STUDY ON ACIDITY TOLERANCE OF MAIZE AND ITS RESPONSE TO PHOSPHORUS AND BORON

Thesis

submitted to

NAGALAND UNIVERSITY

in partial fulfillment of requirements for the Degree

of

Doctor of Philosophy

in

Agricultural Chemistry and Soil Science

by

Merasenla Ao Admn. No. Ph-156/14; Regn. No. 760/2017



Department of Agricultural Chemistry and Soil Science,

School of Agricultural Sciences and Rural Development, Nagaland University, Medziphema Campus – 797106 Nagaland 2020

DECLARATION

I, Ms. Merasenla Ao, hereby declare that the subject matter of this thesis is the record of work done by me, that the contents of this thesis did not form the basis of the award of any previous degree to me or to the best of my knowledge to anybody else, and that the thesis had not been submitted by me for any research degree in any other university/institute.

This is being submitted to Nagaland University for the degree of Doctor of Philosophy in Agricultural chemistry and Soil Science

Date: 06/10/2020 Place: MEDZIPHEMA

(MERASENI

<. Sharma Supervisor

NAGALAND UNIVERSITY Medziphema Campus School of Agricultural Sciences and Rural Development Medziphema – 797106, Nagaland

Dr. Y.K. Sharma Professor Department of Agricultural Chemistry and Soil Science

CERTIFICATE – I

This is to certify that the thesis entitled "Study on Acidity Tolerance of Maize and its Response to Phosphorus and Boron" submitted to Nagaland University in partial fulfillment of the requirements for the award of degree of Doctor of Philosophy (Agriculture) in Agricultural Chemistry and Soil Science is the record of research work carried out by Ms. Merasenla Ao, Registration No. 760/2017 under my personal supervision and guidance.

The results of the investigation reported in the thesis have not been submitted for any other degree or diploma. The assistance of all kinds received by the student has been duly acknowledged.

Date : 06/10/2020 Place : Medziphema

(Dr. Y.K. Sharma)

Supervisor

NAGALAND UNIVERSITY Medziphema Campus School of Agricultural Sciences and Rural Development Medziphema – 797106, Nagaland

CERTIFICATE – II

VIVA VOCE ON THESIS OF DOCTOR OF PHILOSOPHY IN AGRICULTURAL CHEMISTRY AND SOIL SCIENCE

This is to certify that the thesis entitled "Study on Acidity Tolerance of Maize and its Response to Phosphorus and Boron" submitted by Miss Merasenla Ao, Admission No. 156/14, Registration No. 760/2017 to the NAGALAND UNIVERSITY in partial fulfillment of the requirements for the award of degree of Doctor of Philosophy in Agricultural Chemistry and Soil Science has been examined by the Advisory Board and External examiner on 22.03.2021.

The performance of the student has been found Satisfactory/Unsatisfactory.

Member

1. Prof. Y.K. Sharma (Supervisor)

2. Prof. R. K. Kumarjit Singh (External examiner)

3. Dean, SASRD (Pro Vice Chancellor Nominee)

Members of Advisory Committee: 1. Prof. S. K. Sharma

2. Prof. A. K. Singh

3. Dr. A. P. Singh

4. Prof. Amod Sharma

Head Dept. of Agricultural Chemistry and Soil Science

Signature Retermine Retermine Signature Retermine Signature Signat

Dean School of Agricultural Sciences and Rural Development, Medziphema Campus

ACKNOWLEDGEMENT

At the outset, I do hereby acknowledge the Lord Almighty for His never failing presence that inspired me to strive on.

My sincere gratitude to my supervisor Prof. Y. K. Sharma, Department of Agricultural Chemistry and Soil Science, School of Agricultural Sciences and Rural Development, Nagaland University, for his patience and continued support throughout the investigation. His guidance helped me in all the course of my research work and writing of this thesis.

I would also like to thank the members of my advisory committee Prof. S. K. Sharma and Prof. A. K. Singh, Department of Agricultural Chemistry and Soil Science, Dr. A. P. Singh, Department of Agronomy and Prof. Amod Sharma, Department of Agricultural Economics for their encouragement and insightful comments.

I am thankful to all the staffs of the Department of Agricultural Chemistry and Soil Science for their co-operation and support in completing this study. I owe special thanks to Senior Technical Assistant Ms. Mor, for her help and advice whenever needed.

Special thanks to Dr. Damitre Lytan, for his assistance in analysing the statistical data. I also thank my fellow friends Sashilila, Tinatoly, Chumei, Langnyei, Kevineituo and Toshi for their support and encouragement.

I am thankful to my family who were a constant source of encouragement for me.

Date: 06/10/2020 Place: Medziphema

(MERASENLA AO)

CONTENTS

CHAPTER	TITLE	PAGE NO.
1.	INTRODUCTION	1-6
2.	REVIEW OF LITERATURE	7-44
	2.1 Effect of soil pH on plants and soil properties	7-11
	2.2 Effect of lime on plants and soil properties	11-24
	2.2.1 Effect on growth and yield	11-15
	2.2.2 Effect on nutrient composition and uptake	15-18
	2.2.3 Effect on soil properties	18-24
	2.3 Effect of phosphorus on plants and soil properties	24-36
	2.3.1 Effect on growth and yield	24-28
	2.3.2 Effect on nutrient composition and uptake	29-34
	2.3.3 Effect on soil properties	34-36
	2.4 Effect of boron on plants and soil properties	36-42
	2.4.1 Effect on growth and yield	36-39
	2.4.2 Effect on nutrient composition and uptake	39-41
	2.4.3 Effect on soil properties	41-42
	2.5 Tolerance to soil acidity in plants	42-44
3.	MATERIALS AND METHODS	45-55
	3.1 Experimental site	45
	3.2 Climatic condition	45
	3.3 Characteristics of the experimental soil	45
	3.4 Details of experiments	47-49
	3.5 Observations	49-50
	3.6 Plant analysis	50-52
	3.7 Nutrient uptake	52
	3.8 Soil analysis	52-54
	3.9 Agronomic efficiency	55
	3.10 Nutrient use efficiency	55
	3.11 Analysis of data	55

CHAPTER	TITLE	PAGE NO
4.	RESULTS AND DISCUSSION	56-168
	4.1 Effect of soil pH, lime and varieties on the	56-109
	performance of maize and soil properties (Expt-I)	
	4.1.1 Effect of soil pH, lime and varieties	56-95
	on the performance of maize	
	4.1.1.1 Effect on plant height	56-60
	4.1.1.2 Effect on number of leaves	60-62
	4.1.1.3 Effect on leaf area index	62
	4.1.1.4 Effect on cob length, cob girth and	64-66
	cob weight of maize	
	4.1.1.5 Effect on number of grains per row, number of rows and grains per cob	66-70
	4.1.1.6 Effect on test weight, grain yield and stover yield of maize	70-72
	4.1.1.7 Nitrogen content in grain and stover	72-74
	4.1.1.8 Phosphorus content in grain and stover	74-77
	4.1.1.9 Potassium content in grain and stover	77-79
	4.1.1.10 Calcium content in grain and stover	79-81
	4.1.1.11 Boron content in grain and stover	81-83
	4.1.1.12 Nitrogen uptake in grain and stover	83-85
	4.1.1.13 Phosphorus uptake in grain and stover	85-89
	4.1.1.14 Potassium uptake in grain and stover	89-91
	4.1.1.15 Calcium uptake in grain and stover	91-93
	4.1.1.16 Boron uptake in grain and stover	93-95
	4.1.2 Effect of soil pH, lime levels and varieties on soil properties	95-109

CHAPTER	TITLE	PAGE NO.
	4.1.2.1 Effect on soil pH and electrical conductivity	95-98
	4.1.2.2 Effect on percent base saturation and organic carbon	98-101
	4.1.2.3 Effect on available nitrogen, phosphorus and potassium	101-103
	4.1.2.4 Effect on exchangeable calcium and available boron	103-106
	4.1.2.5 Effect on exchangeable hydrogen, aluminium and total potential acidity	106-109
4. po II	2 Effect of lime, phosphorus and boron on the erformance of maize and soil properties (Expt-	109-165
	4.2.1 Effect of lime, phosphorus and boron on the performance of maize	109-154
	4.2.1.1 Effect on plant height	109-114
	4.2.1.2 Effect on number of leaves	114-116
	4.2.1.3 Effect on leaf area index	116
	4.2.1.4 Effect on cob length, cob girth and cob weight	116-120
	4.2.1.5 Effect on number of grains per row, number of rows and grains per cob	120-125
	4.2.1.6 Effect on test weight, grain yield and stover yield	125-127
	4.2.1.7 Nitrogen content in grain and stover	129-131
	4.2.1.8 Phosphorus content in grain and stover	131-133
	4.2.1.9 Potassium content in grain and stover	133-135
	4.2.1.10 Calcium content in grain and stover	135-137
	4.2.1.11 Boron content in grain and stover	137-140

CHAPTER	TITLE	PAGE NO.
	4.2.1.12 Nitrogen uptake in grain and	140-144
	stover 4.2.1.13 Phosphorus uptake in grain and stover	144-146
	4.2.1.14 Potassium uptake in grain and stover	148-150
	4.2.1.15 Calcium uptake in grain and stover	150
	4.2.1.16 Boron uptake in grain and stover 4.2.2 Effect of lime, phosphorus and boron	152-154 158-165
	on soil properties 4.2.2.1 Effect on soil pH and electrical	158
	4.2.2.2 Effect on percent base saturation and organic carbon	158
	4.2.2.3 Effect on available nitrogen, phosphorus and potassium	161
	4.2.2.4 Effect on exchangeable calcium and available boron	163
	4.2.2.5 Effect on exchangeable hydrogen, aluminium and total potential acidity	165
	4.3 Agronomic efficiency and nutrient use efficiency	165-168
5.	SUMMARY AND CONCLUSION	169-189
	REFERENCES	i-xxiii
	APPENDIX	i-xxxiv

LIST OF TABLES

TABLE NO.	TITLE	PAGES
3.1	Physicochemical properties of two different experimental soils	46
4.1.a	Plant height of maize at different growth stages as affected by soil pH, lime and varieties	57
4.1.b	Interaction effect between pH and lime on plant height at 30 DAS	59
4.2	Number of leaves per plant of maize at different growth stages as affected by soil pH, lime and varieties	61
4.3	Leaf area index of maize at different growth stages as affected by soil pH, lime and varieties	63
4.4	Cob length, cob girth and cob weight of maize as affected by soil pH, lime and varieties	65
4.5.a	Number of grains per row, number of rows and number of grains per cob as affected by soil pH, lime and varieties	68
4.5.b	Interaction effect between pH and lime on number of rows per cob	69
4.5.c	Interaction effect between pH and varieties on number of grains per row	69
4.6	Test weight, grain and stover yield of maize as affected by soil pH, lime and varieties	71
4.7	Nitrogen content in grain and stover of maize as affected by soil pH, lime and varieties	73

TABLE NO.	TITLE	PAGES
4.8.a	Phosphorus content in grain and stover of maize as: affected by soil pH, lime and varieties	75
4.8.b	Interaction effect between pH and lime on phosphorus content in grain	76
4.9	Potassium content in grain and stover of maize as affected by soil pH, lime and varieties	78
4.10	Calcium content in grain and stover of maize as affected by soil pH, lime and varieties	80
4.11	Boron content in grain and stover of maize as affected by soil pH, lime and varieties	82
4.12	Nitrogen uptake in grain and stover of maize as affected by soil pH, lime and varieties	84
4.13.a	Phosphorus uptake in grain and stover of maize as affected by soil pH, lime and varieties	86
4.13.b	Interaction effect between pH and lime on phosphorus uptake in grain	87
4.14	Potassium uptake in grain and stover of maize as affected by soil pH, lime and varieties	90
4.15	Calcium uptake in grain and stover of maize as affected by soil pH, lime and varieties	92
4.16	Boron uptake in grain and stover of maize as affected by soil pH, lime and varieties	94
4.17.a	Soil pH and electrical conductivity of post-harvest soil as affected by soil pH, lime and varieties	96
4.17.b	Interaction effect between pH and lime on soil pH of post-harvest soil	97

TABLE NO.	TITLE	PAGES
4.18.a	Base saturation and organic carbon of post-harvest soil as affected by soil pH, lime and varieties	99
4.18.b	Interaction effect between pH and lime on percent base saturation of post-harvest soil	100
4.19	Available nitrogen, phosphorus and potassium of post-harvest soil as affected by soil pH, lime and varieties	102
4.20	Exchangeable calcium and available boron of post-harvest soil as affected by soil pH, lime and varieties	104
4.21.a	Exchangeable H ⁺ , Al ³⁺ and total potential acidity of post-harvest soil as affected by soil pH, lime and varieties	107
4.21.b	Interaction effect between pH and lime on exchangeable hydrogen and aluminium of post- harvest soil	108
4.22.a	Plant height of maize at different growth stages as affected by lime, phosphorus and boron	111
4.22.b	Interaction effect between lime and phosphorus on plant height at 60 DAS	113
4.22.c	Interaction effect of lime, phosphorus and boron on plant height at harvest	113
4.23	Number of leaves per plant of maize at different growth stages as affected by lime, phosphorus and boron levels	115
4.24	Leaf area index of maize at different growth stages as affected by lime, phosphorus and boron	117

TABLE NO.	TITLE	PAGES
4.25.a	Cob length, cob girth and cob weight of maize as affected by lime, phosphorus and boron	119
4.25.b	Interaction effect between lime and phosphorus on cob weight	121
4.26.a	Number of grains per row, number of rows and number of grains per cob as affected by lime, phosphorus and boron	122
4.26.b	Interaction effect between lime and phosphorus on number of grains per row and number of grains per cob	124
4.27.a	Test weight, grain yield and stover yield of maize as affected by lime, phosphorus and boron	126
4.27.b	Interaction effect between lime and phosphorus on grain yield	128
4.28	Nitrogen content in grain and stover of maize as affected by lime, phosphorus and boron	130
4.29	Phosphorus content in grain and stover of maize as affected by lime, phosphorus and boron	132
4.30	Potassium content in grain and stover of maize as affected by lime, phosphorus and boron	134
4.31	Calcium content in grain and stover of maize as affected by lime, phosphorus and boron	136
4.32.a	Boron content in grain and stover of maize as affected by lime, phosphorus and boron	138
4.32.b	Interaction effect between lime and boron on boron content in stover	139

