanea

Species – sativa

Shape Size

- Prolate - 15.7 µ

Pore/Colpi

- Tricolporate

Number of pollen grains - 8

Genus

- Asparagus

Species – acutifolius

Shape Size

- Oblate - 16.4 µ

Pore/Colpi

- Monocolpate

Table – 1. Showing the names of families, genera, species and the number of respective pollen present in the winter honey sample of kubza village per 10 gms

Sl.No.	Family	Genera	Species	No. of Pollen grains
1	Ranunculaceae	Clematis	vitalba	80
	The bost carrie	Helleborous	niger	10
2	Magnoliaceae	Magnolia	grandiflora	90
3	Brassicaceae	Brassica	napus	630
4	Violaceae	Viola	tricolor	5790
5	Cauyophyllaceae	Stellaria	media	90
	Passion	Spergularia	rupicola	80
6	Cistaceae	Helianthemum	chamaecistis	10
7	Malvaceae	Hibiscus	rosa sinensis	170
	Aprecess	Abutilon	theophrasti	80
8	Convolvulaceae	Convolvulus	cantribrica	40
9	Geraniaceae	Geranium	rotuntifolium	110
10	Balsaminaceae	Impatiens	glandulifera	210
11	Rutaceae	Citrius	limon	230
12	Vitaceae	Vitis	vinifera	80
13	Hippocastanaceae	Aesculus	hippocastanum	20
14	Anarcardiaceae	Pistacia	lentiscus	40
15	Platanaceae	Platanus	orientalis	30
		Acacia	dealbata	310
16	Fabaceae	Ulex	galli	60
		Ulex	europacus	60
		Erythrina	indica	60

		Demodium	sp	
		Ononis	pubescens	40
	Ebenactuse	Lupinus	albus	50
17	Caesalpinaceae	Cassia	didimobotrya	100
18	Rosaceae	Malus	domestica	70
		Pyrus	communis	800
		Prunus	dulcis	980
19	Crassulaceae	Sedum	acre	20
20	Myrtaceae	Eucalyptus	gunii	390
21	Passifloraceae	Passiflora	eduli	50
22	Cucurbitaceae	Ecballium	elaterium	140
		Citrullus	lanatus	150
23	Apiaceae	Bupleurum	froticosum	210
24	Araliaceae	Hedera	helix	60
25	Cornaceae	Cornus	sanguinea	450
26	Asteraceae	Ageratum	conyzoides	3400
		Arctium	sp	490
	Plantaginaceae	Carduus	sp	10000
	Polygonacead	Carthamus	sp	70
	Lauraceae	Cirsium	sp	2400
	Thymelacaceae	Eupatorium	sapoicum	300
	Blacagnaccae	Helianthus	annyus	5990
	Licembecces	Inula	sp	1700
	Eunbarblache	Matricaria	sp	510
	Uriescene	Michenia	micranths	5170

	Blmaces	Senecio	sp	130
	Schalaceus	Solidago	canadensis	
27	Ebenaceae	Diospyros	kaki	20
28	Oleaceae	Fraxinus	excelsior	50
29	Boraginaceae	Cynoglossum	creticum	130
	Etflaceae-	Syniphytum	officinal	40
		Alkanna	tinctoria	100
30	Convolvulaceae	Convolvulus	contabrica	40
31	Polemoniaceae	Phlox	drummondii	70
32	Scrophulariaceae	Antirrhinum	majus	160
	Cupressies	Verbena	thapsus	100
		Linaria	vulgaris	30
33	Verbenaceae	Verbena	officinalis	70
34	Lamiaceae	Thymus	sp	120
		Lavandula	agustifolia	20
	Pantis	Rosmarinus	sp	10
	Rangingulacens	Teucrium	sp	10
35	Plantaginaceae	Plantago	lanceolata	37
36	Polygonaceae	Fagopyrom	esculentum	10
37	Lauraceae	Laurus	sp	30
38	Thymelacaceae	Daphne	gnidium	20
39	Elaeagnaceae	Hippophae	rhamnoides	10
40	Loranthaceae	Loranthus	sp	30
41	Euphorbiaceae	Chrozophora	sp	110
42	Urticaceae	Urtica	dioica	10

43	Ulmaceae	Ulmus	procera	160
44	Betulaceae	Alnus	glutinosa	50
45	Fagaceae	Quercus	robur	43
46	Musaceae	Musa	paradisiaca	20
47	Iridaceae	Iris	unguicularis	30
48	Liliaceae	Allium	sphaerocephalum	320
		Asparagus	acutifolius	230
49	Smilacaceae	Smilax	aspera	40
50	Poaceae	Alopecurus	pratensis	20
51	Pinaceae	Pines	sylvestris	20
52	Cupressaceae	Cupressus	sempervirens	50

Table – 2. Showing the names of families, genera, species and the number of respective pollen present in the winter honey sample of Ungma village per 10 gms

SI.No.	Family	Genera	Species	No. of Pollen grains
1	Ranunculaceae	Helleborus	niger	70
		Clematis	vitalba	110
2	Magnoliaceae	Magnolia	grandiflora	260
		Liriodendron	tulipifera	20
3	Berberidaceae	Berberis	darwinii	700
4	Papaveraceae	Нуресоит	procumben	80
5	Brassicaceae	Brassica	napus	580
		Cardamine	pratensis	500
		Brassica	oleifera	680
6	Violaceae	Viola	tricolor	1060

7	Cayophyllaceae	Lychnis	flos-cuculi	40
8	Tamaricaceae	Tamarix	sp	50
9	Malvaceae	Hibiscus	rosa sinensis	120
		Malva	sylvestries	20
10	Tiliaceae	Tilea	petiolories	230
11	Geraniaceae	Geranium	rotuntifolium	110
	Camalpinicese	Geranium	Pratense	100
12	Oxalidaceae	Oxalis	articulata	10
13	Rutaceae	Citrus	limon	50
14	Vitaceae	Vitis	vinifora	890
		Parthenocissus	tricuspidate	950
15	Aceraceae	Acer	pseudoplatanus	10
16	Anarcardiaceae	Pistacia	lentiscus	30
17	Platanaceae	Platanus	orientalis	35
18	Fabaceae	Calycotone	spinosa	220
	Cucarbinaciae	Ceratonia	siliqua	200
		Coronilla	emeros	260
		Colutea	arborescens	700
	Aplaces	Gleditsia	triancanthus	270
	Aminen	Lupinus	albus	310
	Comosene	Onobrychis	vicifalia	290
728	Asieracese	Medicago	sativa	90
		Ononis	pubescens	470
		Ononis	spinosa	420
		Vicia	sp	310

		Vicia	faba	20
		Sophora	japonica	1430
		Trifolium	alexandrinum	540
		Acacia	dealbata	390
		Erythrina	indica	270
		Desmodium	sp	450
19	Caesalpinaceae	Cassia	didmobotrya	390
20	Rosaceae	Malus	domestica	900
		Crataegus	monogyna	600
	Hunkepinsees	Prunus	dulcis	810
	Ebenacere "	Pyrus	communis	1180
21	Crassulaceae	Sedum	acre	520
22	Myrtaceae	Myrtus	communis	1410
	Boraginetese	Eucalyptus	gunii	1980
23	Passifloraceae	Passiflora	edulis	100
24	Cucurbitaceae	Bryonia	dioica	120
	Convolvulacon	Citrullus	lanatus	120
	Polemenineas	Cucurbita	Pepo	10
25	Apiaceae	Bupleurum	fruticosum	220
26	Araliaceae	Hedera	helix	230
27	Cornaceae	Cornus	sangrinea	800
28	Asteraceae	Ageratum	conycoides	1960
		Aster	sp	2030
		Carduus	sp	410
		Carthamus	sp	1480

	Plantagruccae	Eupatorium	sp	2420
	Cheropodiacese	Helianthus	sp	1040
	Polytomocae	Inula	sp	370
	Laurens	Michenia	sp	6200
	Epothsonia	Solidago	sp	880
	Buckerbingers	Taraxacum	officinale	980
	Uniceses	Xanthtium	italicum	2490
29	Ericaceae	Arbutus	unedo	10
	Betilaccae	Eorica	sp	410
30	Plumbeginaceae	Armeria	maritima	720
31	Ebenaceae	Diospyros	kaki	50
32	Oleaceae	Fraxinus	Excelsior	20
33	Buddejaceae	Buddleja	davidii	130
34	Boraginaceae	Symphytum	officinale	430
	Pinaccae	Alkanna	tinctoria	160
		Borogo	officinalis	140
35	Convolvulaceae	Convolvulus	cantabrica	180
36	Polemoniaceae	Phlox	drummondii	140
37	Schrophulariaceae	Verbascum	thapsus	10
	Famuy	Linaria	vulgaris	40
38	Verbenaceae	Verbena	officinalis	40
39	Lamiaceae	Thymus	sp	220
	Denocracese	Lavandula	stoechas	20
	Distriction of the last of the	Mentha	pulegium	220
		Teucrium	sp	10

40	Plantaginaceae	Plantago	lanceolata	60
41	Chenopodiaceae	Chenopodium	sp	160
42	Polyfonaceae	Fagopyrum	esculentum	60
43	Lauraceae	Laurus	nobilis	810
44	Lornthaceae	Loranthus	sp	20
45	Euphorbiaceae	Chrozophora	sp	100
46	Urticaceae	Urtica	dioica	60
47	Ulmaceae	Ulmus	procera	40
48	Betulaceae	Alnus	futinosa	10
49	Fagaceae	Quercus	rubur	210
50	Liliaceae	Allium	sphaerocepalum	440
14	Anarcaudineser	Asphodelus	microcarpus	30
15	Platanecese	Asparacus	acutifolius	350
51	Poaceae	Zea	mays	2310
52	Pinaceae	Pinus	insularis	20

Table – 3 showing the names of families, genera, species and the number of respective pollen present in the winter honey sample of Khensa village per 10 gms