TABLE NO.	TITLE	PAGES
4.33.a	Nitrogen uptake in grain and stover of maize as affected by lime, phosphorus and boron	141
4.33.b	Interaction effect between lime and phosphorus on nitrogen uptake in grain	143
4.34.a	Phosphorus uptake in grain and stover of maize as affected by lime, phosphorus and boron	145
4.34.b	Interaction effect between lime and phosphorus on phosphorus uptake in grain	147
4.35	Potassium uptake in grain and stover of maize as affected by lime, phosphorus and boron	149
4.36	Calcium uptake in grain and stover of maize as affected by lime, phosphorus and boron	151
4.37.a	Boron uptake in grain and stover of maize as affected by lime, phosphorus and boron	153
4.37.b	Interaction effect between lime and phosphorus on boron uptake in grain	155
4.37.c	Interaction effect between phosphorus and boron on boron uptake in grain	156
4.37.d	Interaction effect between lime and boron on boron uptake in grain and stover	157
4.38	Soil pH and electrical conductivity of post-harvest soil as affected by lime, phosphorus and boron	159
4.39	Percent base saturation and soil organic carbon of post-harvest soil as affected by lime, phosphorus and boron	160

TABLE NO.	TITLE	PAGES
4.40	Available nitrogen, phosphorus and potassium of post-harvest soil as affected by lime, phosphorus and boron	162
4.41	Exchangeable calcium and available boron of post-harvest soil as affected by lime, phosphorus and boron	164
4.42	Exchangeable H ⁺ , Al ³⁺ and total potential acidity of post-harvest soil as affected by lime, phosphorus and boron	166
4.43	Agronomic efficiency of phosphorus and boron	168
4.44	Nutrient use efficiency of phosphorus and boron	168

FIGURE	CAPTION	IN BETWEEN
NO.		PAGES
1.	Effect of soil pH, lime and varieties on plant height of maize at 30, 60 DAS and at harvest	57-58
2.	Effect of soil pH, lime and varieties on grain yield and stover yield of maize	71-72
3.	Effect of soil pH, lime and varieties on nitrogen uptake in grain and stover of maize	84-85
4.	Effect of soil pH, lime and varieties on phosphorus uptake in grain and stover of maize	86-87
5.	Effect of soil pH, lime and varieties on potassium uptake in grain and stover of maize	90-91
6.	Effect of soil pH, lime and varieties on calcium uptake in grain and stover of maize	92-93
7.	Effect of soil pH, lime and varieties on boron uptake in grain and stover of maize	94-95
8.	Effect of lime, phosphorus and boron on plant height of maize at 30, 60 DAS and at harvest	111-112
9.	Effect of lime, phosphorus and boron on grain yield and stover yield of maize	126-127
10.	Effect of lime, phosphorus and boron on nitrogen uptake in grain and stover of maize	141-142
11.	Effect of lime, phosphorus and boron on phosphorus uptake in grain and stover of maize	145-146
12.	Effect of lime, phosphorus and boron on potassium uptake in grain and stover of maize	149-150

FIGURE	CAPTION	IN BETWEEN
NO.		PAGES
13.	Effect of lime, phosphorus and boron on calcium uptake in grain and stover of maize	151-152
14.	Effect of lime, phosphorus and boron on boron uptake in grain and stover of maize	153-154

LIST OF ABBREVIATIONS

AE	-	Agronomic efficiency
Al^{3+}	-	Aluminium
@	-	at a rate of
cm	-	Centimeter
CD	-	Critical Difference
В	-	Boron
BUE	-	Boron Use Efficiency
DAS	-	Days after Sowing
°C	-	Degree Celsius
Ca ²⁺	-	Calcium
Cmol	-	Centimol
dSm ⁻¹	-	Decisiemens per meter
E	-	East
EC	-	Electrical conductivity
et al.	-	et allia (and others/co-workers)
Fe ³⁺	-	Iron
Fig.	-	Figure
g	-	Gram
H^+		Hydrogen
ha	-	Hectare
i.e.	-	Id est (that is)
kg	-	Kilogram
LR	-	Lime requirement
m	-	Meter
mg	-	Milligram
Ν	-	North
Ν	-	Nitrogen
NS	-	Non-significant
NU	-	Nagaland University

No.	-	Number
%	-	Per cent
Р	-	Phosphorus
PUE	-	Phosphorus Use Efficiency
Κ	-	Potassium
RDF	-	Recommended Dose of Fertilizers
SASRD	-	School of Agricultural Sciences and Rural
		Development
Sl. No.	-	Serial number
S	-	South
SEm±	-	Standard error of mean
Viz.	-	Namely

INTRODUCTION

CHAPTER I

INTRODUCTION

Maize (*Zea mays L.*) also known as corn belongs to the family of grasses (*Poaceae*) and is widely cultivated throughout the world. It is a versatile crop which is grown over a range of agro-climatic zones *viz.*, the tropical, subtropical and temperate areas of the world. It is globally recognised as the "Queen of cereals" because of its inherent capacity to give higher yield among cereals. Among the countries, the United States of America stands first in maize production with 371.52 million tonnes (mt) followed by China (256 mt), Brazil (94.50 mt), European Union (59.50 mt), Argentina (42.50 mt) and India (26.50 mt) (Maize Outlook, January 2019).

In India, maize is grown throughout the year and is predominantly a *kharif* crop with a total area of 9.18 million hectare (Agricultural Statistics at a Glance, 2019). It is the third most important cereal crop after rice and wheat and accounts for around 10% of total food grain production in the country. It has great significance as human food, animal feed and has numerous applications in various industries such as the food industries where it is used in the manufacture of starch, gluten, sugar, corn oil and syrup, corn meal and flour. Its application in pharmaceutical, cosmetic, textile and paper industries is observed as well. In addition to this, the past few decades have seen a sharp increase in maize production industry with the United States emerging as the leader in this sector (Mohanty and Swain, 2018).

In the North Eastern Region of India, maize is an important cereal crop having high potentiality for large scale cultivation, but the current production is still lower in comparison to our national average. In Nagaland, maize is one of the important cereal crops next to paddy in respect of area and production. Maize is grown in almost all the districts of Nagaland. In Nagaland, it is grown in an area of 68,960 hectares with annual production of 136540 metric tonnes (Statistical Handbook of Nagaland, 2017).

Maize has been found to be grown successfully in a range of soils from loamy sand to clayey loam. However, neutral pH soils with high organic matter content and water holding capacity is considered good for higher productivity. A challenge in maize production faced by the farmers in the North Eastern Regions of India is soil acidity. The acid soils have high Aluminium (Al) content which leads to grain yield losses up to 60% (The et al., 2006). Nearly 65% of the area under North East, India is under acute forms of soil acidity (pH below 5.5) (Sharma and Singh, 2002). Intense weathering associated with humid climate and heavy precipitation is the cause of soil acidity in this region (Kumar et al., 2012). Some of the problems associated with soil acidity include aluminium toxicity, low nutrient status, and nutrient imbalance (Zesith, 2011). In Nagaland where most of the soils are predominantly acidic in nature, acidity is the main constraint in increasing productivity of these soils. The prevailing factors affecting maize productivity on acid soils are toxicity of aluminium (Al) and iron (Fe), fixation of phosphorus (P), low base saturation and reduction in soil biological activity (Patiram, 1991; Kumar et al., 2012). The negative effect of soil acidity on crops is mainly due to P fixation in soils, where Fe and Al sesquioxides fix sizeable quantities of P, thus making it unavailable to plants. Excess Al³⁺ ions due to soil acidity accumulates in the plant roots and prevents the translocation of P, Mo and other ions from the root to the top, as is evident by the inhibition of root elongation and delayed crop development (Ligeyo and Gudu, 2005). The negative effect of H⁺ ions in acid soils is not distinctive as that of Al^{3+} ions, but excess of H^+ ions affects plant root membrane permeability which interferes with ion transport (Ligeyo and Gudu, 2005). In acid soils where the utilization of native P or added P by plants cannot be carried out efficiently (Swift et al., 1994), it becomes critical to correct soil acidity and also to apply P fertilizer.

For decreasing soil acidity, liming becomes essential along with quantification of lime, which also is an important factor. The nature of soil acidity and soil properties basically determines the lime requirement. Addition of lime increases the soil pH, which eliminates most of the major problems of acid soils. Application of lime eliminates actual and exchange acidities, minimizes hydrolytic acidity, raises the calcium content in the soil (Peoples et al., 1995). Liming also increases the availability of several plant nutrients such nitrogen, sulphur and particularly phosphorus, due to enhanced as mineralization of organic matter. Only around 20% of phosphorus applied as fertilizer is taken up by the crop during the year of its application (Bhat et al., 2007). The rest is fixed in the soil in different range of availability to the subsequent crops. Therefore, one of the benefits of liming acid soils is the increased utilization of residual fertilizer phosphorus by crop. Liming creates a suitable environment (pH 6.0 - 6.5) for nitrifying bacteria, increase in aerobic N fixation process and organic matter decomposition process

In P fixing acid soils, combined application of lime and P is crucial for improved plant uptake of applied P. Phosphorus is required for growth, sugar and starch utilization, photosynthesis, nucleus formation, cell division and fat and albumen formation. The energy from photosynthesis and carbohydrate metabolism is stored in phosphate compounds for growth and reproduction (Ayub *et al.*, 2002). Phosphorus can be easily transported within the plants, moving from older to younger tissues (Ali *et al.*, 2002). Sufficient levels of P in plants results in rapid growth and early maturity and enhances the quality of vegetative growth. Purple leaf coloration is commonly associated with phosphorus deficiency in maize. Phosphorus deficiency is also responsible for crooked and missing rows as kernel twist in maize. Among the cereals, maize has been found to be an exhaustive crop with high yield potential and thus absorbs large quantity of nutrients from the soil during different growth stages. Among the essential nutrients, phosphorus is the most important nutrient that is needed in larger quantity to achieve higher yield (Chen *et al.*, 1994).

Another challenge in maize production is its sensitivity to B deficiency (Salisbury and Ross, 1992; Soylu *et al.*, 2004). Previously, maize was considered to have a relatively low boron requirement as compared to other cereals (Martens and Westermann, 1991). However, boron deficiency in maize has been reported across five continents based on field responses to boron application (Bell and Dell 2008; Shorrocks, 1997; Shorrocks and Blaza, 1973). The first observation of boron deficiency in field grown maize was in the 1960s in United States (Shorrocks and Blaza, 1973), where on application of boron, the yield increase was more than 10% (Woodruff *et al.*, 1987).

Nearly 33% of 36800 soil samples analyzed in India have been found to be deficient in boron (Singh, 2004). The problem has been reported to be more extensive in the eastern and north-eastern states of India which may be due to leaching losses by high rainfall and high soil acidity. In acid soils boron availability is decreased due to adsorption on sesquioxides, iron and aluminium hydroxy compounds. Boron deficiency affects various processes in vascular plants such as root elongation, oxidase activity, sugar translocation, carbohydrate metabolism, nucleic acid synthesis, and growth of pollen tube (Goldbach and Wimmer, 2007; Saleem et al., 2011). Visual deficiency symptoms of boron in maize include foreshortened internodes, narrow white and transparent necrotic spots and smaller cobs with less tightly packed kernels. Boron plays an important role in several functions of the plant, viz., meristem tissue cell division, petal and leaf bud formation, vascular tissue repair, transport and metabolism of sugars, metabolism of RNA and indoleacetic acid, stability of membranes, production and transfer of cytokinin, pollen budding and seed formation (Shelp, 1993; Marschner, 1995).

The key to accurate and profitable fertilizer recommendations is to target specific nutrients that are deficient in soils and to evaluate the response to the applied nutrients. In Nagaland due to acidity- induced soil fertility problems and traditionally minimal use of mineral fertilizers, there is a need for soil acidity management and crop productivity improvement like nutrient management for enhancing food security in the region. It is therefore necessary to determine the yield benefits due to individual as well as combined application of lime and chemical fertilizers. The effect of nutrient management is one of the most important factors that must be controlled to ensure that farmers get good quality high yields (Grazia *et al.*, 2003). Since maize is a heavy feeder that requires comparatively higher amounts of nutrients for higher production, use of inadequate quantity of fertilizers and a decrease in the native soil fertility often leads to nutrient deficiencies and reduced production of this crop.

There is substantiation of differences in acidity tolerance among crop species and among varieties of the same species. Tolerance to low P and Al toxicity has been reported (Malama *et al.*, 2000). Thus, it becomes imperative to screen maize varieties for tolerance to soil acidity which would be an efficient and permanent alternative to increase yields on acidic soils and avoiding enormous yield losses as is often observed with sensitive varieties (Horst *et al.*, 1997). The knowledge gained from the response of maize with different lime doses and chemical properties in the acid soil would help to identify efficient and economical practices for the farmers. Lastly, the combination of liming practices together with selection for more tolerant plants to Al toxicity would be an economical approach.

In this backdrop, the present investigation entitled "Study on Acidity Tolerance of Maize and its Response to Phosphorus and Boron" was carried out with the following objectives:

1. To evaluate the acidity tolerance of maize varieties and response to lime application.

- 2. To study the effect of P and B supply on growth, yield and nutrient uptake in case of maize under acidic soil environment.
- 3. To assess the treatment effect on physicochemical properties of the soil.

CHAPTER II

REVIEW OF LITERATURE

REVIEW OF LITERATURE

The information pertaining to the present investigation entitled "Study on acidity tolerance in maize and its response to phosphorus and boron" and related crops have been presented in this chapter.

2.1 EFFECT OF SOIL pH ON PLANTS AND SOIL PROPERTIES

Marquez and Baucas (1990) reported that an increase in soil pH to 5.74, enhanced NPK absorption which in turn resulted to higher yields in various crops.

Goldberg (1997) reported that at lower soil pH, adsorption of B on sesquioxides is greater compared to adsorption of B on soil constituents other than sesquioxides at relatively higher pH. Boron adsorption occurs specifically on mineral surfaces of sesquioxides where innersphere surface complexes are formed which may enhance the irreversibility of B desorption.

Fageria and Zimmermann (1998) studied the influence of pH on nutrient uptake on various crop species and reported varied responses of the crops to soil pH reflecting the genetic diversity among species. In all of the crop species, uptake of calcium (Ca) and magnesium (Mg) decreased with a decrease in soil pH. In general, increasing soil pH decreased the uptake of nitrogen (N), phosphorus (P), and potassium (K) in rice, but uptake of these elements increased in wheat, corn, and common bean.

Alam *et al.* (1999) reported that excess H^+ in the growth medium affects plant growth by two processes: a) H^+ may cause injury to the root tissue, and b) specific effects on root ion fluxes via H^+ competition with base cations for uptake which causes a decrease in the rate of cation absorption as a result of the competition between the similarly charged ions for binding and carrier sites. Gale *et al.* (2001) stated that acidic soil would result in calcium and magnesium deficiency and poor bacterial growth.

Keerthisinghe *et al.* (2001) reported that crops growing in tropical acid soils can only recover and utilize 10–25% of the P fertilizer applied due to their high P fixations by Al and Fe oxides.

Scott *et al.* (2001) stated that incorporating lime and/or use of acid-soil resistant genotypes are integral to the management of acid soils.

Giller *et al.* (2002) reported that soil acidity has also been shown to reduce the agronomic and recovery efficiencies of nutrients such as P by plants.

Nwachukwu (2002) observed that hot water soluble B content decreased with increase in soil pH.

Abreu *et al.* (2003) studied the relationship between acidity and soil chemical properties and reported that pH correlated positively with P, Ca, Mg, K, base saturation and CEC and negatively with Al saturation. The Al³⁺ was the predominant exchangeable cation in 32% of the soils with pH below 5.6.

Adnan *et al.* (2003) in their study reported that the reaction of $H_2PO_4^{2-}$ and HPO_4^{-} with Al and Fe ions forms a precipitate (Al or Fe phosphate minerals) which reduces diffusion of P into plant roots.

Sierra *et al.* (2003) reported that soil acidity reduces maize and other grain yields by about 10% in tropical areas.

Arora and Chahal (2005) found significant positive correlation between soil pH and available B content.

Mariano and Keltjens (2005) reported that Al had negative effects on the uptake of macro and micronutrients. Al effects were most pronounced on the uptake of Ca and Mg, with respective reductions of 61% and 72%, when averaged across the 10 genotypes. Among the micronutrients, the most pronounced effects of Al were noted on Mn and Zn.

Kifuko *et al.* (2007) stated that an increase in soil pH results in precipitation of exchangeable and soluble Al as insoluble Al hydroxides.

Ligeyo (2007) reported that Al toxicity and P deficiency reduce maize grain yield by about 16 and 28%, respectively, in acid soils.

Mallarino (2011) in a study reported that maize growth and performance is affected by soil reaction. The study revealed that the optimum performance of maize was observed at pH 6.0 and 6.5 in areas with high pH subsoil and in areas without high pH (calcareous) subsoils, respectively.

Awani (2012) stated that soil acidification leads to a decrease in soil available phosphorus.

Bruckner (2012) reported that lower soil pH indicates larger number of hydrogen ions in the soil. Hydrogen ions can appear in varying amount in the soil environment which can affect the level of electrical conductivity. Higher amount of hydrogen ions in the soil will show a higher rate of electrical conductivity. Hence, low soil pH due to large number of hydrogen ions in the soil may encourage soil electrical conductivity.

Kai *et al.* (2012) noted that crop performance is affected indirectly by low soil pH from aluminium and manganese toxicity resulting from overly acidic conditions of the soil.

Lauchli and Grattan (2012) stated that though plants differ in their tolerance to extreme pH, most agricultural plants perform optimally at a pH near neutrality.

Lin *et al.* (2012) in their study on effect of acidic growth conditions on soybean reported that low pH effects the root growth and development and absorption of water and nutrients

Dewi-Hayati *et al.* (2014) reported that grain yield reduction in acid soils varied from 2.8 to 71%.

Sahin (2014) reported that pH of soils is not effective upon the increase of boron concentration in maize with boron fertilizer implementation.

Behera and Shukla (2015) reported that soil pH is positively and significantly correlated with exchangeable K⁺, Ca²⁺ and Mg²⁺ content in soils.

Tandzi *et al.* (2015) found maize yield reduction under acid soils to be up to 69%.

Khadka *et al.* (2016) in a study on relationship between soil pH and macronutrients revealed that soil pH was significantly and negatively correlated with total N, available P_2O_5 and extractable K₂O, while there was significant and positive correlation between soil pH and extractable Ca, extractable Mg and available S. In addition, with the increase in soil pH by one unit, N, P_2O_5 and K₂O decreases by 0.056, 51.86 and 3.90 units, respectively and vice-versa. On the other hand, Ca, Mg and S increased by 382.2, 46.09, 1.2 units, respectively and vice-versa.

Kundu *et al.* (2017) reported that available boron was significantly and negative correlated with pH. The result indicates that available boron increased with decreased in pH value. Boron adsorption by soils increased as a function of solution pH in the range of pH 3 to 9 and decreased in the range of pH 10 to 11.5.

Otieno *et al.* (2018) observed positive relationships between pH and N, P and K nutrient uptake in soybean.

Kaur *et al.* (2019) stated that in acidic soils, effects of Al toxicity in plants include inhibition of root growth, inhibition of root cell division, modification of the cytoskeleton and inhibition of nutrient uptake. In most acidic soils, P ends up being fixed by Fe which degrades the condition for crop production.

Pan *et al.* (2020) studied the growth and yield of maize on two soil pH of 4.8 and 5.0 and observed that changes in soil pH below the critical soil pH, increased soil pH significantly and increased maize height and the yield of maize shoots and roots. Increase in the pH also increased N accumulation in maize, the N derived from fertilizer in maize, physiological N use efficiency, and N use efficiency (NUE) by maize. Changes in soil pH above the critical soil pH however showed no significant effect on these parameters.

2.2. EFFECT OF LIME ON PLANTS AND SOIL PROPERTIES2.2.1 Effect on growth and yield

Venkatesh *et al.* (2002) reported that liming @ 0.25 LR increased both grain and straw yields of maize by about 10 per cent, which might be due to the increase in soil pH, exchangeable bases, available P and reduction in exchangeable Al.

Sierra *et al.* (2003) observed that grain yield, number of grains, test weight and number of rows per ear of maize was higher in limed plots than in non-limed plots. In non-limed plots, soil acidity affected aerial growth in two ways: (i) the appearance of leaves was delayed, and (ii) the individual leaf area and LAI were reduced.

Chatterjee *et al.* (2005) reported that incorporation of lime at 10, 50 and 100% LR increased the pod yield and nutrient uptake of groundnut, where maximum yield was observed at 100% LR.

Brajendra *et al.* (2006) reported that there was a significant increase in plant height, cob length, number of grains/cob and grain yield of maize in all the levels of lime addition (0.5, 1.0, 1.5 and 2.0 t/ha) over no lime.

Dixit (2006) reported that grain and stover yield of maize increased significantly due to lime and P interaction effect. The P application increased grain yield significantly up to 39 kg P ha⁻¹ at no lime, 26 kg P ha⁻¹ at half of the lime requirement and 13 Kg P ha⁻¹ at full lime requirement. Increase in stover yield at no lime and half of the lime requirement was up to 26 kg P ha⁻¹ and at full lime requirement, a significant increase was recorded at 13 kg P ha⁻¹.

Adhikary and Karki (2007) studied the effect of lime and fertilizer on local and improved varieties of maize and suggested that irrespective of the maize varieties application of fertilizers (60: 30: 30) with agricultural lime (4 t ha⁻¹) was beneficial for increased grain production.

Bhat *et al.* (2007) found that application of lime caused a significant increase in grain (GY) and straw yield (SY) of wheat. The magnitude of increase in GY and SY due to liming was 26.8 and 18.6 per cent, respectively over the no lime treatment, irrespective of the sources and levels of lime.

Busari *et al.* (2008) reported that application of lime along with poultry manure in maize, significantly improved plant height, stover and cob yield as well as highest grain yield of maize as compared to control.

Fageria and Baligar (2008) in their review stated that in acid soils with a $pH (H_2O) < 5.0-5.5$, initial lime applications result in large increases in crop growth and yield, mainly due to the amelioration of Al toxicity. Lime rates that raise the soil pH (H₂O) to 6 or greater are often associated with lime-induced depressions in plant growth, mostly due to micronutrient deficiencies, and sometimes deficiencies of K and Mg. Lime-induced depressions in plant P content have also been recorded.

Singh *et al.* (2009) reported that application of NPK with lime resulted in the highest grain yield of maize which produced 5.6 t ha⁻¹, whereas application of NPK alone (no lime) in acid soil produced 1.8 t ha⁻¹ grain yield of maize

Andric *et al.* (2012) found that liming resulted in maize grain yield increases of up to 33% in the first year and 35% in the second year.

Kumar *et al.* (2012) reported that liming at 300 kg ha⁻¹ (furrow application) caused 32% yield increase over control. Combined application of NPK+ lime further boosted the yield improvement up to 147% over control.

Kumar *et al.* (2014) reported that growth attributes like plant height and leaf area index and yield and yield attributes of ricebean were significantly higher with the application of lime at 0.6 t ha⁻¹ than lower levels of lime (0.2 t ha⁻¹ and 0.4 t ha⁻¹) and control.

Kumar (2015) reported that increasing levels of lime (control, 0.2, 0.4 and 0.6 t/ha) significantly increased cob length, number of row/cob, number of grain/row, number of grain/cob and 1000-grain weight than lower doses of lime. The grain, stover and biological yield of maize also significantly increased with increasing levels of lime up to 0.6 t ha⁻¹.

Behera *et al.* (2017) reported that liming of soil and their combined application with FYM resulted in increasing root length, cob length, diameter and seed weight cob^{-1} . Inorganic (lime) amelioration of acid soil resulted in 32 per cent higher yield compared to the yield of 22 q ha⁻¹ due to inorganic nutrition only.

Lokya *et al.* (2017) observed that combined application of soil test based recommended dose (STD) + vermicompost (VC) @ 2.5 t ha⁻¹+ lime @ 0.2 LR increased the concentration, uptake and recovery of the macro primary nutrients in maize crop.

Opala (2017) reported a significant effect of lime, P, and P by lime interactions on plant heights and dry matter. Without lime application, plant height and dry matter increased with increasing P rates but, with lime, it increased up to $30 \text{ kg P} \text{ ha}^{-1}$ and thereafter declined from 30 to $100 \text{ kg P} \text{ ha}^{-1}$.

Bekele *et al.* (2018) in a study demonstrated that highest leaf area index (5.91), plant height (3.48 m), cob length (47.83 cm), number of grain per cob (644) and above ground dry biomass yield (22 t ha-1) were exhibited by the application of lime along with vermicompost and mineral P fertilizer over the control.

Ferdous *et al.* (2018) reported that growth parameters, yield components and yield of rice increased with increasing lime rate (0, 0.50, 1.00, and 1.50 t ha⁻¹) along with fertilizer application in acidic soil. The highest grain yield (2.90 t ha⁻¹) was recorded from the application of 1.50 t ha⁻¹ lime and the lowest (2.06 t ha⁻¹) was from control (0 t ha⁻¹), irrespective of fertilizer application.

Opala *et al.* (2018) in a study on effect of lime and fertilizers on maize yield observed that maize did not respond to lime without fertilizer. Maize responded to the fertilizers containing N and P but not to application of lime without fertilizer at all experimental sites.

Adikuru *et al.* (2019) reported that lime application of 2 t ha⁻¹ significantly increased number of grain rows cob^{-1} by 5.7 %, number of grains row⁻¹ by 17.9 %, number of grains cob^{-1} by 24.9 %, weight of hundred seeds by 19.6 % and grain yield by 58.5 % over control.

Devkota *et al.* (2019) reported that plant and yield characters showed significant variation due to variety, lime and their interaction. The highest significant grain yield was given by Manakamana-3 maize variety followed by

Deuti variety. Application of lime at the rate of 1.5 to 1 t ha⁻¹ showed statistically significant highest grain yield.

Teshome and Garadew (2019) reported that in an experiment with treatments of five levels of potassium fertilizer (0, 20, 40, 60, and 80 kg ha⁻¹) and two levels of lime (0 and 4.6 t ha⁻¹), response of soybean in terms of grain yield was highest when 60 kg K₂O ha⁻¹ and 4.6 ton lime ha⁻¹ was applied while the lowest yield was recorded in the control treatment.

2.2.2 Effect on nutrient composition and uptake

Friesen *et al.* (1980) reported that liming alleviates Al toxicity in soil leading to improvement in root growth of plants which eventually results in increased uptake of nutrients as the roots are able to explore greater volume of soil.

Gupta *et al.* (1989) conducted a study to observe the effect of lime on various crops and response to liming was variable among the crops. The uptake of N, P, K increased in all the crops where soybean recorded maximum uptake of all the three nutrients followed by maize, barley and wheat.

Quaggio *et al.* (1991) reported that liming of the surface soil promoted amelioration of the soil down to 100 cm, increased root growth and nutrient absorption in maize, mainly nitrogen, which increased about ten fold.

Su et al. (1994) reported an increase in B uptake in alfalfa due to liming.

Oya and Khondaker (1996)) reported the enhanced concentration of N in common millet as a result of addition of lime in acid soil.

Rosolem and Caires (1998) studied response of peanut to liming and Mo application and reported that their application increased N contents in the leaves as well as N uptake.
Jibrin *et al.* (2002) reported that application of lime at 1.35 t CaO ha⁻¹ significantly raised the P concentration of index leaves and the total P uptake in maize. Liming also significantly raised the concentrations of Ca and Mg concentrations of index leaves of maize by 12 and 5 %, respectively.

Busari *et al.* (2005) reported an increase in N and P uptake in maize due to liming.

Dixit (2006) reported that phosphorus uptake by grain and stover significantly increased with increase in lime dose up to full lime requirement (3.08 t ha⁻¹). The increase over non lime in P uptake by maize grain was to the tune of 68 and 41 % during first and second year, respectively.

Bhat *et al.* (2007) found that application of lime caused a significant increase in N and P content of crop. The mean increase of N concentration was 17.1% over no lime and phosphorus concentration was 51.5% over no lime. Application of lime did not show any specific trend of increase or decrease in concentration of K in wheat plants. The concentration of K was increased to about 0.92% in limed soils.

Ranjit *et al.* (2007) reported that in groundnut, application of 100% lime requirement recorded higher total nitrogen, phosphorus, potassium and calcium uptake than other levels of lime.

Busari *et al.* (2008) reported that lime improved the maize leaf N concentration when provided in combination with poultry manure (PM) and NPK while the combination of PM, lime and NPK gave significantly higher leaf P and K concentrations than any of the other treatments or the control.

Ossom and Rhykerd (2008) in a study reported that N concentration in tubers of sweet potato increased as a result of liming in acid soil.

Singh *et al.* (2009) studied that lime application with NPK resulted in significantly higher uptake of P, Ca, Mg, S and B by maize as compared to NPK alone in acidic soils. Uptake of K by maize in NPK and NPK + lime treated plots was at par. Hot water soluble B content showed a decreasing trend with lime + NPK compared to NPK application without lime.