Sl.No.	Family	Genera	Species	No. of Pollen grains
1	Ranunculaceae	Helleborus	niger	150
2	Magnoliaceae	Magnolia	grandflora	280
3	Berberidaceae	Berberis	darwinii	430
4	Brassicaceae	Brassica	пари	22
		Candamin	pratensis	290
5	Violaceae	Viola	tricolor	800

6	Caryophyllaceae	Lychnis	flos – cuculi	70
7	Malvaceae	Abutilen	indicum	10
	Passifinguese	Hisbiscus	rosa – sinensis	220
8	Tiliaceae	Tilia	petiolaris	130
9	Geraniaceae	Geranium	rotuntifoloous	200
		Gerantum	pratense	130
10	Oxalidaceae	Oxalis	articulate	130
11	Balsaminaceae	Impatiens	glandulifera	130
12	Rutaceae	Citrus	limon	210
13	Vitaceae	Vitis	sp	
	Astronomic Co.	Perthenocissus	tricuspidata	390
14	Anarcaudiaceae	Pistacia	lentiscus	10
15	Platanaceae	Platacia	orientalis	210
		Acacia	dealbata	460
16	Fabaceae	Colutea	arborescens	45
		Gleditsa	iniancanthus	350
		Onoprychis	vicifolia	100
		Ononsi	pubescens	370
		Sophora	taponica	370
		Vlex	gall	70
17	Caesalpinaceace	Casesia	didimobotrya	370
18	Rosaceae	Eriobotrya	japonica	390
		Malus	domestica	520
	to the same	Prunus	dulcis	640
		Crataegus	monogyna	37

		Pyrus	communis	650
19	Myrtaceae	Myrtus	communis	1000
20	Passifloraceae	Passiflora	edulis	30
21	Cucurbitaceae	Bryonia	dioica	150
	Bridge Section:	Citrullus	lanatus	200
		Ecballium	elaterium	300
22	Apiaceae	Bupleureum	fruticosum	310
	Tables in	Smyrnium	olusatrum	20
23	Araliacae	Hetera	helix	20
24	Cornaceae	Cornus	sanguinea	730
25	Asteraceae	Ageratum	conyzoides	5080
		Aster	sp	3330
	Samuel and the	Cartuns	sp	1500
	Patrician in	Carthamus	sp	170
		Circium	sp	120
	Charles and	Eupatorium	sp	1030
		Inula	sp	220
	Selection of the select	Heloanthus	sp	2750
		Michenia	sp	150
		Senecio	sp	100
		Solidago	sp	510
		Taraxacum	officinale	920
		Xanthium	italicum	1370
26	Ericaceae	Erica	sp	290
	Oleaceae	Fraxinus	excelsior	50

27	Boraginaceae	Echium	italicum	160
	Posceae	Symphytum	officinale	320
28	Convolvulaceae	Covolvulus	cantabrica	110
29	Polemoniaceae	Phlox	drummondii	150
30	Srophuloriaceae	Anthinumm	majus	70
	4. Showing the name	Lianaria	vulgarias	140
31	Verbenaceae	Verbena	officinalis	40
32	Lamiaceae	Havandula	ajustifolia	70
	Rannelacon	Lavantula	stoechas	90
	Papaveneese	Mentha	pulegium	220
	Bussieser	Rosmarinos	sp	50
33	Plantaginaceae	Plantago	lanceolata`	330
34	Chenopodiaceae	Chenopodium	sp	110
35	Polygonaceae	Fagopyrum	esculentum	40
36	Lauraceae	Laurus	nobilis	360
37	Elaeagnaceae	Hippophae	rhamnoides	10
38	Loranthaceae	Lorathus	sp	10
39	Eubhorbiaceae	Chrozophora	sp	40
40	Urticaceae	Urtica	dioica	10
41	Ulmaceae	Ulmus	procera	130
42	Fagaceae	Querus	robur	330
43	Salicaceae	Populus	tricocarpa	80
44	Musaceae	Musa	parasidica	10
45	Iridaceae	Iris	unguicularis	20
46	Liliaceae	Asparagus	acutifolis	270

48	Smilacaceae	Smilax	aspera	40
49	Poaceae	Alopecurus	pratensis	440
		Zea	mays	2770
50	Cupressaceae	Cupressus	sempevirens	10

Table – 4. Showing the names of families, genera, species and the number of respective pollen present in the summer honey sample of Kupza village per 10 gms

CLAI	Б			No.of
Sl.No.	Family	Genera	Species	pollen grains
1	Ranunculaceae	Clematis	vitalba	200
2	Papaveraceae	Papaver	rhoes	300
3	Buassicaceae	Mathiola	sp	400
4	Malvaceae	Gossypium	sp	300
5	Tiliaceae	Tilla	petiolaris	200
6	Geraniaceae	Geranium	praense	600
		Geranium	rotundifolium	2500
7	Aceraceae	Acer	sp	800
8	Platanaceae	Platanus	orientalis	300
9	Fabaceae	Coronilla	sp	600
		Glycine	max	100
30		Melilotus	alba	500
		Sophora	sp	200
10	Eaesalpinaceae	Cassia	didimobotrya	700
11	Rosaceae	Filipendula	ulmaria	800
12	Myrtaceae	Myrtus	communis	200
13	Cucurbitaceae	Cucurbita	pepo	600
		Cucumis	sativus	700

especti		Citrullus	lanatus	400
		Diplotaxis	erucoides	200
14	Apiaceae	Bupleurum	fruticosum	1400
15	Cornaceae	Cornus	sanguinea	100
16	Caprifoliaceae	Sambucus	nigra	200
17	Asteraceae	Ambrosia	sp	1600
		Chrysanthemum	sp	2800
		Matricaria	sp	2000
		Michenia	micrantha	1200
18	Boraginaceae	Cynoglossum	cueticum	700
19	Loranthaceae	Loranthus	europaeus	100
20	Santalacene	Thesium	humile	200
21	Carylaceae	Carylus	avellana	1200
		Carpinus	betulus	500
22	Fagaceae	Facus	sylvatica	1100
23	Salicaceae	Populus	tricocarpa	1100
24	Musaceae	Musa	paradisiaca	200
25	Amaryllidaceae	Galanthus	nivalis	100
26	Poaceae	Zea	mays	400

Cocumus

Meuroltaceae

Table – 5. Showing the names of families, genera, species and the number of respective pollen present in the summer honey sample of Ungma village per 10 gms

	West bombons			
				No.of
Sl.No.	Family	Genera	Species	pollen grains
1	Magnoliaceae	Liriodendron	tulipifera	600
2	Papaveraceae	Papaver	rhoes	1200
3	Brassicaceae	Brassica	napus	1100
4	Malvaceae	Abutilon	theophrasti	300
		Hibiscus	rosa – sinensis	250
		Gossypium	sp	
5	Geraniaceae	Geraniom	rotundifolium	900
6	Hippocastanaceae	Aesculus	hippocastanum	1500
7	Aceraceae	Acer	sp	800
8	Anacardiaceae	Cotinus	sp	500
9	Tabaceae	Ceratonia	coccineus	900
		Phaseolus	pseudacacia	300
		Robinia	siliqua	400
10	Rosaceae	Eriobotrya	japonica	700
		Filipendula	ulmaria	600
11	Myrtaceae	Eucalyptus	gunii	200
12	Cucurbitaceae	Cucumis	sativus	400
13	Apiaceae	Bupleurum	fruticosum	2500
14	Asteraceae	Artemisia	vulgaris	1000
		Chrysanthemum	sp	3500
		Helianthus	annuus	3100
		Michenia	micrantha	1400
15	Boraginaceae	Cynoglossum	reticum	5400

	1			
16	Serophulariaceae	Antirrhinum	majus	300
17	Polygonaceae	Fagopyrum	esculentum	1900
18	Euphorbiaceae	Mercurialis	sp	200
19	Urticaceae	Urtica	dioica	800
20	Juglandaceae	Juglans	sp	400
21	Salicaceae	Populus	triandra	
	Folygonscon	Salix	tricocarpa	1900
22	Typhaceae	Typha	angustifolia	600
23	Musaceae	Musa	paradisiaca	500
24	Iridaceae	Crocosmia	sp	1100
25	Liliaceae	Allium	sphaerocephalum	500

Table – 6. Showing the names of families, genera, species and the number of respective pollen present in the summer honey sample of Khensa village per 10 gms

Sl.No.	Family	Genera	Species	No.of pollen grains
1	Ranunculaceae	Ranunculus	bulosus	800
2	Brassicaceae	Diplotaxis	erocoides	20
3	Malvaceae	Habiscus	rosa-sinensis	2400
4	Tiliaceae	Tilla	petiolaris	500
5	Geraniaceae	Geranium	rotundifolium	2300
6	Hippocastanaceae	Aesculus	hippocastanum	300
7	Platanaceae	Platanus	orientalis	700
8	Fabaceae	Glycine	max	100
	H BISQUEINGC. III	Phaseolus	coccineus	600
	Meliaceat	Robinia	pseudacacia	300

9	Caesalipendula	Cassia	didimobotrya	200
10	Rosaceae	Filipendula	ulmaria	700
11	Myrtaceae	Eucalyptus	gunii	100
12	Cucubitaceae	Cucumis	sativus	300
13	Apiaceae	Bupleurum	fruticosum	400
14	Asteraceae	Chrysanthemum	sp	400
15	Polygonaceae	Fagopyrum	esculentum	600
16	Santalaceae	Thesium	humile	800
17	Euphorbiaceae	Mercurialis	perennis	100
18	Corylaceae	Corylus	avellena	700
19	Fagaceae	Facus	sylvatica	300
20	Typhaceae	Thypha	angustifolia	200
21	Musaceae	Musa	paradisiaca	100
22	Iridaceae	Iris	pseudoacorus	400
23	Liliaceae	Allium	sphaerocephalum	2600
24	Poaceae	Alopecurus	pratensis	200
		Zea	mays	100

Table - 7. Showing the names of families, genera, species and the number of respective pollen present in the summer honey sample of market per 10 gms

Sl.No.	Family	Genera	Species	No.of pollen grains
1	Brassicaceae	Brassica	napus	100
2	Capparidaceae	Capparis	spinosa	300
3	Violaceae	Viola	tricolour	200
4	Balsaminaceae	Impatiens	glandulifera	100
5	Meliaceae	Melia	azedarach	100
6	Fabaceae	Glycine	max	100

		Melilotus	albameticus	300
		Ononis	pubescens	100
		Trifolium	pratense	300
7	Rosaceae	Prunus	domestica	100
8	Cucurbitaceae	Cucumis	sativus	100
9	Apiaceae	Bupleurum	fruticosum	700
10	Boraginaceae	Alkanna	tinctoria	100
		Echium	italicum	200
11	Convolvulaceae	Convolvulus	cantabrica	200
12	Lauraceae	Laurus	nobilis	200
13	Fagaceae	Castanea	sativa	800
14	Liliaceae	Aspauagus	acutifolius	100

SUMMARY

Qualitative analysis of honey samples.