Sultana *et al.* (2009) reported that the concentrations of P and Ca of wheat grain remained unaffected on liming whereas total uptake of P and Ca were increased due to application of lime which was mainly associated with increased wheat yields.

Kovacevic and Rastija (2010) examined the effect of liming on maize and reported that liming improved maize nutritional status and increased P, Ca, Mg and Mo concentration and decreased high Mn content to the adequate range.

Moreira and Fageria (2010) reported that N, Ca, and Mg concentrations in alfalfa increased with increasing lime rates which may be associated with higher N₂-fixation rate by N₂-fixing bacteria and higher Ca and Mg uptake with increasing lime rates.

Beukes *et al.* (2012) reported that liming significantly affected P, K, Ca, Mg and Mn concentrations of maize leaves but had no clear effect on leaf N, Cu, Fe and Zn. The increased uptake of the former nutrient elements indicates that liming improved the use efficiency of these elements by the maize crop.

Muindi *et al.* (2015) reported a significant increase of P concentrations in plant tissues of maize after lime application.

Meena and Varma (2016) reported that different lime levels (100, 200 and 300 kg ha⁻¹) improved NPK uptake in mungbean, where maximum was observed at 200 kg ha⁻¹ as compared to control.

Lynrah and Nongmaithem (2017) reported that application of lime at 1.5 t ha⁻¹ gave the highest response in terms of NPK uptake in soybean under acidic soil of Nagaland.

Bhindhu *et al.* (2018) in a study on influence of lime on plant nutrient content of rice reported that content of potassium, calcium and sulphur in plant increased at the lime level as per lime requirement.

Otieno *et al.* (2018) in a study on effects of farmyard manure, lime and inorganic fertilizers in soybean, reported that combined treatments of NPK+lime and manure+lime+NPK recorded higher uptake of N, P and K nutrients rather than individual treatments.

Han *et al.* (2019) reported that K uptake by maize and wheat crops under lime application significantly increased by 37.6% to 155.1% compared with the no-lime treatments.

Yadesa *et al.* (2019) reported significant increase in P uptake of maize with application of P and lime compared to the control experiments.

Zhihao *et al.* (2019) observed that increase of soil pH due to lime application increased the contents of soil available nitrogen and exchangeable Ca^{2+} and exchangeable Mg^{2+} , which promoted the uptake of nitrogen, phosphorus and potassium nutrients of crops.

2.2.3 Effect on soil properties

Mikko (1972) reported that liming, in general, increased the B retention capacity of soil due to formation of insoluble metaborate.

Garica (1975) in a study reported that the pH of acid soils increases due to liming, and adsorption is higher with higher rate of lime application and calcium deficiencies are ameliorated. Haynes (1982) in a study on effects of liming on phosphate availability in acid soils reported an increase in P availability due to increase in mineralization of P by lime.

Haynes and Naidu (1998) reported that increase in K availability in soil increases due to displacement of exchangeable K by Ca in the soil exchange complex.

Andersson and Nilsson (2001) in a study reported that liming increases microbial activity which enhances the mineralization of organic matter and thus releasing inorganic nutrients such as N and P.

It was reported by Ghosh and Mukhopadhyay (2001) that liming generally increased K fixation in soil, though liming soils of acidic reaction increased the cumulative release of potassium over unlimed system.

Caires et al. (2002) reported that application of lime in acid soil increases the soil pH as a result of precipitation of Al as Al(OH)₃.

Venkatesh *et al.* (2002) observed that there was an overall beneficial effect of liming on available P which appeared to be related to the suppression of exchangeable Al content to the extent of 88%. Liming resulted in a significant increase in pH, exchangeable Ca and Mg, whereas exchangeable K decreased which may be due to the entrapment of exchangeable K by different forms of Al precipitated upon liming the soil. Liming @ 2 t ha⁻¹ (25% LR) resulted in remarkable decrease in exchangeable Al content of soil from 0.92 to 0.11 cmol (P⁺) kg⁻¹. Exchangeable acidity, which is a direct function of exchangeable Al⁺ and H⁺ registered a decline up to 72% on liming.

Hue (2004) in a study conducted on acid soils reported an increase in pH of soil due to liming possibly due to precipitation of Al as as Al(OH)₃.

Chaterjee *et al.* (2005) reported a decrease in soil acidity parameters (exchangeable and total acidity, exchangeable H^+ , Al^{3+} and Fe^{3+} and total Fe and Al) due to liming which was maximum at 100% LR.

A study conducted by Rahman *et al.* (2005) in acidic alluvial soils at Dinajpur and Rangpur in Bangladesh reported that soil pH, available P, exchangeable Ca, and hot-water-extractable B were increased by liming. The available P, exchangeable Ca and hot-water-extractable B showed a percent increase of 235.185 and 186.792 %, 132.114 and 130.28 % and 62.5 and 69.38 %, respectively at Dinajpur and Rangpur.

Brown *et al.* (2008) reported that one-time broadcast application of calcitic lime @ 7000 kg ha⁻¹ significantly increased the pH by 0.7 units, while Al significantly lowered relative to the control.

Sarkar (2009) found that within the forms of K, water soluble K increased but exchangeable K decreased in all the soils after liming.

Singh *et al.* (2009) reported that with lime and fertilizer use, the pH of acid soil (5.5) increased by 0.9 units, while it decreased in unlimed plots.

Basak (2010) reported that liming in general increased the potassium availability in soils which is often low in acid soils due to the formation of soluble K salt by soil acids and their loss by leaching from the soil.

Dasog *et al.* (2010) found significant increase in soil pH with addition of lime, where application of lime at 100% lime requirement recorded a highest soil pH of 6.03 and the lowest exchangeable soil acidity. Also the highest available phosphorus in the soil was recorded at 100% of lime requirement.

Kumar *et al.* (2010) showed that liming has positive influence in exchangeable and reserved form of K.

Andric *et al.* (2012) observed that as a result of liming, soil pH increased from 4.75 to 5.28 and was close to neutral after application of 5 t ha^{-1} and 20 t ha^{-1} of lime, respectively. Liming also considerably affected soil nutrient availability because of increases in P (+44%), Ca (+58%), Mg (+74%), and B (+86%) and decreases in Mn (-33%) and Fe (-28%), compared to the control.

Beukes *et al.* (2012) observed that liming at various levels had highly significant effects on soil pH, extractable acidity (EA), acid saturation (AS), P, Ca and Mg. Soil pH (KCl), P, Ca and Mg increased, whereas EA and AS decreased significantly with an increase in lime application. Significant lime \times fertiliser interaction was also observed in terms of soil P. The trends in the latter whereby at low soil pH there was no significant increase in P, followed by an increase as pH increased due to liming.

Costa (2012) reported that liming leads to an increase in soil organic matter (includes organic carbon).

Lege (2012) reported low pH value in soils of control treatment which was attributed to low exchangeable basic cations at the soil exchange sites due to exhaustive uptake of exchangeable bases by plants during the growing period.

Voor (2012) stated that soil acidity results in low exchangeable base cations due to aluminium toxicity.

Kisinyo *et al.* (2013) observed that lime significantly reduced exchangeable Al^{3+} and increased soil pH and available P. Lime increased soil pH because of the likely displacement of Al^{3+} , H^+ and Fe³⁺ ions by Ca²⁺ ions it contains. This led to the observed reduction in P sorption at all the sites.

21

Osundwa *et al.* (2013) reported that addition of lime had significant increase on the soil pH above the control treatment with lime addition of 1.0, 1.5 and 2 t/ha. The soil available P was highest with lime addition of 2 t ha⁻¹.

Amer *et al.* (2014) observed a significant increase in the pH in lime treated soil and lowest total and exchangeable acidity were found to be with application of 20% lime of LR.

Das and Saha (2014) reported that liming of acid soil with half of its requirement and combined application of N and K fertilizers significantly reduced K fixation and increased water soluble K which can easily be taken up by the growing plants. Addition of lime decreased the exchangeable K in soil system the reason being the precipitation of exchangeable aluminium (Al³⁺) and iron (Fe³⁺) ions as hydroxy-aluminium and hydroxy-iron polymers which act as props between layer silicates, thereby inhibiting the collapse of the layers for the K ions. Also, the amount of exchangeable K was more in full limed as compared to half limed soil. This is due to liming with higher doses; higher amount of Ca²⁺ occupy the exchange sites which encourages release of non-exchangeable K in the exchangeable phase.

Kamaruzzaman *et al.* (2014) reported that the application of different rate of lime increased the K availability of soils and better concentration of available K was obtained with lime rate at 1.5 t ha⁻¹.

Kisinyo *et al.* (2014) reported that lime could reduce the P sorption in acid soils resulting in increased available P for plants.

Badole *et al.* (2015) in a pot experiment reported that decrease in values of all the forms of acidity i.e., total acidity, hydrolytic acidity, exchange acidity, electrostatically bound aluminium (EBAl³⁺), and electrostatically bound hydrogen (EBH⁺) was greater in full lime (L₁) than in half lime (L_{1/2}) treatment under Entisols of the terai zone, followed by Entisols of coastal saline zone, Inceptisols, and Alfisols.

Benjala *et al.* (2015) reported that liming effects caused exchangeable potassium (K) to increase with increasing rates of dolomitic limestone application. This is because raising the pH through liming soils particularly those with pH-dependent charge increases the soils' cation exchange capacity (CEC) and thus increases the soils' capacity to adsorb K.

Dey and Nath (2015) reported that liming along with integrated nutrient management practices showed improved status of post harvest soil analysis in organic C, N and P, but available K status declined.

Samant (2015) reported that application of lime at 100% LR was beneficial in improving the fertility status of acid soil. Maximum pH, available P, K, exchangeable Ca and Mg in post harvest soil was recorded at 100% LR with RDF.

Sethi (2015) narrated that lime application (inorganic) increased pH, base saturation, and cation exchange capacity and decreased Al, Fe, and Mn availability, acidity and P fixation.

In a field experiment carried out by Toppo and Kumar (2018), it was observed that liming showed no significant result in soil organic carbon. Also, the availability of phosphorus was more in limed plots as compared to that of control plot although it was treated with SSP. The reasons for this have been attributed to breaking of the complex of iron and aluminum phosphates by lime on acid soils and increasing the availability of phosphates in the form of calcium phosphates.

Han *et al.* (2019) reported that concentration of available K and exchangeable Ca^{2+} in soil had a positive relationship with lime addition rates.

Shen *et al.* (2019) reported that lime might be a better choice over byproduct amendments to improve chemical properties of the acidic soil. Results showed that lime significantly increased the pH and electrical conductivity of soil.

2.3. EFFECT OF PHOSPHORUS ON PLANTS AND SOIL PROPERTIES

2.3.1 Effect on growth and yield

Hamdi and Woodard (1995) reported an increase in leaf area index of corn with increase in P levels (0, 12, and 24 mg P kg⁻¹ soil).

Arya and Singh (2000) reported that P application of 90 kg P_2O_5 ha⁻¹ resulted in significantly more grain and straw yields compared with 60, 30 and 0 P_2O_5 ha⁻¹.

Sarma *et al.* (2000) in a field study on a composite maize variety 'Vijoy' found that on supplying three levels of P (20, 40 and 60 kg P_2O_5 ha⁻¹), maize responded significantly up to 60 kg P_2O_5 ha⁻¹ but the optimum P rate for maize was 20 kg P_2O_5 ha⁻¹.

Ayub *et al.* (2002) reported that leaf area of maize increased with increase in nitrogen and phosphorus levels.

Venkatesh *et al.* (2002) reported that on application of 0, 30, 60 and 90 kg P_2O_5 ha⁻¹, the grain and straw yield of maize increased significantly at phosphorus level up to 90 kg P_2O_5 ha⁻¹ over control when applied without FYM or lime. However, the P levels responded significantly only up to 60 kg P_2O_5 ha⁻¹ when applied along with lime/ FYM or lime + FYM. Similar trend as in grain and straw yields was also seen in case of test weight of maize. Phosphorus use efficiency by maize increased up to 60 kg P_2O_5 ha⁻¹, thereafter it significantly reduced indicating that PUE decreases at higher doses of P due

to the fact that plants grown in extremely P deficient soil exhibit greater P sorption at lower doses of P.

Alias *et al.* (2003) studied the effect of varying levels of phosphorus (0, 50, 75, 100, 125 and 150 kg P ha⁻¹) on yield and yield components of maize on a sandy loam soil and reported that there was a significant increase in leaf area per plant, number of grains per cob and 1000- grains weight with increasing levels of P.

Assuero *et al.* (2004) in a study reaffirmed the important role of phosphorus in cell division and cell enlargement, wherein they reported a decreased cell production, and ultimately leaf elongation rates, in P-deficient maize plants.

Khan *et al.* (2005) studied the effect of different levels of phosphorus (0, 25, 50, 75 and 100 kg P_2O_5 ha⁻¹) on maize and reported that phosphorus significantly increased grain weight per cob, number of grains per cob, cob weight and 1000-grain weight and maximum values was observed at 75 kg P_2O_5 ha⁻¹.

Hussain *et al.* (2006) reported an increase in plant height of maize with increasing levels of phosphorus and attributed the promotion effect of P on plant height to better root development and nutrient absorption.

Hussain *et al.* (2007) studied that maximum number of grains cob⁻¹ and cob weight of maize was obtained with 90 kg P ha⁻¹ while minimum grains cob⁻¹ was obtained with 30 kg P ha⁻¹ and minimum cob weight was obtained with 30 and 60 kg P ha⁻¹. Increase in phosphorus level also increased 1000-grain weight and grain yield of maize where maximum was obtained with 90 kg P ha⁻¹.

Ali *et al.* (2008) reported that P application of 60, 90, 120 kg ha⁻¹ resulted in increase in number of cobs plant⁻¹, number of grains cob⁻¹, thousand

grain weight and grain yield as compared to control. P dose of 60 kg ha⁻¹ resulted in maximum increase in number of grains cob⁻¹, thousand grain weight and grain yield.

Amanullah *et al.* (2009) reported an increase in plant height, leaf area index, grain and stover yield, harvest index, shelling percentage, thousand grain weight and grains ear^{-1} of maize with increase in levels of phosphorus fertilizer.

Amanullah *et al.* (2010) reported that among P levels of 30, 60 and 90 kg P ha⁻¹, the highest level of 90 kg P ha⁻¹ increased plant height, number of leaves per plant, mean leaf area, dry weight of leaf, stem and ear as well as biomass yield and harvest index of maize.