(Table - 1 - 10)

As shown in table - 1, Honey same

villages of the Mokokchung district in Nagaland state extrant

village the winter-honey contains pollen grains of 20 per

conifers (rýmnosporms). Whereas, Kubza valtage termine is

polen grains of \$5 genera teleproins to 45 feetles to

the Linema village shows the redesition recolors

SUMMARY

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seem in Kubes sample two, The hosey samples onlicited mos-

during August And September reveal that the potten graces of 14 c

present in the summer sample of Uniting village also polled in

families (gynnospennspollen are absent).

## **SUMMARY**

# Qualitative analysis of honey samples

(Table - 1 - 10)

#### Winter Sample

As shown in table – 1, Honey samples collected from three villages of the Mokokchung district in Nagaland state exhibit that in Khensa village the winter honey contains pollen grains of 86 genera belonging to 44 families of dicots angiosperms, 5 families of monocots angiosperms and one family of conifers (gymnosperms). Whereas, Kubza village winter honey sample shows pollen grains of 85 genera belonging to 45 families of dicot angiosperms, 5 damilies of monocot angiosperms and two families of conifers (gymnosperms). But the Ungma village shows the maximum number of genera i.e., 101 genera belonging to 49 families of dicot angiosperms, one family of monocot angiosperms and one family of conifers (gymnosperms).

#### Summer Samples

As shown in table – 2, the honey samples collected from Khensa village during summer season i.e., August – September shows pollen grains of 27 genera belonging to 19 families of dicot angiosperms and 5 families of monocot angiosperms but gymnosperms pollen grains were absent. Whereas, Kubza village honey samples exhibit pollen grains of 37 genera belonging to 23 families of dicot angiosperms and 3 families of monocot angiosperms. The gymnospermous are absent in Kubza sample too. The honey samples collected from Ungma village during August And September reveal that the pollen grains of 34 genera belonging to 21 families of dicot angiosperms and 4 families of monocot angiosperms are present. In the summer sample of Ungma village also pollen grains of conifer families (gymnospermspollen are absent).

Table - 8. Showing the number of genera and families whose pollens are present in winter honey sample of investigated localities during 2002 - 2004.

SI. Name of locality	No. of genera	enera	No. of families	nilies	No. of genera	No. of families	
	Monocot	dicot	Monocot	dicot	Gymnosperm	Gymnosperm	_
Khensa village	5	92	5	44	1	1	
Kubza village	9	77	2	45	2	2	
Ungma village	4	96	2	49	-	1	
	Khensa village Kubza village Ungma village		Monocot 5 6 4	Monocot dicot 5 76 6 77 4 96	Monocot dicot Monocot 5 76 5 5 6 77 5 4 96 2	Monocot         dicot         Monocot         dicot           5         76         5         44           6         77         5         45           4         96         2         49	Monocot         dicot         Monocot         dicot         Gymnosperm           5         76         5         44         1           6         77         5         45         2           4         96         2         49         1

**Table – 9.** Showing the number of genera and families whose pollens are present in summer honey sample of investigated localities during 2002 – 2004.

## Quantitative analysis of honey samples

## Winter honey samples

The quantitative analysis shows the presence of total number of 44620 pollen/10 gm of honey, collected during winter season in Khensa village. Out of these 40940 pollens belong to dicot angiospermous taxa, 3670 belong to monocot angiospermous taxa, whereas, only 10 pollens belong to gymnospermous taxa.

The winter honey sample of Kubza village exhibit the presence of 31013 pollen grains, out of which 660 belongs to monocots angiospermous taxa, 30323 belongs to dicot angospermous taxa and 30 pollen belongs to gymnospermous (conifers) taxa per 10 gm of honey.

The winter honey samples collected from Ungma village exhibit a total number 60420 pollen per 10 gm of honey, out of which 57280 belong to dicot angiospermous taxa, 3130 pollen belong to monocot angiospermous taxa and only 10 pollen grains belong to gymnospermous taxa.

## Summer honey samples

The quantitative analysis of honey sample collected summer season (August – September, 2002 – 2004) from Khensa village exhibit a total number of 16400 pollens per 10 gm. Out of these 12800 belong to dicot angiospermous taxa, 3600 monocot angiospermous taxa and no pollen of gymnospermous taxa.

The summer honey sample collected from Kubza village shows the presence of 25700 pollen per 10 gm. Out of which 2500 belongs to dicot angiospermous taxa, 700 monocot angiospermous taxa and no pollen of gymnospermous taxa.

The quantitative analysis of honey samples collected from Ungma village exhibit a total number 37500 pollens per 10 gm. Out of which 34900 pollens belong to dicot angiospermous taxa, 2600 pollen to monocot angiospermous taxa but gymnospermous pollen were absent.

Table - 10. Showing the number of pollen grains per 10 gm of honey sample collected during winter and summer from various localities of investigation during the year 2002 - 2003 and 2003 - 2004.

		Total	number of taxa			16400	25700	37500
SUMMER SAMPLE	No. of pollen	to	gymnospermous	t l	ket	Nil	Nii	Nil
	No. of	belonging	to monocot	angiospermous	taxa	3600	200	2600
	No. of	belonging	to Dicot	angiospermous	taxa	12800	25000	34900
	Total number of taxa					44620	31013	60420
	No. of pollen	to	gymnospermous taxa			10	30	10
E	No. of	belonging	to	angiospermous	taxa	3670	099	3130
WINTER SAMPLE	No. of	to belonging	Sl. no Name of angiospermous	taxa	ictes	40940	30323	57280
	Name of locality					Khensa Village	Kubza Village	Ungma
SI. no						1.	5.	3.

## Market sample

## Qualitative analysis of market honey sample

Qualitative melissopalynological analysis of market honey samples collected from Mokokchung daily market exhibit pollens of 18 genera belonging 13 families of dicot angiosperms. However, the gymnosperms pollen grains were absent in market honey samples.

## Quantitative analysis of market honey sample

Quantitative melissopalynological analysis of market honey sample collected from the Mokokchung daily market reveals the presence of 4000 pollen grains per 10 gm of honey. Out of which 3900 pollen belong to dicot angiospermous taxa and only 100 pollen grains belong to a single family viz., Liliaceae and single genus *Asparagus*.

## Poisonous/Allergic pollen grains in various honey samples

Repeated observation of winter and summer honey samples of various localities reveal the presence of few allergic pollens, viz., *Xanthium*, *Strumarium*, *Alnus glutinosa*, *Betula pendula*, *Artamisia sp*. And others. These pollen grains when eaten by the cattles in bulk along with the respective plants, causes poisoning in them (www.allergenica.com/greer. asp; www.labspec.co.za/l weed. htm).

# RESULTS

# AND

# **DISCUSSION**

# Results and Discussion

Melissopalynological investigaton of three localities, viz., Khensa village, Ungma village and Kubza village of Mokokchung district during the winter months (January – February) and summer months (August – September) for two years i.e., 2002 -2003 and 2003 – 2004 reveal that the honey samples collected during winter season contain maximum number of pollen contents. The number of pollen ranges from minimum 31013 pollens per 10 gm in Kubza village honey followed by Khensa village honey samples i.e., 44620 pollen per 10 gm and the maximum number of pollen i.e., 60420 pollen per 10 gm was found in the Ungma village honey samples. Whereas, summer samples of honey in all the three localities exhibit rather low frequency of pollen grains. E.g., summer honey samples from Khensa village shows minimum number of pollen grains i.e., 16400 per 10gm, but the summer honey samples from Kubza village shows 25700 pollens per 10 gm. Whereas, the honey samples from Ungma village shows 37500 pollens per 10 gm. of honey which is the maximum number of pollens present per 10 gm in the investigated samples.

Therefore, on the basis of present investigation and the results, it can be concluded that out of three localities (Khensa, Kubza and Ungma), the Ungma village and its surroundings possesses maximum diversity of plants which contribute as pollen flora (table - 9) to the honey bees of this locality. These results are also found correct by qualitative analysis of honey samples of these localities. According to the qualitative analysis of winter samples of honey from these three localities it is evident that in Ungma village pollen contributot taxa belongs to 100 genera of 52 families. Out of this 96 genera belongs to the 49 families of dicot angiosperms (table – 7).

However, the contributor plants are rather less in number e.g., Ungma village summer honey sample exhibit 34 genera belonging to 25 families of angiospermous taxa but the pollen grains of gymnospermous taxa are absent. Out of these, the number of dicot families and genera is much more in comparision to monocot. i.e., 30 genera belonging to 21 families of dicot angiospermous. Whereas only 4 genera of 4 families of monocot angiospermous are there.

The maximum number of families and genera contributing the pollen flora in summer honey sample was recorded in Khensa village i.e., 26 genera belonging to 24 families of angiospemous. Out of these 19 families with 21 genera of dicots and 5 families with 6 genera of monocot represent the pollen flora. In summer sample not even a single pollen grain of gymnospermous taxa has been observed (table -7).

On the basis of these qualitative and quantitative analysis, it is concluded that the maximum number of pollen grains in the honey in Mokokchung district during winter as well as summer comes from the taxa belonging to dicotyledonous angiosperms and the number of pollen from monocotyledonous angiosperms is rather less. The presence of pollen grains of gymnospermous taxa is very less and hence can be considered as contamination because during January – February the pollen grains of *Pinus sylvestris* and *Cupressus sp* are very frequent in the air. The presence of a few fungal spores and hyphae in honey sample is also found as contamination.

## Comparison of results with that of the Market samples

The seasonal (winter and summer) qualitative and quantitative analysis of honey samples collected from Khensa, Kubza and Ungma villages of Mokokchung district shows a high frequency of pollen grains I.e., ranging from – 31013 to 60420 per 10 gm in summer samples with a wide range of families (50 – 52) and genera (82 – 101) belonging to angiosperms as well as gymnosperms. But the qualitative and quantitative analysis of market sample exhibit pollen flora belonging only 17 genera of 13 dicot families and 1 genus of one monocot family. However, the pollen grains of gymnospermous plant are not present in market sample. The total number of pollen grains in market sample in comparison to that of natural bee hive samples from investigated localities, also under represented i.e., only 4000 pollen grains per 10 gm of honey. These comparison of pollen frequency per 10 gm of honey sample between natural honey sample collected by bees and market sample, clearly indicate adulteration of market honey in Mokokchung town.

## Colour of the honey

In the light of present investigation (quantitative as well as qualitative) it has been concluded that the colour of winter honey is always dark brown and the summer honey is always light brown. This may be interpreted as due to the presence of pollen grains during winter and lesser number i.e., almost half of the number of that of the winter sample. But when examine the market honey colour, it is always dark brown, whereas, the number of pollen grains per 10 gm is nearly 12 to 15 times lesser than winter honey sample. These result also indicate that market samples are not pure.

## Unifloral and multifloral honey

Winter sample honey of Khensa and Ungma villages exclusively shows the presence of Asteraceae pollen grains as maximum e.g., Khensa village shows 50.87%, Kubza shows 70.06% and Ungma shows 49.04%. Therefore, the winter honey can be termed as Asteraceous honey (here the single floral honey can not be given the name), whereas, the summer honey cannot be categorized in any category as the presence of pollen grains in Ungma sample in Asteraceous 24% as maximum. The Kubza sample exhibit Asteraceous pollen 29.57% which is maximum and Khensa sample shows Malvaceous pollen 14.63% followed by Geranceaceous pollen 14.02% and Liliaceous pollen as 15.85%. Hence, the summer honey samples of all the three villages may be categorized as multifloralhoney.

#### REFERENCES

A lon, M.Y. 1928 h. Pellen Grains. Ber World 9: 86.

Attenue, E.O. 1936, Zam. Policiner, remain des la neverselles

REFERENCES

46 (1925)

Annual VIII. and Jones G.D. (500). The

Reput Jr. Vaughn M. 2301, Police Control of Control

toron il M-1973, An Analysis of Horsey Bes Police !

Change of Autumn House .

M. 1889, Police Analysis of Some Spring House & a

## REFERENCES

Allen, M. Y. 1928 a, Armchair Thoughts. Bee world 9:57.

Allen, M.Y. 1928 b, Pollen Grains. Bee World 9:66.

Allen, M.Y. 1929, Pollen Grains IV. Bee World 10: 114-118.

Alfonsus, E.O. 1933, Zum Pollenverbrauch des Bienenvolkes. Archiv Für Bienenkunde 14:220-223.

Armbruster, L. 1921, Vergleichende Eichungsversuche auf Bienenkund Wespen. Archiv für Bienenkunde 3: 219 – 230.

Barth, O.M., 2004, Melissopalynology in Brazil: A Review of Pollen Analysis of Honeys, Propolis and Pollen Loads of Bees. Sci. Agric. (Piracicaba, Braz.), 61 (3): 342 – 350.

Betts, A.D.1923, The Identification of Pollen Sources. Bee World 5:43
- 45. (1925)

Betts, A.D. 1925, notes on Pollen Identification. *Bee World* 7:90.

Bryant V.M. and Jones G.D. 2001, The R-Values of Honey: Pollen Coefficients, *Palynology* 25, 11-28.

Bryant Jr. Vaughn M. 2001, Pollen Contents of Honey. CAP Newsletter 24(1): 10-24.

Chaturvedi, M. 1973, An Analysis of Honey Bee Pollen Loads from Banthra, Lucknow, India. *Grana*, 13: 139 – 144.

Chaturvedi, M. 1983, Pollen Analysis of Autumn Honeys of Kumaon Region. Pcoc. Indian Natn. Sci. Acad, B49: 125 – 133.

Chaturvedi, M. 1989, Pollen Analysis of Some Spring Honeys from the Western Himalayan Region of Uttar Pradesh, India. *Proc. Indian* 

Acad. Sci. (Plant Sci.), 99 (3): 241 - 246.

Chaturvedi, S.K. 1983, Studies in Pollination of Some Indian Plants.

D.Phil Thesis. Department of Botany Allahabad University, Allahabad.

Chaturvedi, S.K.1993, Studies on Insect Visitors/Pollinators of Some angiospermous Plants in Allahabad. New Approaches in Agricultural Technology. 2:301-334.

Chaturvedi, S.K., and Shonali Chaturvedi., 2001, Biology of Reproduction in Angiosperms. Bioved Research Society, Allahabad. PP: 1-130.

Chaturvedi, S.K., and Temsunungla., 2004, Melissopalynology in Ungma village of Mokokchung District in Nagaland. Nagaland University

Research Journal, 2:95-99.

Chaubal, P.D. and Deodikar, G.B. 1963, Grana Palynol. 4:3, 393 - 397.

Chaubal.P.D. and Deodikar, G.B. 1965, Morphological characterization of pollen grains of some major honey yielding plants of the Western ghats (India). *India Bee Jorn.* XXVII: I, 1-28.

Chaubal, P.D. and Deodikar, G.B. 1965, Palynological Bulletin, I: 56 - 58.

Chaubal, P.D. 1966, Palynological Studies on the family Acanthaceae

from Western Ghats. Registrar, Poona University, Poona (India).

Chaubal, P.D. and Deadikar, G.B. 1966 – 1967, Palynological Bulletin II – III, 84 – 88.

Chaubal, P.D. 1976, Palynological Aspects of Compositae. Shivaji University, Kolhapur (India).

Chaubal, P.D. 1980, Melittopalynological Studies of Sagarmal Area (Kolhapur), Maharashtra, India, Dring Monsoon, PP: 103 – 107. In Nair,

P.K.K (Edt) Advances in pollen Spore Research. Vol. V - VIII, Today and Tomorrow Printers and Publishers, New Delhi (India).

Cooke, T.1967, The flora of the Presidency of Bombay. I – II, 2<sup>nd</sup> Edition, Botanical Survey of India, Calcutta.

Deodikar, G.B., Shah Chaubal, P., Thakar, C.V. and Salvi, S.R. (1958)

Proc. 17th Int. Bee Keeping Congr. Balagna - Rome. 2: 214 - 217.

Deodikar, G.B., Thakar, C.V., Phadke, R.P., Shah Chaubal P. (1958 B) . *Bee World*, 39 (5),118–120.

Deodikar, G.B., Thakar, C.V., Shah (Chaubal), P., Salvi, S.R., Chitale, P.S. (1958 C), Bee World, 39 (5): 12 – 121.

Demianowicz, Z. 1961, Pollenkoeffizienten als Grundlage der quantitativen Pollenanalysis des Honigs. Pszczelnicze Zeszyty Naukowe 5 (2): 95 - 105.

Demianowicz, Z. 1964, Characteristik der Einartenhonige. Annales de l'Abeille 7: 273 – 228.

Dietz, A. 1975, Nutrition of the Adult Honeybee. In Dadant, C., and Dadant, C.P., (eds): *The Hive and the Honeybee*, Dadant & Sons, Carthage, Illinois, pp 125-126.

Eckert, J. E. 1942, The Pollen Required by a Colony of Honeybees.

Journal of Economic Entomology, 35: 309-311.

Erdtman, G. 1952, Pollen Morphology and Plant Taxonomy Angiosperms.

Chronica Botanica Co., Waltman, Mass, USA.

Erdtman, G. 1960, The Acetalysis Method. Svensk Botanisk Tidskrift, 54: 561-564.

Fehlman, 1911, In Bryant. Jr, M.V. 2001. Pollen Contents of Honey. CAP

News letter 24(1):10-24.

Garg, A. 1996, Palynocontents of Bee - Collected Pollen Loads of Autumn Season in Bhimtal, India. *Taiwania*, 49 (3): 197 - 207.

Garg, A. And Nair. P.K.K. 1994, Seasonal Analysis of Honey Bee Pollen Loads from a Temperate Zone in Kumaon Himalaya in Uttar Pradesh, India. J. Palynol, 30:1-34.

Gary, N.E. 1975, Activities and Behavior of Honeybees. In Dadant, C., and Dadant, C.P., eds.: *The Hive and the Honey Bee*, Dadant & Sons, Carthage, Illionois, P. 185 – 264.

Herrero, B., Barrera, R.M.V., Martin, R.S., Pando, V., 2002, Characterization of Honey by Melissopalynology and Statistical Analysis. *Can. J. Plant Sci.* 82:75 – 82.

Jones, G.D., Bryant, V.M. Jr., Lieux, M.H. Jones, S.D. and Lingren, P.D.

1995, Pollen of the Southeastern United States with Emphasis on

Melissopalynology and Entomopalynology, CAP Newsletter 18 (3): 22-23.

Lieux, M. H. 1969, A Palynological Investigation of Louisiana Honeys.

Unpubl. Doctoral Dissertation, Department of Botany and Plant

Pathology, Lousiana State University, U.S.A., 113 pp.

Lieux, M. H. 1975, Dominant Pollen Types Recovered from Commercial Louisiana Honeys. *Economic Botany* 29: 78 – 96.