Masood *et al.* (2011) found that P at the rate of 100 kg ha⁻¹ was the optimum rate for the yield components of maize like plant height, number of grains per cob and thousand grain weights. Further increase in P above 100 kg ha⁻¹ did not have a direct proportional effect on the yield components. However, number of cobs per plant and grain yield was found to be optimum at 100 kg P_2O_5 ha⁻¹ followed by 150 kg P_2O_5 ha⁻¹.

Khan *et al.* (2014) reported that the response of maize to three levels of phosphorus (60, 90 and 120 kg ha⁻¹) was found to be non-significant in case of plant height and leaf area while number of leaves plant⁻¹ and dry weight of plants were significant with highest values where P was applied @ 120 kg ha⁻¹ followed by 90 kg ha⁻¹.

Nsanzabaganwa *et al.* (2014) found that the yield attributes of maize such as number of grains per cob, cobs per plant, test weight were found to have no significant effect on P application except on test weight, which was significantly higher at 26.4 kg P ha⁻¹. The effect of P on grain and stover yield was found to be significant and the highest was reached at 26.4 kg P ha⁻¹ after which further increase declined the yield. There was an increase in grain yield, amounting to 12.6, 17.5, and 14.3% with 13.2, 26.4, and 39.6 kg P ha⁻¹ over control. In the case of stover yield, the corresponding increases were 28.6, 45.8, and 27.0%. Increase beyond 26.4 kg P ha⁻¹ caused a reduction in stover yield by 28.7%.

Amanullah and Khan (2015) reported that phosphorus applied at the two higher rates (75 and 100 kg P ha⁻¹) increased number of grains ear⁻¹ and grains row⁻¹, 1000 grains weight, grain yield, harvest index and shelling percentage of maize.

Umeri *et al.* (2016), observed an increase in maize plant height and number of leaves, with the application of 40kg N and 40 kg P ha⁻¹, respectively.

Chandrakala *et al.* (2017) reported higher phosphorus use efficiency (PUE) of maize when P was applied at the rate of 75 % of recommended dose along with recommended dose of N and K. This could be due to maximum utilization and uptake of added P and further application beyond this level was non beneficial and was also non economical to farmers as the PUE by the crops were only 20-40 per cent in general, the remaining amount was fixed in the soil.

Khan *et al.* (2017) reported that application of phosphorus significantly influenced the cob weight, grains row⁻¹ (35.30), grain rows cob⁻¹, grains cob⁻¹ and 100-grain weight of sweet corn. The treatment consisted of 5 levels of phosphorus (40, 50, 60, 70 and 80 kg P_2O_5 ha⁻¹) and maximum values for all was observed at 60 kg P_2O_5 ha⁻¹.

Kumar *et al.* (2017) reported that application of increasing levels of both phosphorus (90 kg P_2O_5 ha⁻¹) and sulphur (45 kg S ha⁻¹) resulted in a significant increase in seed and stover yield of soybean.

Opala (2017) reported that the highest dry matter yield of maize was obtained with 30 kg P ha^{-1} .

Pal *et al.* (2017) reported that phosphorus levels (40, 50 and 60 kg P_2O_5 ha⁻¹) significantly influenced the growth parameters, yield attribute and yield of maize *viz.*, plant height, number of cobs per plant, cob length, cob girth, number of grains per cob, 100 grain weight, grain yield and stover yield and highest values of these growth parameters, yield attribute and yield parameters were observed with application of 60 kg P_2O_5 ha⁻¹ while the lowest values were recorded under 40 kg P_2O_5 ha⁻¹.

Sadiq *et al.* (2017) reported that P applied at 120 kg ha⁻¹ increased rows ear⁻¹ by 40%, grains row⁻¹ by 18%, grains ear⁻¹ by 41%, thousand grains weight of maize by 8%, biological yield by 41%, stover yield by 34%, grain yield by 55% and harvest index by 10% over control.

Timlin *et al.* (2017) reported an increase in leaf appearance rate and area of maize due to P application

Sharma *et al.* (2018) reported that application of phosphorus at 75 kg ha⁻¹ was found to be best over its lower levels ((0, 25 and 50 kg ha⁻¹) on yield attributes (cob length, number of grains cob⁻¹ and 1000 grain weight) and yield (grain and stover) of hybrid maize which was closely followed by phosphorus at 50 kg ha⁻¹.

Irfan *et al.* (2019) studied the interactive effect of P and B on wheat wherein results revealed that yield and yield related attributes increased linearly with the addition of B at each P level. Nonetheless, the significant interactive effect of both nutrients was most pronounced in the treatment having 90 kg P ha⁻¹ and 1.5 kg B ha⁻¹.

2.3.2 Effect on nutrient composition and uptake

Chirnogeanu *et al.* (1997) found that concentration of absorbed phosphorus in maize leaves was 70-80% higher in plants which received 120 kg P_2O_5 ha⁻¹, as compared to unfertilized variants. However, there was an increase in potassium content in plants without phosphorus application. The highest values of potassium content corresponded to the lowest values of plant calcium content.

Dubey (2000) with reference to phosphorus use efficiency in crops stated that the decrease in phosphorus use efficiency at higher doses of P is because plants grown in P deficient soil exhibit greater P sorption at lower doses of P.

Shankarlingappa *et al.* (2000) reported that combined application of P and S up to 50 kg P_2O_5 and 20 kg S ha⁻¹ showed significant synergistic effect on uptake of N, P and S by cowpea which was attributed to higher concentration of nutrients as a result of increased availability of nutrients from soil and fertilizer.

Khan *et al.* (2002) reported a linear increase in total biomass, straw yield and grain yield of mungbean was observed with increasing rates of phosphorus fertilizer. Maximum increase in these parameters was found with phosphorus application at 100 kg P_2O_5 ha⁻¹. The P concentration was higher at 100 kg P_2O_5 ha⁻¹, while uptake was higher at 75 kg P_2O_5 ha⁻¹.

Venkatesh *et al.* (2002) reported that in maize significant increase in uptake by grain up to 60 kg P_2O_5 ha⁻¹ was observed when applied with or without FYM and lime but its value was maximum at 60 kg P_2O_5 ha⁻¹ when applied with FYM and lime.

Mehta *et al.* (2005) found that the application of P up to 40 kg P_2O_5 ha⁻¹ significantly improved nitrogen and phosphorus uptake by seed and stover over 20 kg P_2O_5 ha⁻¹.

Jaetzold *et al.* (2006) stated that positive influence of phosphorus application on nitrogen uptake in plants is due to the role of phosphorus in cell division of shoot and expanded growth of meristematic tissues or foliage.

Kumar *et al.* (2006) reported an increase on N, P, K uptake in maize due to application of increasing doses of phosphorus fertilizer. Agronomic efficiency in maize was reported to decrease with increasing level of phosphorus doses.

Nekesa (2007) reported a decrease in phosphorus use efficiency with increasing rate of phosphorus.

Ranjit *et al.* (2007) reported that application of different levels of phosphorus influenced nitrogen, phosphorus, potassium and calcium uptake in groundnut significantly where 112.5 kg P_2O_5 ha⁻¹ recorded maximum uptake over 37.5 and 75 kg P_2O_5 ha⁻¹.

Islam *et al.* (2008) studied the effect of phosphorus on nutrient uptake of Japonica rice variety (Nipponbare) and Indica rice variety (IR-28) and it was observed that phosphorus content in the plant and grain significantly increased with phosphorus levels up to 800 kg P_2O_5 ha⁻¹, while P uptake also significantly increased with phosphorus levels though in some cases, variation occurred which may be due to the variation in dry matter production of the two varieties. However, P application at different levels did not show any effect on N and K content and uptake in plant as well as grain.

Sharma *et al.* (2008) stated that increase in nutrient supply in green gram due to phosphorus application is due to its positive effect on better root system.

Hussein (2009) reported that increasing rate of phosphorus application was found to decrease phosphorus use efficiency and phosphorus agronomic efficiency.

Nedunchezhiyan *et al.* (2010) found significantly higher N, P and K uptake by maize grain on application of 32.7 kg P ha⁻¹ over 26.2 kg P ha⁻¹ for three successive years.

Awomi *et al.* (2012) reported that N, P, K uptake in mungbean increased with increasing levels of P, where maximum uptake was recorded at 60 kg P_2O_5 ha⁻¹ which was significantly superior to the lower levels of 20, 40 kg P_2O_5 ha⁻¹ and control.

Rashid and Iqbal (2012) showed that P concentration increased with its application. The maximum P concentration in maize was observed at highest P level of 57 kg P ha⁻¹.

Singh and Singh (2012) reported that application of 75 kg P_2O_5 ha⁻¹ gave higher total nitrogen, phosphorus, potassium and sulphur uptake in pigeonpea, which was at par with 50 kg P_2O_5 ha⁻¹ and significantly superior over 25 kg P_2O_5 ha⁻¹ and control.

Darwesh *et al.* (2013) reported an increase in nutrient uptake (N, P, K) due to phosphorus application and attributed the result to the influence of phosphorus on enhanced root growth, hair length and surface area which increased the nutrient content of plant.

Prajapati *et al.* (2013) reported that that soil applied phosphorus and sulphur increased the nitrogen content in seed and stover of mungbean.

Sentimenla *et al.* (2013) reported an increase in N, P, K and B content in both seed as well as stover of soybean with the application of increasing rates of both phosphorus and boron. The highest agronomic efficiency of phosphorus was recorded with the lowest dose of P application.

Sepat and Rai (2013) reported that response of maize to P application of 90 kg P_2O_5 ha⁻¹ through SSP recorded highest N and P uptake over 45 kg P_2O_5 ha⁻¹ and control.

Kisinyo *et al.* (2014) reported that agronomic phosphorus use efficiency efficiency and phosphorus fertilizer recovery efficiencies decreased with increase in P fertilizer rates.

Nsanzabaganwa *et al.* (2014) reported that application of N up to 240 kg ha⁻¹ and P up to 26.4 kg ha⁻¹ consistently maintained higher levels of N in maize plants over the control at almost all the growth stages. P application at 39.6 kg ha⁻¹ reduced N content than that observed at 26.4 kg P ha⁻¹ at all the growth stages. The concentration of N and P either declined or remained unchanged due to P application beyond 26.4 kg ha⁻¹. Phosphorus application also caused an increase in N uptake, but not at the same magnitude as N application. It led to an increase in N uptake in maize by 22.7, 40.2, and 24.4% due to application of 13.2, 26.4, and 39.6 kg P ha⁻¹, respectively over control. It also resulted in an increase in P uptake by 20.6, 34.0, and 22.7%, respectively with 13.2, 26.4, and 39.6 kg P ha⁻¹ over control.

Kumar *et al.* (2015) reported positive effect of phosphorus application on N, P, K content and uptake in urd bean.

Snehlata (2015) reported that application of 50 kg P_2O_5 ha⁻¹ significantly enhanced N, P and K accumulation by grain and stover of maize over 40 kg P_2O_5 ha⁻¹ and 30 kg P_2O_5 ha⁻¹.

Bak *et al.* (2016) in a field study on maize observed that the form of phosphorus applied as fertilizer showed no significant effect on P contents in

the maize organs, as well as on the total accumulation of this nutrient in the plant.

Kumar *et al.* (2017) reported that application of increasing levels of phosphorus (0, 30, 60 and 90 kg P_2O_5 ha⁻¹) resulted in a significant increase in N, P, K, Ca, Mg and S content of soybean seed where maximum values were observed at 90 kg P_2O_5 ha⁻¹.

Muhlbachova *et al.* (2017) reported a positive effect of P on the plant growth and B uptake by plants. High positive correlations between B content in plants and B uptake was also reported.

Saeed *et al.* (2017) reported higher P contents in plants under elevated P applications where maximum was recorded at 200 kg P ha⁻¹ which declined with the declining P application rates of 150 and 100 kg P ha⁻¹.

Etabo *et al.* (2018) reported that there was no significant effect on plant tissue nitrogen and phosphorus due to phosphorus applications on soils, though increased level of P application consistently increased plant tissue nitrogen and phosphorus in the plant studied. It was observed that application of phosphorous @ 75 kg ha⁻¹ gave highest plant-tissue nitrogen and phosphorus content in rice plant over application of 0, 25 and 50 kg P ha⁻¹.

Uygur and Sen (2018) reported that phosphorous fertilization affected grain P, N and Mn concentration in wheat positively, while it affected the concentration of Ca, Mg, Na, Fe, Cu and Zn negatively. On the other hand, effect of phosphorus was non-significant on K concentration in wheat grains.

Wafula *et al.* (2018) reported that phosphorus application significantly influenced the calcium content of finger millet grains. They reported that application of phosphorus probably enhanced exchange reactions in the soil by releasing hydrogen ions in the microbial biomass that probably resulted in more availability of Ca and hence increased its uptake.

Irfan *et al.* (2019) reported that interactive effect of phosphorus and boron on their concentration in maize was such that application of B level of 2 kg ha⁻¹ resulted in relatively higher P concentration in maize grains and straw at P level of 90 kg ha⁻¹ contrarily to 45 and 135 kg P ha⁻¹. The B concentration in maize grains and straw increased with corresponding addition of B at each P level but at variable rate, with the maximum response at higher P level.

2.3.3 Effect on soil properties

Rekhi *et al.* (2000) reported that application of higher dose of phosphorus with organic manures raised the available phosphorus from 3 mg kg⁻¹ soil to 11.5 mg kg⁻¹ soil.

Okalebo *et al.* (2002) observed that in highly weathered soils of the tropical and subtropical acid soils, the applied P fertilizers readily react with Al and Fe sesquioxides to form sparingly soluble P forms which normally results in very low soil available P for plant absorption.

Venkatesh *et al.* (2002) reported that available P and exchangeable Ca and Mg in post-harvest soil increased due to application of P, FYM and lime. Phosphorus @ 90 kg P_2O_5 ha⁻¹ resulted in reduction of exchangeable Al content of soils which may be due to the formation of relatively stable hydroxy Al-phosphate complexes and their polymerisation.

Weisz *et al.* (2003) reported an increase in P content in soil due to P application.

Fageria *et al.* (2010) attributed the increase of P concentration in plant tissue after liming to 'P spring effect' of lime, where amount of available phosphorus in soil increases because of reduced adsorption-precipitation reaction between Al and P at the root surface and root free space.

Iqbal *et al.* (2010) reported that P can reduce exchangeable acidity by precipitating the Al in solution.