Lieux, M. H.1978 Minor Honeybee Plants of Louisiana Indicated by

Pollen Analysis. Economic Botany 32: 418-432.

Louveaux, J., Maurizio, A. and Vorwohl, G. 1970, Methods of

Melissopalynology, Bee World 51, 125 - 131.

Louveaux, J. Maurizio, A. Vorwohl, G. 1978, Methods of

Melissopalynology. Bee World 59: 139-157.

Lutier, P. M., and Vaissiere, B. E. 1993, An Improved Method for Pollen Analysis of Honey. Review of Palaeobotany and Palynology 78: 129 – 144.

Maurizio, A. 1951, Pollen Analysis of Honey. Bee World 32:1-5.

Maurizio, A., Louveaux J., 1965, Pollen de Plantes Melliferes d' Europe,

Union des groupements Apicoles Français, Paris.

Moar, N. T., 1985, Pollen Analysis of New Zealand Honey. New Zealand Journal of Agricultural Research 28:39-70.

Nair, P. K. K. 1964, A Pollen Analytical Study of Indians Honeys. J. Indian Bot. Soc., 18 (2): 179 – 191.

Oertel, E. 1939, Honey and Pollen Plants of the United States. U. S. Department of Agriculture; Circular 554, Washington, D. C.

Ohe, W.V.D., Oddo, L.P., Piana, M.L., Morlot, M., and Martin, P., 2004,

Harmonized Method of Melissopalynology. Apidologie 35: S 18 - S 25.

Pammel, L. H., and King, C. M. (eds) 1930, Honey Plants of Iowa;

Iowa Geological Survey Bulletin 7. States of Iowa, Des Moines, Iowa.

Parker, R. L., 1923, Some Pollen gathered by Bees. American Bee Journal 63:16-19.

Pellett, F. C., 1930, American Honey Plants, together with those which are of Special value to the Beekeeper as Sources of Pollen. (3<sup>rd</sup> ed.)

American Bee Journal, Hamilton, Illinois.

Pfister, R., 1895, Versuch Einer Mikroskopie des Honigs. Forschber.

Lebensmitt, U. Ihre Bez. Z. Hygiene, Forens. Chem., Pharmakogn. (2) (1/2):

1 - 9; 29 - 35.

Ricciardelli, d, albore, G. (ed) 1997, Textbook of Melissopalynology, Apimondia, Bucharest, 300 pp.

Ricciardelli, d, albore G. 1998, Mediterranean Melissopalynology,
Istituto di Entomologia Agraria, Universita Degli Studi, Di Perugia,
466 pp.

Sawyer, R., 1981, Pollen Identification for Bee Keepers. Cardiff Academic Press, Cardiff, Wales, 112 pp.

Sawyer R., 1988, Honey Identification, Cardiff Academic Press, Cardiff, Wales, UK.

Seedly, T. D., 1985, <u>Honeybee Ecology</u>. Princeton University Press, Princeton, New Jersey.

Sharma, M., 1970, An Analysis of Pollen Loads of Honey Bees from kangra, India. *Grana*, 10:35-42.

Snodgrass, R. E. and Erickson, E. H. 1992, The Anatomy of the Honeybee. In: Dadant C and Dadant C.P. (eds.) The Hive and the Honey Bee, Dadant R Sons, Hamilton, Carthage, Illinois, P. 103 – 169.

Teale, E.W. 1942, The Golden Throng: A Book about Bees. Robert Hale Ltd., London.

Todd, F. E., and Vansell, G. H., 1942, Pollen Grains in Nectar and Honey. The Journal of Economic Entomology 35: 728-731.

Van Der Ham, R. W.J. M., Kaas, J. P., Kerkvliet, J. D. and Neve, A. 1999, Pollenanalyse. Stichting Landelijk Proefbedrijf Voor Insektenbestuiving en Bijenhouderij Ambrosiushoeve. Hilvarenbeek, Netherlands. 156 pp. Whitcomb, W. Jr., and Wilson, H. F., 1929, Mechanics of Digestion of Pollen by the Adult Honeybee and the Relation of Undigested Parts to Dysentery of Bees. Wisconsin Agricultural Experiment Station Research Bulletin, 92.

Winston, M. L., 1987, The Biology of the Honey Bee. Harvard University Press, London.

Young, W. J., 1908, A Microscopical Study of Honey Pollen. <u>United</u>

<u>States\_Bureu of Chemistry, Bulletin, 110</u>, Washington, D.C. 93pp.

Zander, E. 1935 – Pollengestaltung und Herkunftsbestimmung bei Blütenhonig. Reichsfachgruppe Imker, Berlin.

Zander, E. 1937 – Pollengestaltung und Herkunftsbestimmung bei Blütenhonig. Liedloff, Loth & Michaelis.

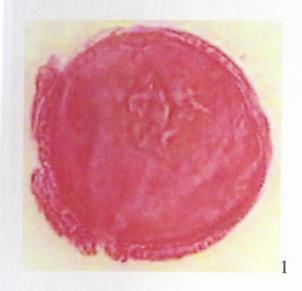
Zander, E. 1941 – Pollengestaltung und Herkunftsbestimmung bei Blütenhonig. Liedloff, Loth, Michaelis.

Zander, E. 1949 – Studien Zur Heskunftsbestimmung bei Waldhonigen. Ehrenwirth, Munich.

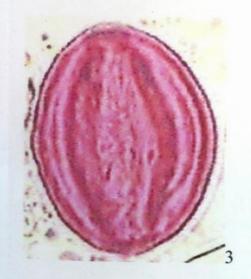
Zender, E. 1951 – Letzte Nachtrage Zur pollengestaltung und Herkunftsbestimmung bei Blütenhonig. Liedloff, Loth & Michaelis, Leipzig.

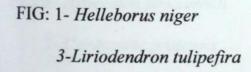
# PHOTOMICROGRAPH PLATES

# WINTER SAMPLE:....











4-Magnolia grandiflora

2- Clematia vitalba







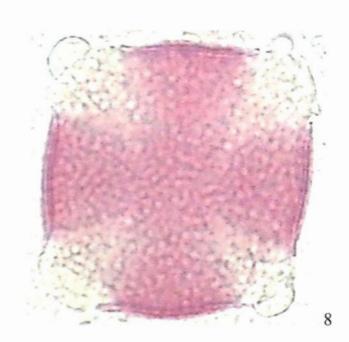
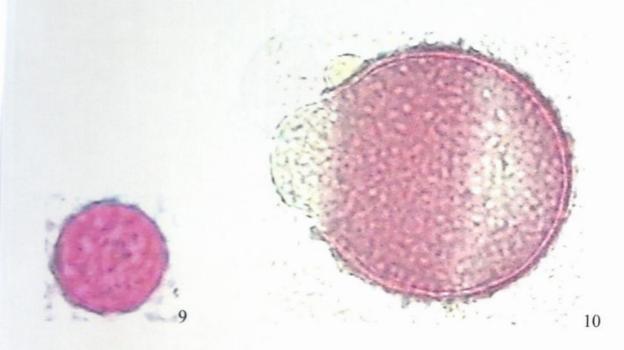


FIG:5- Hypecoum procumben 6 - Brassica napus 7- Cardamine pratensis

8- Viola tricolor



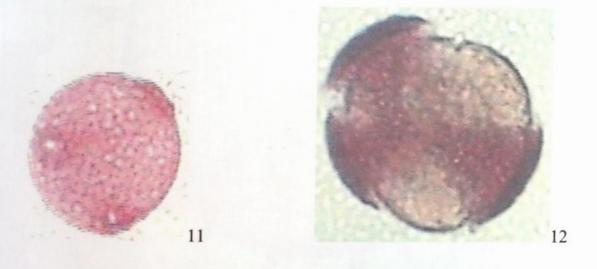
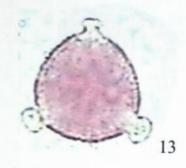


FIG: 9- Lychnis flos-cuculi 10- Spercularia rupina
11- Stellaria media 12- Tamarix gallica



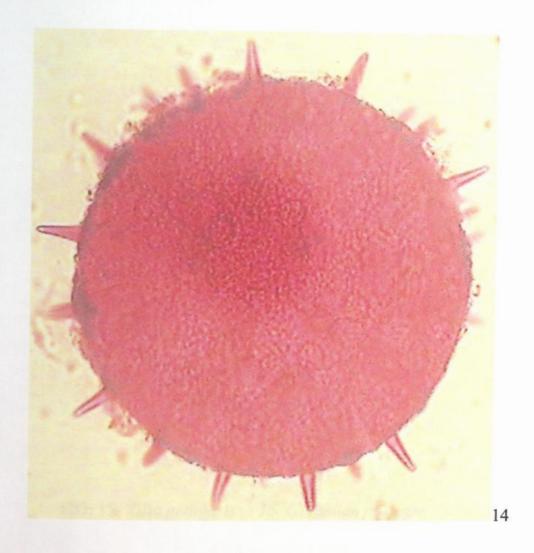


FIG: 13- Helianthemum chamaecistus 14-Hibiscus rosa-sinensis



15

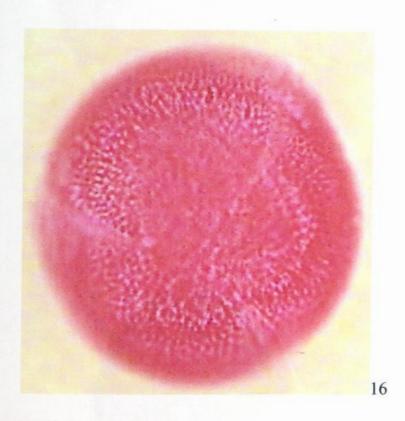


FIG: 15- Tilia petiolaris 16- Geranium pratense

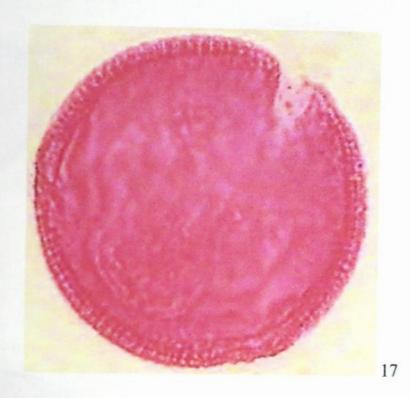




FIG: 17- Geranium rotundifolium 18- Oxalis articulata



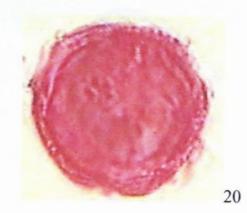
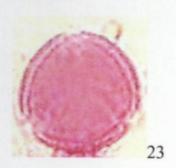






FIG: 19- Impatiens glandulifera 20- Citrus limon 21- Parthenocissus tricuspidata 22- Vitis vinifera





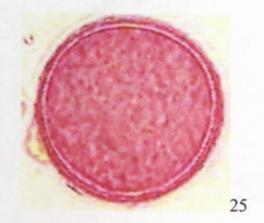




FIG: 23-Aesculus hippocastanum 24- Acer pseudoplatanus 25- Pistacia lentiscus 26- Platanus orientalis

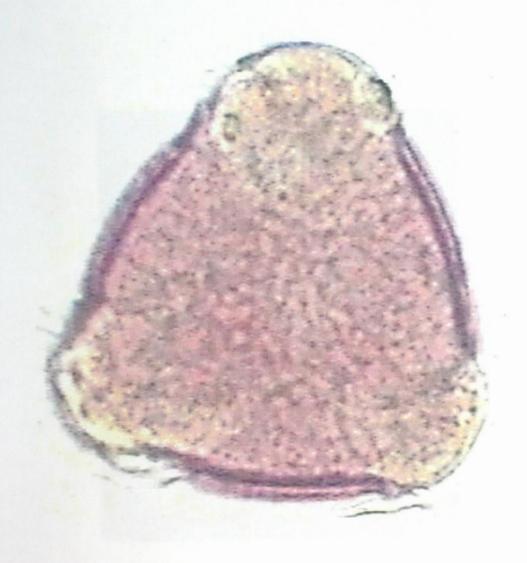


FIG: 27- Coronilla emerus

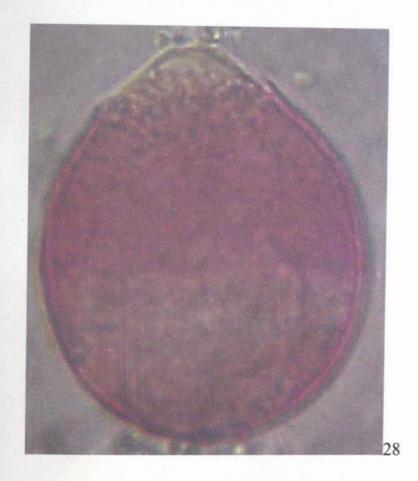
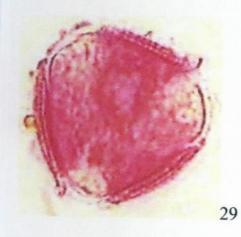
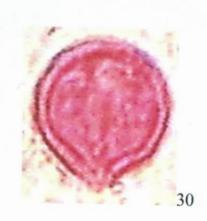


FIG: 28- Calycotome spinosa

## Plate - 11





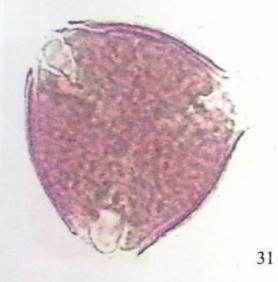
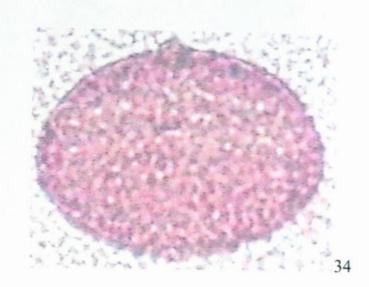




FIG: 29- Gleditsa triancanthos
31- Lupinus albus

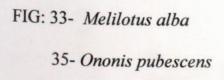
30- Glycine max32- Medicago sativa

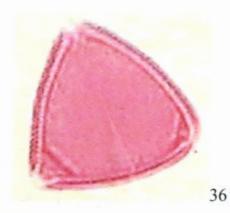












34- Onobrychis vicifolia 36- Robinia pseudoacacia









FIG:37- Sophora japonica
39- Vicia faba

38- Ulex galli40- Cassia didibotrya

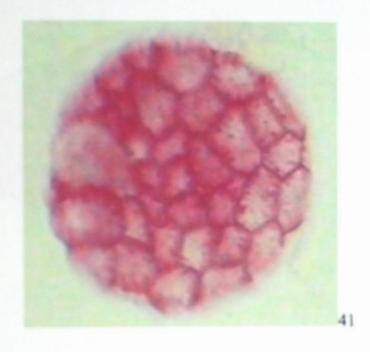


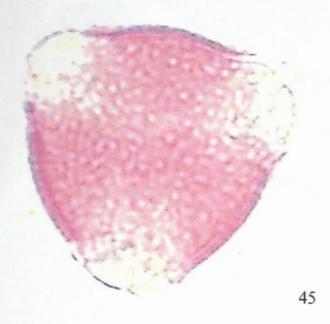


FIG:41- Acacia dealbata 42- Crataegus monogyna





FIG: 43-44- Malus domestica



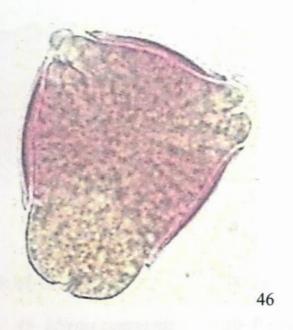
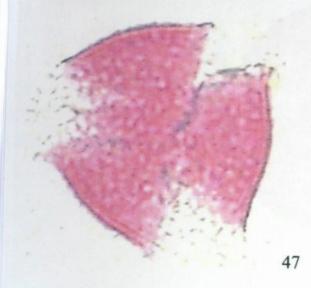
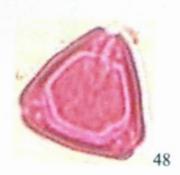


FIG: 45- Prunus dulcis 46- Pyrus communis







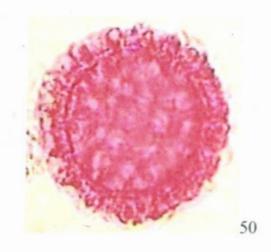


FIG: 47- Sedum acre
49- Myrtus communis

48- Eucalyptus gunni 50- Passiflora edulis



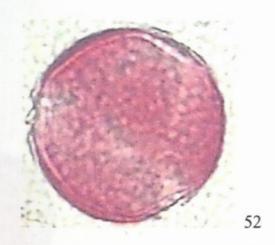


FIG: 51- Bryonia dioica

52- Citrullus lanatus







FIG: 53 -Ecballium elaterium

55- Bupleurum fruticosum

54- Cucubita pepo

56-Smyrnium olusatrum







59

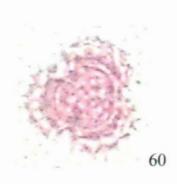
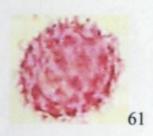


FIG: 57- Hedera helix

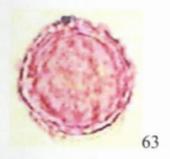
58- Cornus sanguinea

59 - Ageratum conizoides 60- Arctium sp

## Plate - 21







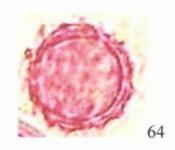
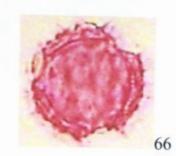
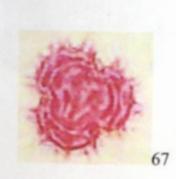


FIG: 61- Aster sp 62-Carduus sp

63- Carthamus sp 64- Circium sp







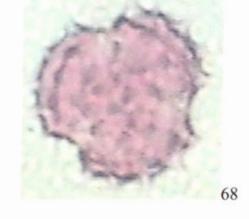


FIG: 65- Helianthus annuus 66- Inula sp
67- Matricaria sp 68- Michenia micrantha



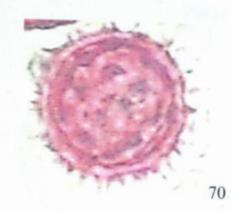




FIG: 69- Solidago canadensis 70- Taraxacun officinale 71- Xanthium strumarium



72- Arbutus unedo









FIG: 73- Erica sp

74- Diospyros kaki

75- Fraxinus excelsior

76- Alkanna tinctoria







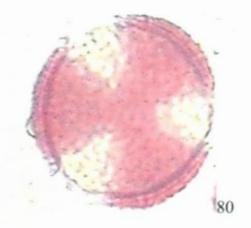


FIG: 77- Cynoglossum craticum 78- Echium italicum

79-Symphytum officinale 80- Convolvulus cantabrica



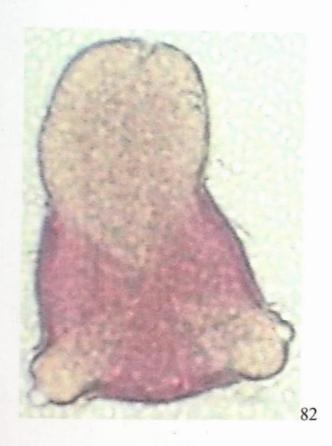
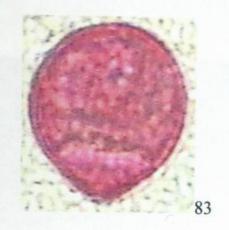