Sharma *et al.* (2011) found that application of P at 0, 30 and 60 kg ha⁻¹ showed no significant change in the soil pH over its initial value. The available N and K were significantly higher at zero level of applied P compared to when P was applied at 30 and 60 kg ha⁻¹. This may be attributed to the reason that removal of N and K was lower at zero level of applied P. However, available P increased significantly with successive levels of applied P registering an increase of 45.7 % and 94.2 % with 30 and 60 kg ha⁻¹ of applied P over zero level of P. A slight increase in exchangeable Ca and K was found with increase in the dose of applied P.

Kisinyo *et al.* (2014) reported that application of P fertilizer increased soil available P. However, higher soil available P was observed in combined application of lime and P fertilizer than when either of them was applied alone.

Kisinyo (2016) reported that application of TSP reduced the soil pH because of the release of phosphoric acid during its dissolution. Phosphate fertilizer had significant effect on soil available P due to release of phosphate ions into the soil solution during its dissolution. However, soil available P was more when lime and P fertilizer were applied in combination than when either of them was applied alone. This was probably because lime reduced P sorption making both the soil native and applied P fertilizer available for plants uptake as well as enhancing root growth.

Opala (2017) reported that only at high rates of P application (100 kg ha^{-1}) with high lime rates of 10 or 20 t ha^{-1} the available P exceeded the critical value of 10 mg kg⁻¹ which has been found to be adequate for maize production.

Goswami *et al.* (2019) observed a significant effect of phosphorus application on soil organic carbon, available N and K and electrical conductivity of soil and a non-significant effect on soil pH. A slight increase in soil organic carbon, available N and K and a slight decrease in electrical conductivity of soil with higher levels of P (118 kg P_2O_5 ha⁻¹) were observed.

2.4. EFFECT OF BORON ON PLANTS AND SOIL PROPERTIES 2.4.1 Effect on growth and yield

Singh *et al.* (2002) observed that higher level of B (1.0 and 2.0 kg ha⁻¹) alone showed advantage in yield but it reduced the yield in presence of lime. Treatment of 0.5 kg B ha⁻¹ + 250 kg lime ha⁻¹ stood second in respect of grain (2555 kg ha⁻¹) and stover (8835 kg ha⁻¹) yields, resulting in 44.8 and 50.3% higher grain and stover yields over the control respectively. Application of boron @ 1.0 kg B ha⁻¹ even in absence of lime was found good enough for the maximum yield of maize grain (2830 kg ha⁻¹).

Rahim *et al.* (2004) reported an increase in number of grains per row and grains per cob in maize due to boron application.

Adiloglu and Adiloglu (2006) found that dry matter yield of maize decreased with B application which may be due to the fact that B uptake of plants are more in Zn deficient soils. Therefore, B toxicity is exposed and the plant growth is adversely affected by this situation.

Ceyhan *et al.* (2007) reported that B application increased the plant height of chickpea by 4 and 5% at 3 kg B ha⁻¹ and 6 kg B ha⁻¹, respectively over control.

Ahmed *et al.* (2008) reported that the growth parameters of cotton increased from 0-5 mg kg⁻¹ B levels, but thereafter reduced sharply in an irrigated arid climatic condition.

Chowdhury *et al.* (2008) assessed the impact of boron on grain yield of diverse wheat genotypes under foothill region of West Bengal. The wheat cultivars irrespective of genotypic variations showed significant positive responses to B application. Among the different methods of boron application, soil application of borax @ 10 kg ha⁻¹ was found to be best for increasing growth and yield parameters, like number of effective tillers, length of panicle, grains/spike and total kernel weight.

Akhter and Mahmud (2009) reported that the yield of maize grain increased significantly due to added boron up to 2.0 kg B ha⁻¹ and yield components like plant height, ear height and straw yield were influenced significantly due to application of boron.

Debnath and Ghosh (2011) conducted a pot culture experiment on rice in Terai Zone soils of West Bengal. B was applied at 0, 0.5, 1.0 and 1.5 mg kg⁻¹ soil. The critical concentration of B in soils and plant was 0.32 and 12.5 mg kg⁻¹, respectively. It was found that B significantly increased the average shoot yield from 5.76 to 10.75 g pot⁻¹ up to 1.5 mg B kg⁻¹ soil below the critical value and 9.99 to 10.29 g pot⁻¹ up to 1 mg B kg⁻¹ soil above the critical value and thereafter the decrement in yield occurred. It was found that application of 1.5 mg B kg⁻¹ can be applied safely for rice in the soils of Terai Zone of West Bengal where the available B is below 0.32 mg kg⁻¹.

Somroo *et al.* (2011) reported that boron application in maize increased the number of green leaves which might be due to the availability of boron at later growth stages of maize.

Tahir *et al.* (2012) observed that, application of boron at the rate of 0.30 kg ha⁻¹ significantly increased plant height, leaf area, stem diameter, cob weight, number of grains per cob and grain yield. However, further increase in boron dose decreased the yields.Muhammad *et al.* (2015) reported that, application of graded boron to maize increase all the agronomic growth

parameters of maize. The increase was achieved with the application of 8 kg boron (granubor).

Gazala *et al.* (2016) observed that the application of boron at different rates in different crops have shown a positive influence on yield and other agronomical attributes in different crops, thereby proving the vital role boron plays in improving yield of different crops.

Saleem *et al.* (2016) reported that height of plant, stem width /girth, green and dry leaves per plant, ear head length and grain yield of maize were significantly affected with the application of 3 kg B ha⁻¹.

Arunkumar and Srinivasa (2018) in a field experiment on sandy loam soil conducted in maize crop observed that application of 5 kg borax ha⁻¹ recorded significantly higher grain yield (89.86 q ha⁻¹), stover yield (160.78 q ha⁻¹) and yield attributing parameters over control and 200 kg gypsum ha⁻¹.

Borase *et al.* (2018) reported that application of the micronutrients boron, iron and zinc along with RDF gave higher growth parameters in *kharif* maize *viz.*, plant height, number of functional leaves and leaf area.

Rahman *et al.* (2018) in a three year experiment revealed that for hybrid maize the highest grain yield was obtained with 2.5 kg B ha⁻¹, which was marginally significant with control but statistically identical to rest of the boron levels. Considering the boron fertilty of soil, response of maize to boron and economic return, the optimum dose of boron was found to be 0.5 kg ha⁻¹ for the cultivation of hybrid maize for medium level boron fertile soil.

Rehim *et al.* (2018) in a field experiment of maize crop observed that grain yield was increased to 65% with combined fertilization of K and B @125 and 16 kg ha⁻¹, respectively, with reference to control. Maximum plant height, cob length and 1000-g weight was observed @ 150 kg K ha⁻¹ and 16 kg B ha⁻¹.

Chanchal *et al.* (2020) in a study on rice-wheat cropping sequence, where wheat was grown after rice in the same pot without fresh application of B, observed that application of RDF + 1.5 kg B ha⁻¹ gave maximum grain and straw yield in rice, whereas in wheat, maximum grain and straw yield was observed in RDF + 3 kg B ha⁻¹. A significant residual effect of B application in rice was noticed even after the harvest of wheat crop particularly at highest doses of B application.

2.4.2 Effect on nutrient composition and uptake

Inal and Tarakcioglu (2001) reported an increase in N concentration of plants when B was applied.

Adiloglu and Adiloglu (2006) narrated that N, P and K concentration in maize increased with increasing rates of B and Zn application. But these increases were not found significant statistically.

Ayden and Sevin (2006) reported that, boron and zinc significantly increases the concentrations of phosphorous and potassium in maize.

Byju *et al.* (2007) reported that fertilizer B recovery of sweet potato was generally higher at the smallest B rate and decreased with further increase in B rate, but the differences were rather small.

Adem *et al.* (2011) suggested that addition of 7.7 kg ha⁻¹ of boron increased levels of N Ca, Mg, P, K and Mn in shoot and leaves tissues of maize, but decrease Fe, Zn and Cu content.

Aref (2011) reported that the use of 3 and 6 kg ha⁻¹ B increased grain N uptake from 129.07 at zero B level to 150.78 and 148.56 kg ha⁻¹, respectively. Highest P uptake by the grain was seen at boron application of 3 kg ha⁻¹ which significantly increased P uptake by the grain from 24.15 to 29.13 kg ha⁻¹, showing an increase of 20.62% as compared with the no B level. The use of 3

and 6 kg ha⁻¹ B, significantly increased K uptake by the grain from 34.86 to 39.5 and 39.06 kg ha⁻¹, respectively (13.31 and 12.05% increase relative to the no B level).

Günes *et al.* (2011) reported that boron fertilizer application in corn increased both leaves and shoot tissue N, Ca, Mg, P, K, and Mn but decreased tissue Fe, Zn, and Cu content, respectively. The 2 years average leaf and shoot tissue B content in the control treatments was 5.59 and 3.63 mg kg⁻¹ dry weight, respectively. This increased to 20.61 for leaf and 13.43 mg B kg⁻¹ for shoot, when B fertilizer was applied at 7.7 kg B ha⁻¹.

Khurana and Arora (2012) reported that increase in B levels from 0.75 to 1.25 kg B ha⁻¹ recorded a decrease in agronomic efficiency 286.7 to 78.4 and 293.3 to 244 kg seed kg⁻¹ B applied through borax in lentil and soybean, respectively.

Chuan *et al.* (2013) reported that agronomic efficiency for a nutrient increased with yield response increasing, but the amount of increase became smaller as the yield response became larger. A lower yield response indicates higher soil indigenous nutrient supply or higher soil fertility, resulting in lower agronomic efficiency. In contrast, a larger yield response means lower soil nutrient supply and relatively higher agronomic efficiency.

Mantovani *et al.* (2013) stated that the mobility of boron within the phloem is low, as a result of which boron fertilization as a function of doses and forms of application is low.

Sahin (2014) reported that boron concentration in maize increased significantly in contrast to where boron fertilization was not applied. This rate of increase was nearly 3 times more. Also with boron fertilization, total nitrogen concentration in maize increased by 10%.

Trautmann *et al.* (2014) stated that susceptibility of boron to leaching in soil may be attributed to its high mobility in soil, high solubility in water and low reactivity with the soil.

Arunkumar and Srinivasa (2018) reported that higher nutrient content and uptake in maize was observed at 5 kg borax ha⁻¹ over control and 200 kg gypsum ha⁻¹ and was at par with, 2.5 kg borax ha⁻¹, 200 kg gypsum + 2.5 kg borax ha⁻¹ and 200 kg gypsum + 5 kg borax ha⁻¹.

Rehim *et al.* (2018) in a field experiment of maize crop observed that concentrations of K in leaf and grains were increased 253% and 322% with combined fertilization (150 kg ha⁻¹ of K and 8 kg ha⁻¹ of B). Fertilization of K and B also increased B concentration in leaf and grains by 179% (150 kg ha⁻¹ of K and 16 kg ha⁻¹ of B) and 370% (125 kg ha⁻¹ of K and 16 kg ha⁻¹ of B).

Shrestha *et al.* (2020) observed that application of B and Zn, either sole or in combination, generally increased the respective nutrient concentration in the biomass of wheat. The application of B fertilizer increased tissue B contents from 5 to 22 mg B kg⁻¹ in wheat.

2.4.3 Effect on soil properties

Saha and Haldar (1998) found that the application of B and lime on a boron deficient Aeric Haplaquept increased the content of available B in soil the extent being 2.85 and 4.29 times over no-B at lower and higher level of B application. The combined effect of lime and B application also showed an increase in the content of extractable B in soils, the extent of increase being 3.3 and 3.5 times over that in the control at lower level of applied B coupled with lower and higher level of lime applied, respectively.

Barman *et al.* (2014) reported that there was no effect of B application on the status of available N, P, K, Ca, Mg, S, Zn, Cu, Fe, and Mn in soil.

Sarkar *et al.* (2015) in a laboratory incubation study on Inceptisols and Alfisols, found that only <50 percent of applied B was recovered from the soils in available form. Such recovery was lower in Alfisols than that in Inceptisols due to adsorption of a greater amount of added B with Fe and Al oxides in the former soil group. Required dose of lime showed an increase in availability of native soil B, particularly in Inceptisols (26%), and a net decrease in recovery of added B (32.5 %) as compared to no lime control (41.6%). The results thus suggested that liming to acidic soils increase extractable B.

Patil *et al.* (2017) observed that soil application of borax showed nonsignificant response on the soil properties; however there was significant increase in available B content in soil. Significantly highest soil available B was recorded where borax was applied at 10 kg ha⁻¹ followed by 7.5 and 5 kg borax ha⁻¹.

Kumar *et al.* (2020) observed that there was no significant effect of different sources and doses of boron on soil pH, EC, OC and available NPK. The different forms of boron increased over its initial status with the application of different sources and doses of boron but results did not vary significantly. While among different doses applied, application of 1.5 kg B ha⁻¹ was observed to be at par with the application of 2.0 kg B ha⁻¹ in increasing the amount of boron in different available forms.

2.5. Tolerance to soil acidity in plants

Mugwira (1980) reported that uptake of nutrients such as Ca, P, Mg, K, Fe, Zn and Cu was reduced in sensitive maize cultivars to soil acidity than in tolerant ones.

Foy (1984) stated that inter- and intra-specific plant differences for tolerance to soil acidity/Al toxicity have been reported. Differences in yield

and nutrient uptake have been related to root development (elongation and absorption), translocation, and shoot demand per unit of nutrient absorbed.

Mendes *et al.* (1985) conducted a field experiment to compare the acid tolerance of 48 commercially grown maize cultivars and reported that on an average, 'tolerant' cultivars were 33% taller than 'intolerant' cultivars in the absence of lime, produced more than double the yield, and had a 60% lower leaf Al content. Significantly, several of the least acid tolerant cultivars were among the best performers in the absence of acidity.

Clark and Mgema (1993) noted that Al generally reduced P, Ca, Mg, Fe, Zn, Mn and Cu uptake in sensitive maize cultivars than in tolerant ones. partly because they have higher efficiency of utilization of absorbed essential nutrients, particularly P.

Malama *et al.* (2000) reported that tolerance to soil acidity is a complex character involving not only tolerance to low pH but also Al toxicity, and in some cases Mn and Fe toxicity and P, Ca and Mg deficiencies. The difference between varieties or species in terms of Al tolerance seems to be positively correlated with differences in P translocation rates in the presence of Al.

Dewi Hayati *et al.* (2014) in a study to check the performance of maize varieties on acid soil tolerance reported that reduction of grain yields in acid soil varied greatly within genotypes and acid soil conditions, ranging from 2.8 to 71%. Check varieties produced high yield on limed soil, but produced low yield on acid soil. It showed higher yield reduction compared to average yield reduction of hybrids.