FIG: 81- Antirrhinum majus 82- Verbascum thapsus



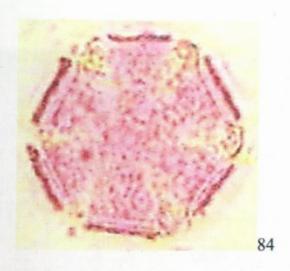
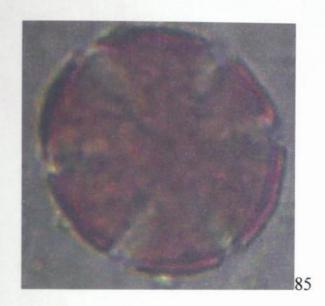


FIG: 83- Verbena officinalis 84 – Lavandula agustifolia



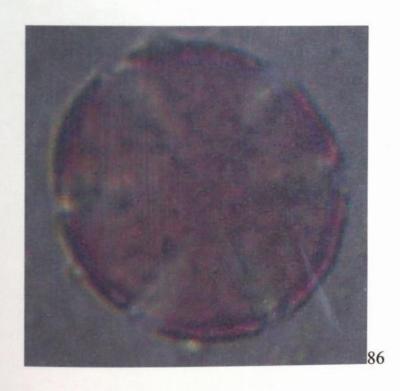
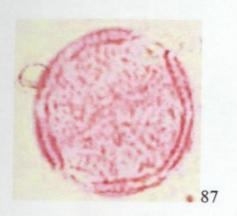
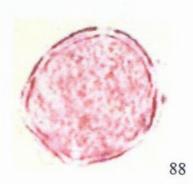


FIG:85 – 86 - Mentha pulegium





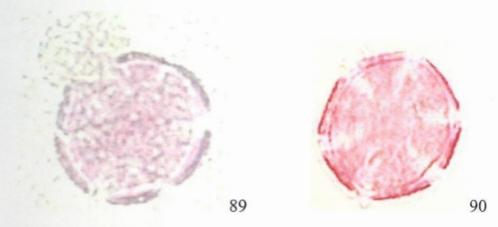


FIG: 87-88-Rosmarinus sp 89-Teucrium sp 90-Thymus sp

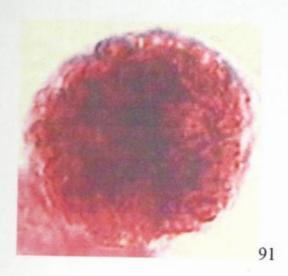
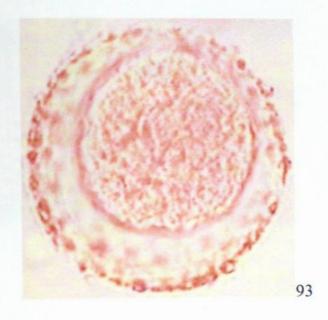




FIG: 91- Chenopodium sp 92- Fagopyrum esculentum



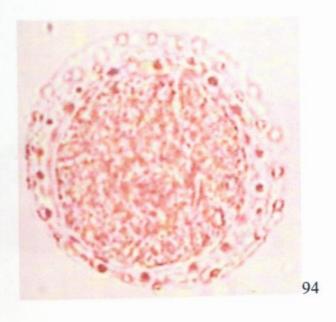
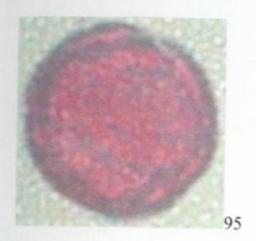
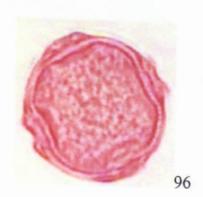
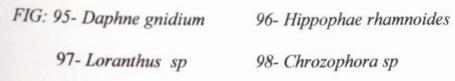


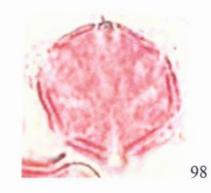
FIG: 93-94- Laurus nobilis











98- Chrozophora sp

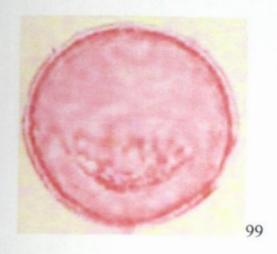
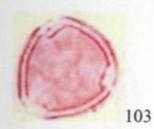
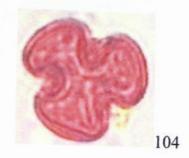


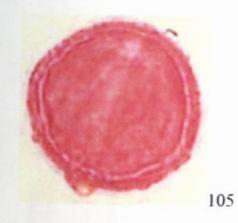




FIG:99- Urtica dioica 100- Ulmus procera
101- Alnus glutinosa 102 - Betula pendula







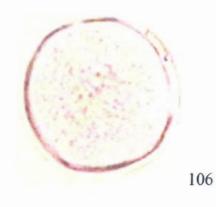
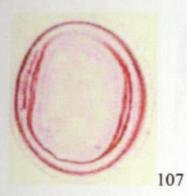
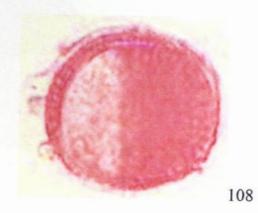


FIG: 103- Castanea sativa

105 - Populus tricocarpa 106 - Musa x paradisiaca

104- Quercus robur







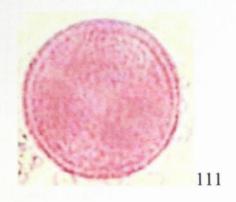


108- Iris unguicularis

109- Allium cepa

FIG: 107- Cronus purpurea

110- Asphodelus microcarpus



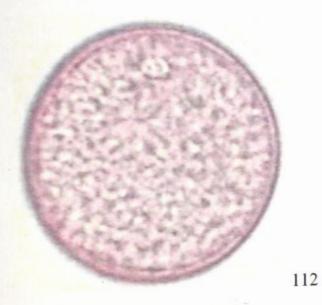


FIG: 111 - Alopecurus pratensis

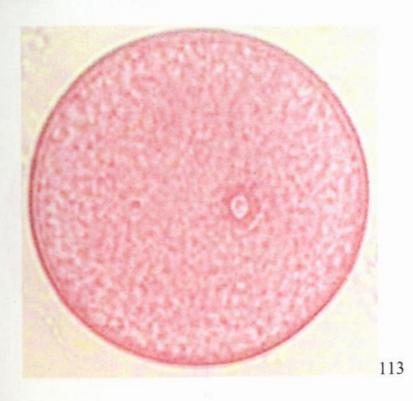


FIG: 112-113- Zea mays







FIG: 114 – 116 - Pinus insularis

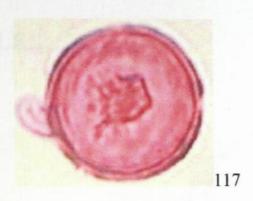


FIG: 117- Cupressus sempeviren

## IMMER SAMPLE:....





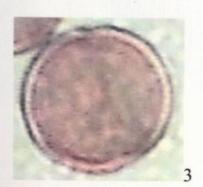


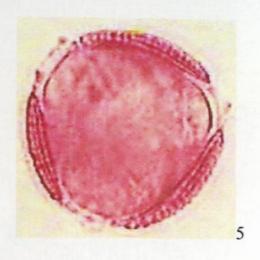


FIG: 1- Clematis vitalba

3- papaver rhoes

2- Liriodendron tulipifera

4- Brassica napus



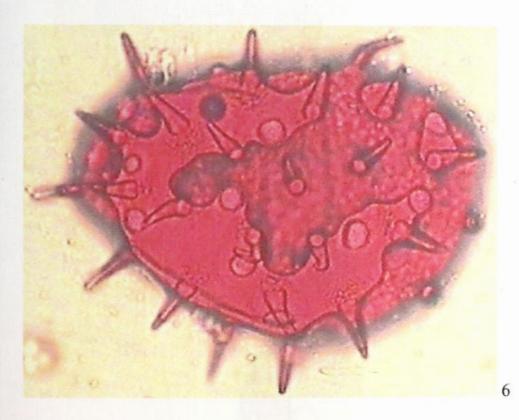


FIG: 5- Diplotaxis erocoides

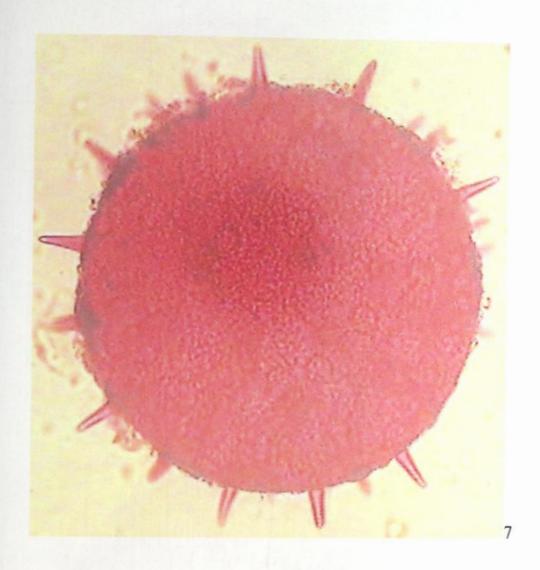
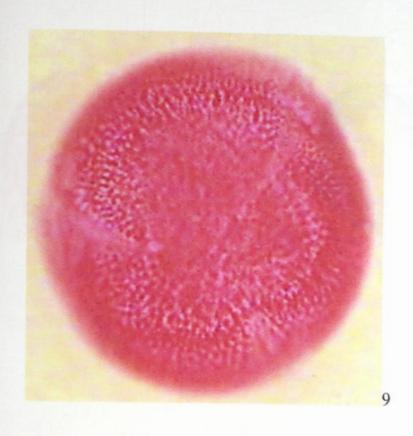




FIG: 6 - 7- Hibiscus rosa - sinensis 8- Tilia petiolaris



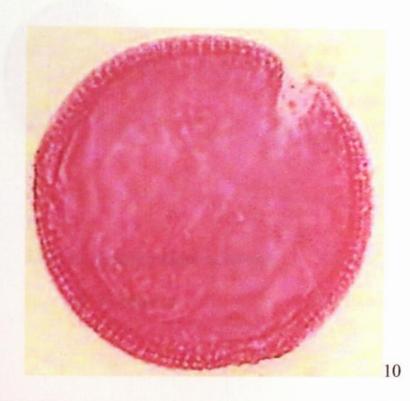
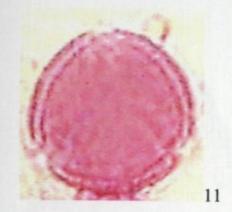


FIG:9- Geranium pratense 10-Geranium rotundifolium





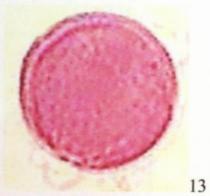


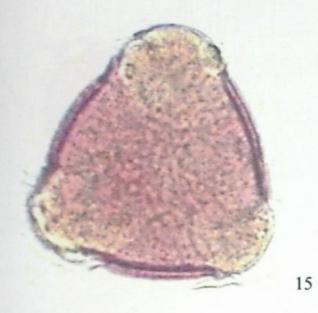


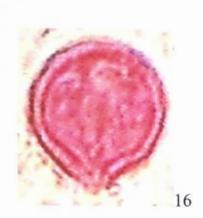
FIG: 11 Aesculus hippocastanum

13- Cotinus sp

12- Acer sp

14- Platanus orientalis





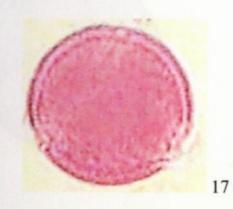


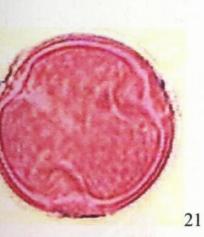


FIG: 15 - Coronilla sp 16 - Glycine max

17- Phaseolus coccineus 18- Robinia pseudoacacia







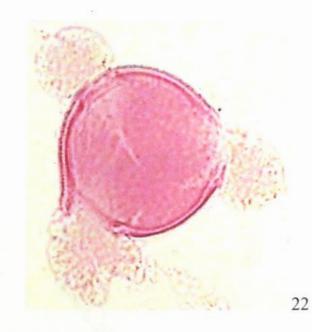
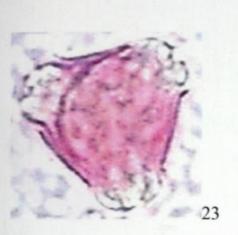


FIG:19- Melilotus alba 21- Cassia didibotrya

20- Sophora japonica22- Filipendula ulmaria





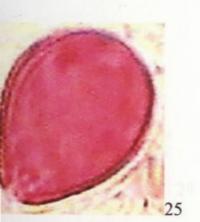




FIG:23- Myrtus communis

24 – cucurbita pepo 25- cucumis sativus 26- citrullus lanatus





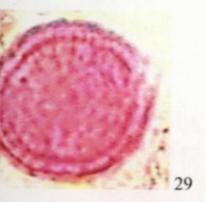
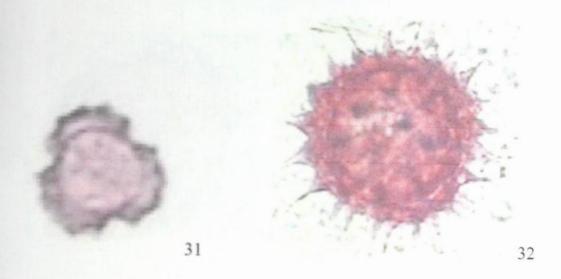




FIG: 27- Bupleurum fruticosum
29- Ambrosia maritima

28-Cornus sanguinea
30- Artemesia vulgaris



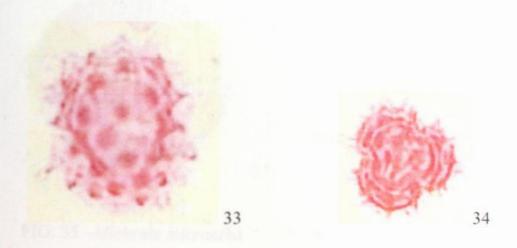
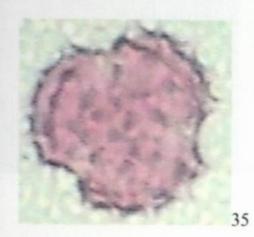


FIG: 31- Chrysanthemum sp 32- Helianthus annuus 33- 34 - Matricaria sp





36



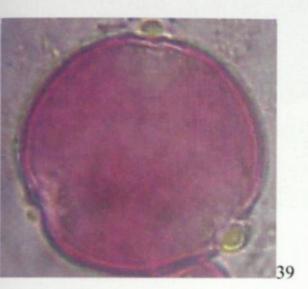


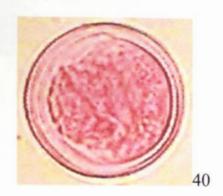
FIG: 35 -Michenia micrantha

37- Fagopyrum esculentum

36- Cynoglossum creticum

38- Loranthus euspaeus







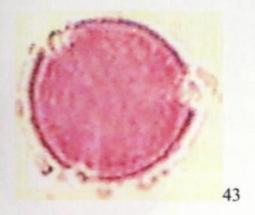
41



42

FIG: 39- Mercurialis sp 41- Juglans sp

40- Urtica dioica 42-Carpinus betulus



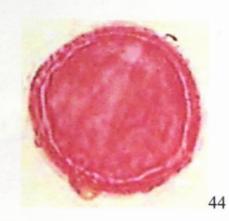




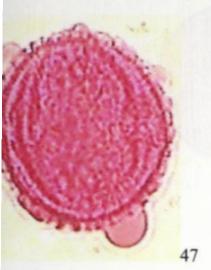


FIG: 43-Facus sylvatica

44- Populus trichocarpa

45- Salix triandra

46- Musa x pradisiaca







49

FIG: 47- Crocosmia sp 48- Galanthus nivalis
49- Allium cepa



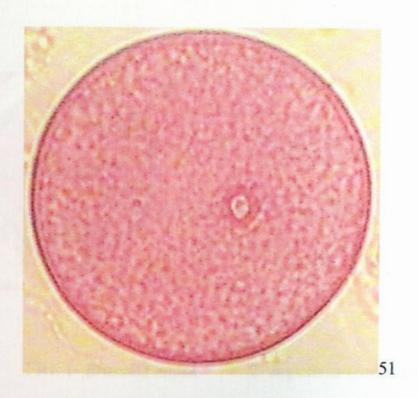
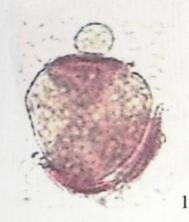
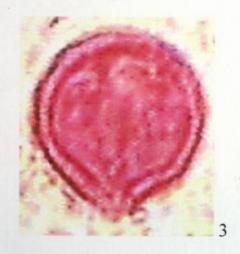


FIG: 50 – Alopecurus pratensis 51 – Zea mays

## MARKET SAMPLE:







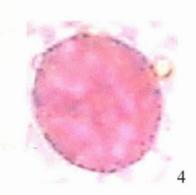


FIG: 1- Brassica napus

3- Glycine max

2- Impatiens glandulifera

4- Melilotus alba

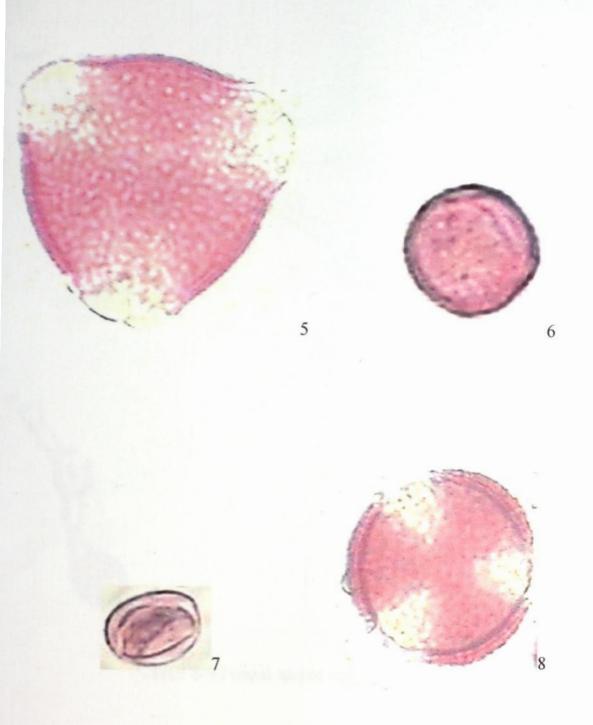


FIG:5- Prunus domestica

7- Alkanna tinctoria

6- Bupleurum fruticosum

8- Convolvulus arvensis



FIG: 8-9 Fungal spores and mycelium

## UNGAL SPORES AND MYCELIUM:.....

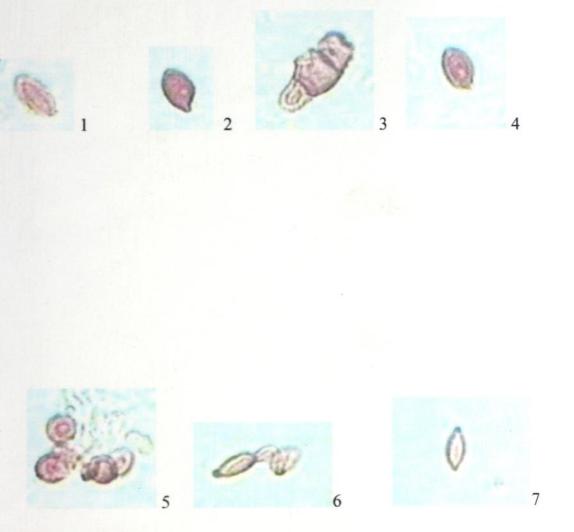


FIG: 1-7-Fungal spores and mycelium