Adie and Krisnawati (2016) studied the performance of 12 soybean genotypes on three acid soil locations with a pH of 5.87, 5.04, and 4.73, respectively. They reported that among the genotypes, G4AB consistently produced highest yield at pH 5.04 as well as at pH 4.73. Hence, the genotype

G4AB was not only adaptive at low pH but also relatively productive. Soybean genotype adaptive to acid soil was characterized by its ability to maintain the plant height, and followed by a high number of nodules per plant and pod per plant.

Moroni *et al.* (2018) reported that a significant range of differential responses to the acidic soil was shown among the cereals where oat was more tolerant than wheat and barley. Among wheat, there was a wider range of responses (32 to 100%) than barley (27 to 60%).

CHAPTER III

MATERIALS AND METHODS

MATERIALS AND METHODS

Two experiments were conducted in the greenhouse of the Department of Agricultural Chemistry and Soil Science, School of Agricultural Sciences and Rural Development (SASRD), Nagaland University, Medziphema, Nagaland during the *kharif* season of 2016 and 2017 to carry out the investigation entitled, "Study on Acidity Tolerance of Maize and its Response to Phosphorus and Boron". The details of materials used and techniques adopted during the course of investigation are briefly described in this chapter.

3.1. Experimental site

The experimental site lies at 25° 45' 15.95" N latitude and 93° 51' 44.71" E longitude at an elevation of 310 meter above mean sea level.

3.2. Climatic condition

The experimental site lies in humid sub-tropical zone with an average rainfall from 2000-2500 mm annually spread over 6 months *i.e.*, April to September, while the remaining period from October to March remains dry. The mean temperature ranges from 21^oC to 32^oC during summer and rarely goes below 8^oC in winter due to high atmospheric humidity.

3.3. Characteristics of the experimental soil

The soil used for experimentation was collected from two different locations i.e., (i) from Research Farm, Department of Agricultural Chemistry and Soil Science, SASRD (pH 4.6) and (ii) from farmer's field of Medziphema village (pH 5.2). The composite soil sample collected from 0-15 cm depth and mixed well was subjected to analysis for some important physicochemical properties. The results of analysis are presented in Table 3.1.

UOD I	VALUES					
SUIL DADAMETEDS	Soil I Se		Soi	il II	METHODS	
IAKANILIEKS	2016	2017	2016	2017		
Soil pH	4.6	4.6	5.2	5.2	Jackson (1973)	
Lime requirement (t ha ⁻¹)	8.6	8.6	6.6	6.6	Shoemaker et al. (1961)	
EC (dSm ⁻¹)	0.22	0.23	0.20	0.21	Jackson (1973)	
Base Saturation (%)	23.90	24.32	23.78	24.06	1 <i>N</i> NH ₄ OAc at pH 7.0 (Chapman, 1965)	
Organic C (%)	1.52	1.55	1.53	1.56	Rapid titration method by Walkley and Black (Jackson, 1973)	
Available N (kg ha ⁻¹)	237.29	242.20	240.94	242.05	Alkaline permanganate method (Subbiah and Asija, 1956)	
Available P (kg ha ⁻¹)	8.45	8.50	9.50	9.13	Bray P ₁ method (1945)	
Available K (kg ha ⁻¹)	144.48	146.45	150.22	149.20	Ammonium acetate method (Jackson, 1973)	
Exchangeable Ca {cmol (p ⁺) kg ⁻¹ }	2.88	2.85	3.04	2.99	1 <i>N</i> ammonium acetate extracts of soil by titration against EDTA (Black, 1965)	
Available B (mg kg ⁻¹)	0.45	0.40	0.50	0.54	Curcumin method (Dible <i>et al.</i> , 1954)	
Total potential acidity $\{ cmol (p^+) kg^{-1} \}$	15.09	15.03	14.8	14.57	BaCl ₂ - triethanolamine extract buffered at pH 8.0 to 8.2 (Baruah and Barthakur, 1997)	
Exchangeable Al^{3+} {cmol (p ⁺) kg ⁻¹ }	1.63	1.70	1.41	1.44	NaF solution (4%) in 1 N KCl extract titrated against 0.1N HCl (Baruah and Barthakur, 1997)	
$ \begin{array}{c} Exchangeable \ H^{\scriptscriptstyle +} \\ \{cmol \ (p^{\scriptscriptstyle +}) \ kg^{\scriptscriptstyle -1} \} \end{array} $	0.90	0.93	0.64	0.53	Exchangeable H^+ =Exchangeableacidity-Exchangeable Al^{3+}	
Mechanical analysis						
Sand (%)	50.4	48.9	45.4	42.6		
Silt (%)	27.0	25.1	23.7	25.7		
Clay (%)	22.6	26	30.9	31.7	International pipette method	
	Sandy	Sandy	Sandy	Sandy	(Piper, 1966)	
Textural class	clay	clay	clay	clay		
	Ioam	Ioam	Ioam	Ioam		

 Table 3.1: Physicochemical properties of two different experimental soils

3.4. Details of experiments

In order to study the effects of treatments, two pot experiments were carried out in greenhouse of the Department of Agricultural Chemistry and Soil Science during Kharif season of 2016 and 2017. The details of experiments are given below:

3.4.1 Experiment-I: To study the acidity tolerance and lime requirement of maize varieties

Treatments		
i) Soil pH levels	2 [pH 4.6 (pH ₁) and pH 5.2 (pH ₂)]	
ii) Lime levels	4 [Control (L_0), 30 (L_{30}), 60 (L_{60}) and 100 (L_{100}) % of LR]	
iii) Varieties	3 [RCM-75 (V ₁), RCM-76 (V ₂) and RCM-1-1 (V ₃)]	
iv) Crop	Maize	
v) Number of treatment combinations	24	
vi) Number of replications	3	
vii) Experimental design	CRD	
viii) Total number of pots	72	

Experimental procedure:

Earthen pots of 30 cm diameter size were filled with 15 kg of soil. Two type of soils (pH 4.6 and 5.2) was used to develop the pH levels. Lime levels were developed by calcite (CaCO₃). Recommended dose of nitrogen, phosphorus and potassium (100 kg N, 60 kg P₂O₅ and 50 kg K₂O ha⁻¹) were applied through urea, single superphosphate and murate of potash, respectively. Half dose of nitrogen, full dose of phosphorus and full dose of potassium was applied one day before sowing. Remaining half dose of nitrogen was applied 15 days after emergence of crop. Lime was applied 10 days before sowing. The soil was well mixed after applying calculated quantities of fertilizer and lime. Three seeds of maize in each pot were sown as per variety treatments on 28th May, 2016 and 25th May, 2017. After 10 days of

germination, thinning was done and one healthy plant was allowed to grow. Weeding was done to check the weed growth.

3.4.2 Experiment-2: To find out the	P and B requ	irement of maize	under
acidic soil environment			

Treatments	
i) Lime levels	2 [Control (L ₀) and 25 (L ₂₅) % of LR]
ii) Phosphorus levels	4 [Control (P ₀), 30 (P ₃₀), 60 (P ₆₀) and 90 (P ₉₀) kg P ₂ O ₅ ha ⁻¹]
iii) Boron levels	3 [Control (B ₀), 1 (B ₁) and 2 (B ₂) kg B ha ⁻¹]
iv) Crop	Maize
v) Number of treatment combinations	24
vi) Variety	RCM-75
vii) Number of replications	3
viii) Experimental design	CRD
ix) Total number of pots	72

Experimental procedure:

Earthen pots of 30 cm diameter size were filled with 10 kg soil (soil II pH 5.2). Lime, phosphorus and boron levels were developed through calcite (CaCO₃), single superphosphate and borax, respectively. Recommended dose of nitrogen and potassium (100 kg N and 50 kg K₂O ha⁻¹) were applied through urea and murate of potash, respectively. Calculated amount of lime was applied 10 days before sowing. Half dose of nitrogen and full dose of phosphorus, potassium and boron were applied one day before sowing. Remaining half dose of nitrogen was given as top dressing 15 DAS. The soil was mixed properly after fertilizer and lime application. Three seeds of maize in each pot were sown at optimum soil moisture to ensure the germination. Seeds were sown on 28th May, 2016 and 25th May, 2017. Thinning was done 10 days after germination and one plant in each pot was allowed to grow. Time to time weeds was removed from the pots. Standard agronomic practices were adopted during entire crop growing period.

3.4.3 Harvesting and Threshing

The crop was harvested during last week of September (29th of September in 2016 and 25th of September in 2017). The harvested plants from each pot were cleaned and sun dried and weight was recorded for yield data.

3.5. Observations

3.5.1 Plant height

The plant height was measured in centimeter (cm) from the ground level to the top of the plants at 30, 60 DAS and at harvest. The average height of the plant for each treatment was calculated.

3.5.2 Number of leaves per plant

The number of leaves per plant was recorded from each pot at 30, 60 DAS and at harvest. The average number of leaves per plant for each treatment was calculated.

3.5.3 Leaf area index

The leaf area index was recorded at 30, 60 DAS and at harvest. The leaf area index was calculated as the ratio of leaf area to the ground area available to the maize plant. The leaf area was calculated by using the equation given by Montgomery (1911) as,

$LA{=}L\times\!\!W{\times}\,A$

Where, LA, L, W and A is leaf area, leaf length, leaf maximum width and a constant, respectively. The value of the constant A is 0.75.

3.5.4 Cob length (cm)

The length of the cob of maize from each pot was measured in cm.

3.5.5 Cob girth (cm)

The girth of the cob from each maize plant was measured in cm.

3.5.6 Number of rows per cob

The rows of the cob from each maize plant were counted.

3.5.7 Number of grains per row

The grains from the rows of the cob from each maize plant were counted.

3.5.8 Number of grains per cob

The grains of the cob from each pot were counted.

3.5.9 Cob weight (g)

The cob of maize from each pot was weighed in gram.

3.5.10 Test weight (g)

Test weight is the weight of 1000 grains. 100 viable grains were counted from the threshed grains and their weight was recorded which was multiplied by a factor of 10 for each treatment.

3.5.11 Grain yield

After proper sun drying of the grains, the grain yield of each pot was taken on treatment basis and the yield per pot of each treatment was obtained as g pot-¹.

3.5.12 Stover yield

The plant harvested from each pot was sun dried for about a week and their weight was taken and stover yield was obtained by deducting grain yield from total weight of the plant.

3.6. Plant analysis

The grain and stover samples were oven dried at a temperature of 60°C to 70°C to attain a constant weight. The dried seed and stover samples were then powdered and stored in polythene bags for chemical analysis. The powdered seed and stover samples were analyzed for N, P, K, Ca and B content.
3.6.1 Nitrogen

Half a gram powdered sample was digested with concentrated H_2SO_4 in presence of digestion mixture (CuSO₄ + K₂SO₄) till the digest gave clear bluish green colour. The digested sample was further diluted carefully with distill water to known volume. Then a known volume of aliquot was transferred to distillation unit (Micro kjeldahl - apparatus) and liberated ammonia was trapped in boric acid containing mixed indicator. Later it was titrated against standard H_2SO_4 and the amount of ammonia liberated was estimated in the form of nitrogen as per the procedure given by Black (1965).

3.6.2 Boron

For extraction of boron, dry ashing of the ground sample in a muffle furnace was done at 550^oC for 1 h and subsequent extraction with 0.36 N H₂SO₄ (Gaines and Mitchell, 1979). For boron estimation curcumin method as described by Dible *et al.*, 1954 was carried out, where 1 mL of sample aliquot and 4 mL of curcumin solution was evaporated to dryness at $55\pm5^{\circ}$ C. The rosocyanin colour such developed was dissolved in 95% ethanol to make the 25 mL mark in a volumetric flask. Readings were taken at 540 nm in the spectrophotometer within 2 h.

3.6.3 Digestion of plant samples for other nutrients

Half a gram powdered sample was pre-digested with concentrated HNO_3 overnight. Further predigested sample was treated with di-acid (HNO_3 : $HClO_4$ in the ratio 10 : 4) mixture and kept on hot plate for digestion till colourless thread like structures was obtained. After complete digestion precipitate was dissolved in 6*N* HCl and transferred to the 100 mL volumetric flask through Whatman No. 42 filter paper and finally the volume of extract was made to 100 mL with double distilled water and preserved for further analysis.

3.6.3.1 Phosphorus

Phosphorous in the digested sample was determined by vanadomolybdate yellow colour method (Jackson, 1973) by using spectrophotometer at 470 nm.

3.6.3.2 Potassium

The potassium content in the digested sample was determined by flame photometer after making appropriate dilution as described by Chapman and Pratt (1961).

3.6.3.3 Calcium

Calcium was determined in the di-acid digest of plant sample by versenate (EDTA) method (Prasad, 1998).

3.7. Nutrient uptake

The uptake values of nitrogen, phosphorus, potassium, calcium and boron by maize were calculated by using the nutrient content (%) in plant and corresponding yield. The uptake values of nutrients were calculated using the following relationship.

Nutrient uptake (mg pot⁻¹) = Nutrient content (%) × yield (g pot⁻¹) ×10

3.8. Soil analysis

Soil samples were collected from each pot after harvest of the crop. The soil samples were air dried in shade, ground using mortar and pestle and sieved through 2 mm sieve and stored in polythene bags with proper labeling for the analysis of various soil parameters using standard methods as mentioned below.

3.8.1 Soil pH

The soil pH was determined in 1:2.5 water suspensions and analyzed using Glass electrode pH meter (Jackson, 1973).

3.8.2 Electrical conductivity (EC)

The electrical conductivity (EC) was determined using Conductivity Bridge (Jackson, 1973).

3.8.3 Percent base saturation

Base saturation is the percentage of total CEC occupied by Ca^{2+} , Mg^{2+} , K^+ and Na^+ . The base saturation was worked out by using the formula given below.

Base saturation (%) = $[(Ca^{2+} + Mg^{2+} + K^{+} + Na^{+}/CEC] \times 100$

3.8.4 Organic carbon

Organic carbon content in soil was determined by rapid titration method outlined by Walkley and Black (1934) and expressed in g kg⁻¹ as described by Jackson (1973).

3.8.5 Available nitrogen

The available nitrogen in soil was determined using alkaline potassium permanganate method as described by Subbiah and Asija (1956).

3.8.6 Available phosphorus

Available phosphorus was extracted with 0.03N NH₄F in 0.025N HCl solution. The procedure is primarily meant for soils which are moderate to strongly acidic pH and determined by Brays and Kurtz method (1945).

3.8.7 Available potassium

The available potassium content in soil was extracted with neutral normal ammonium acetate (pH 7.0). The potassium content in the extract was determined by flame photometre (Jackson, 1973).

3.8.8 Exchangeable calcium

The exchangeable calcium in soil was determined by versenate method, where a known volume of soil extract was titrated with standard 0.01N versenate (EDTA, ethylene diamine tetra acetic acid disodium salt) solution using murexide (ammonium purpurate) indicator in the presence of NaOH solution (Black, 1965).

3.8.9 Available boron

The available boron in soil was determined by curcumin method as described by Dible *et al.* (1954). Boron forms a reddish brown complex withcurcumin, a phenol dye. After evaporation to dryness, the B-curcumin complex was determined in ethanol at a wavelength of 540 nm in spectrophotometer.

3.8.10 Total potential acidity

The total potential acidity of soil includes all the acidity components like extractable acidity, non-exchangeable acidity, weak acidic carboxylic and phenolic hydroxyl groups of soil organic matter and partially neutralised hydroxyl Al polymers that could be present even in soils of pH >7. The total potential acidity was determined by using BaCl₂- triethanolamine extract buffered at pH 8.0 to 8.2 as described by Baruah and Barthakur (1997).

3.8.11 Exchangeable Al³⁺

The exchangeable Al^{3+} was determined by adding 5 ml of NaF solution (4%) in 1*N* KCl extract. This solution was then titrated against 0.1*N* HCl solution until the pink colour disappeared (Baruah and Barthakur, 1997).

3.8.12 Exchangeable H⁺

The exchangeable H^+ was estimated by the difference between exchangeable acidity and exchangeable Al^{3+} .

Exchangeable H^+ = Exchangeable acidity – Exchangeable Al^{3+}

3.9. Agronomic efficiency

Agronomic efficiency is calculated in units of yield increase per unit of nutrient applied. It more closely reflects the direct production impact of an applied fertilizer and relates directly to economic return. It is expressed in kg kg^{-1} or g g^{-1} .

It is calculated using the formula as follows:

Agronomic efficiency
$$=\frac{Y - Y_0}{F}$$

Where, Y = Yield in nutrient applied plot

 Y_0 = Yield in control plot

F = Amount of nutrient applied

3.10. Nutrient use efficiency (NUE)

The nutrient use efficiency (NUE) is a term used to indicate the relative balance between the amount of fertilizer taken up and used by the crop *versus* the amount of fertilizer lost.

It is calculated using the formula as follows:

NUE (%) =
$$\frac{U - U_0}{F} \ge 100$$

Where, U = Nutrient uptake in nutrient applied plot

 U_0 = Nutrient uptake in control plot

F = Amount of nutrient applied

3.11. Analysis of data

The collected data was processed, classified, tabulated and systematically and statistically analysed by applying the techniques of analysis of variance and the significant of different source of variations was tested by '*F*' test (Gomez and Gomez, 1984).

CHAPTER IV

RESULTS AND DISCUSSION

RESULTS AND DISCUSSION

The results pertaining to "Study on Acidity Tolerance of Maize and its Response to Phosphorus and Boron" carried out during 2016 and 2017 are presented in this chapter. The performance of the crop under various treatments is illustrated by the use of tables and graphs incorporated at appropriate places. The data recorded were analyzed and significant variations have been discussed.

4.1. EFFECT OF SOIL PH, LIME AND VARIETIES ON THE PERFORMANCE OF MAIZE AND SOIL PROPERTIES (EXPT-I)

4.1.1 Effect of soil pH, lime and varieties on the performance of maize4.1.1.1 Effect on plant height

It is evident from Table 4.1.a and Fig. 1 that there was an appreciable increase in plant height with the advancement of days and also a significant difference among the treatments. Irrespective of treatments the pooled plant height at 30, 60 DAS and at harvest varied from 39.70 to 49.36 cm, 153.47 to 188.48 cm and 187.43 to 228.66 cm, respectively. In the year 2016 as well as 2017, at all stages of crop growth, it can be observed that maximum plant height was recorded in pH₂ soil (pH 5.2), while minimum plant height was recorded in pH₁ soil (pH 4.6). Thus, reduced plant height was observed in the lower soil pH (pH 4.6) which may be due to high solubility of aluminium to levels that are toxic which restricts the growth of root systems and in turn plant growth (Matsumoto, 2000).

It can be observed from Table 4.1.a and Fig. 1 that lime had a significant positive effect on plant height at all growth stages. It is apparent that in the year 2016 as well as 2017, the maximum plant height at 30, 60 DAS

	Plant height (cm)								
Treatment		At 30 DAS			At 60 DAS			At harvest	
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Soil pH									
pH_1	44.95	43.67	44.31	166.02	172.30	169.16	209.04	216.36	212.70
pH_2	47.40	48.90	48.15	181.26	187.71	184.49	213.09	221.68	217.39
SEm±	0.49	0.41	0.32	0.75	0.82	0.55	0.80	0.82	0.57
CD (P=0.05)	1.40	1.16	0.90	2.14	2.32	1.56	2.29	2.34	1.61
Lime levels									
L ₀	39.00	40.40	39.70	150.27	156.66	153.47	183.44	191.41	187.43
L ₃₀	46.47	47.41	46.94	175.99	181.91	178.95	213.58	220.54	217.06
L_{60}	48.88	48.96	48.92	183.22	189.57	186.39	222.68	231.37	227.03
L ₁₀₀	50.35	48.37	49.36	185.09	191.87	188.48	224.56	232.76	228.66
SEm±	0.70	0.58	0.45	1.06	1.16	0.78	1.14	1.16	0.81
CD (P=0.05)	1.98	1.65	1.27	3.02	3.28	2.20	3.23	3.30	2.28
Varieties									
\mathbf{V}_1	43.84	43.84	43.84	168.58	175.97	172.27	205.06	214.13	209.60
V_2	48.93	49.02	48.97	177.22	185.01	181.12	215.86	223.78	219.82
V ₃	45.76	46.00	45.88	175.13	179.03	177.08	212.27	219.15	215.71
SEm±	0.60	0.50	0.39	0.92	1.00	0.68	0.98	1.01	0.70
CD (P=0.05)	1.71	1.43	1.10	2.62	2.84	1.91	2.80	2.86	1.98

Table 4.1.a: Plant height of maize at different growth stages as affected by soil pH, lime and varieties



Fig 1: Effect of soil pH, lime and varieties on plant height of maize at 30, 60 DAS and at harvest

and at harvest was recorded in the treatment receiving full dose of lime i.e., at L_{100} . Treatment L_{100} , however, was found to be at par with L_{60} in all growth stages. The minimum plant height at all growth stages *i.e.*, 30, 60 DAS and at harvest was recorded in control *i.e.*, L_0 . At harvest, application of 100% lime enhanced the pooled plant height by 22% over control. The significant increase in maize growth at higher dose of lime may be attributed to decrease in aluminium toxicity which is found to inhibit root growth due to reduction in cell elongation and division (Foy, 1988). Liming decreases Al toxicity by precipitating soluble and exchangeable Al as positively charged monomeric AlOH²⁺ and Al(OH)₂⁺ species which may polymerize to form both large and small positively charged polynuclear complexes. The positively charged polynuclear sorbed to clay mineral and organic matter surfaces around the root zone (Stol *et al.*, 1976), which improves the ability of plants to explore the soil volume for nutrients and water.

From the data depicted in Table 4.1.a and Fig. 1, the results revealed that in both years at all growth stages, maximum plant height was recorded in the variety V₂ (RCM-76), while minimum plant height was recorded in the variety V₁ (RCM-75). Variation in plant height of varieties was significant at all growth stages during both the years. At harvest, pooled plant height of RCM-76 was 4.8 and 1.9% higher than RCM-75 and RCM-1-1, respectively. On the basis of pooled plant height at harvest, superiority of varieties was recorded in the order of RCM-76 > RCM-1-1 > RCM-75. The reason for differences in plant height among the varieties may be due to the variation in the genetic makeup of these varieties.

Interaction effect of pH and lime on plant height

It can be observed from Table 4.1.b that in the year 2016 as well as 2017, the effect of pH and lime had a significant effect on plant height at 30 DAS which showed an increasing trend with increase in lime levels, irrespective of soil pH levels. Higher soil pH level along with higher lime

		Plant height at 30 DAS (cm)						
Soil pH levels		Lime	levels					
	L ₀	L ₃₀	L ₆₀	L ₁₀₀				
		20	016					
pH_1	37.70	45.59	46.01	50.49				
pH_2	40.30	47.34	51.74	50.21				
SEm±	0.98							
CD (P=0.05)	2.80							
	2017							
pH_1	36.70	43.83	46.86	47.29				
pH_2	44.10	50.99	51.06	49.46				
SEm±		0.	82					
CD (P=0.05)		2.	33					
		Poo	bled					
pH_1	37.20	44.71	46.43	48.89				
pH ₂	42.20	49.17	51.40	49.83				
SEm±	0.64							
CD (P=0.05)		1.	80					

Table 4.1.b: Interaction effect between pH and lime on plant height at 30 DAS

levels yielded in higher plant height, which may be attributed to the fact that at higher soil pH coupled with lime application, the soil reached the desirable neutral pH and also decreased the solubility of aluminium ion, which thus favoured the growth of maize. It could be observed that at pH 4.6 (pH₁), maximum plant height was recorded at 100% lime of LR (L₁₀₀), whereas at pH 5.2 (pH₂), maximum plant height was recorded at 60% lime of LR (L₆₀). This observation is in line with the findings of Muindi *et al.* (2015) who reported a similar trend in the plant height of maize due to liming grown in two types of soil - extremely acidic and strongly acidic soil. However, further examination of the pooled data elucidated that in pH₂ soil, plant height at L₆₀ was at par with L₁₀₀. Conversely, minimum plant height in both pH was observed in L₀. This may be due to the fact that at lower pH, there is higher concentration of aluminium ions in exchangeable forms, thus hampering crop growth.

4.1.1.2 Effect on number of leaves

From Table 4.2, it can be clearly seen that the effect of pH on number of leaves was non-significant in the year 2016 as well as 2017. However, it can be observed that there was a noticeable increase in the number of leaves with the advancement of days.

It is apparent from Table 4.2 that lime had a significant effect on the number of leaves. The number of leaves at 30 and 60 DAS was found to be highest at 100% lime of LR (L_{100}) during 2016 as well as 2017, and treatment L_{100} was at par with treatment L_{60} . On the other hand, the number of leaves at harvest was highest at 60% lime of LR (L_{60}) in both the year 2016 and 2017. Pooled data further elucidated that treatment L_{60} was at par with treatment L_{100} . The lowest number of leaves in all the growth stages was observed in control (L_0) during 2016 and 2017. Thus, the significant positive effect of liming on number of leaves may be due to liming, the soil pH increased towards neutral condition which increased the plant nutrient availability in the soil and resulted

	Number of leaves plant ⁻¹								
Treatment		At 30 DAS			At 60 DAS			At harvest	
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels									
pH_1	6.92	7.11	7.01	9.94	10.17	10.06	13.00	13.33	13.17
pH ₂	7.00	7.25	7.13	10.11	10.39	10.25	13.22	13.53	13.38
SEm±	0.08	0.09	0.06	0.08	0.09	0.06	0.09	0.07	0.06
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS
Lime levels									
L ₀	6.50	6.78	6.64	9.50	9.89	9.69	12.67	13.11	12.89
L ₃₀	6.56	7.11	6.83	9.83	10.11	9.97	13.17	13.39	13.28
L_{60}	7.33	7.33	7.33	10.33	10.50	10.42	13.33	13.78	13.56
L ₁₀₀	7.44	7.50	7.47	10.44	10.61	10.53	13.28	13.44	13.36
SEm±	0.11	0.13	0.08	0.11	0.12	0.08	0.13	0.10	0.08
CD (P=0.05)	0.31	0.36	0.23	0.31	0.35	0.23	0.36	0.28	0.23
Varieties									
\mathbf{V}_1	6.67	6.92	6.79	9.63	9.92	9.77	12.83	13.25	13.04
V_2	7.25	7.42	7.33	10.42	10.63	10.52	13.42	13.58	13.50
V ₃	6.96	7.21	7.08	10.04	10.29	10.17	13.08	13.46	13.27
SEm±	0.09	0.11	0.07	0.09	0.11	0.07	0.11	0.09	0.07
CD (P=0.05)	0.26	0.31	0.20	0.26	0.31	0.20	0.31	0.25	0.20

Table 4.2: Number of leaves per plant of maize at different growth stages as affected by soil pH, lime and varieties

in absorption of more nutrients by plants and ultimately enhanced the plant growth.

From the data (Table 4.2) it can be inferred that significant differences existed among the varieties with regard to leaves per plant and the highest number of leaves was recorded in the variety RCM-76 (V₂), while the minimum number of leaves was recorded in treatment RCM-75 (V₁) during 2016 and 2017. It may also be inferred here that the reason for differences in number of leaves among the varieties may be due to the variation in their genetic makeup.

4.1.1.3 Effect on leaf area index

The results on the leaf area index in different treatments have been presented in Table 4.3. The effect of pH on leaf area index was non-significant at all crop stages during both the years of experimentation.

It is apparent from Table 4.3 that lime did not have any significant effect on the leaf area index at 30 DAS. However, at 60 DAS and at harvest, it could be observed that lime had a significant positive effect on leaf area index. The maximum leaf area index at 60 DAS as well as at harvest was recorded at 60% lime of LR (L_{60}), while minimum leaf area index in both crop stages was recorded in control (L_0) during 2016 and 2017. At 60 DAS, treatment L_{60} was at par with L_{100} and at harvest, treatment L_{60} was at par with L_{30} and L_{100} . Higher value of leaf area index was recorded at 60 DAS growth stage and thereafter it declined at harvest due to drying of leaves. In general, the application of lime was observed to boost the growth of the crop due to increase in the soil pH. Similar results have also been reported by Kumar *et al.* (2014); Muindi *et al.* (2015) and Singh *et al.* (2016) in various crops, who all reported a significant increase in leaf area index with lime application.

From table 4.3, it is clear that varieties did not have any significant effect on the leaf area index of maize at any growth stage.

	Leaf area index								
Treatment		At 30 DAS			At 60 DAS			At harvest	
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels									
pH_1	0.61	0.64	0.63	1.30	1.37	1.33	1.17	1.21	1.19
pH_2	0.63	0.69	0.66	1.32	1.36	1.34	1.18	1.21	1.19
SEm±	0.01	0.02	0.01	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	NS	NS	0.04	NS	NS	NS	NS	NS	NS
Lime levels									
L ₀	0.59	0.63	0.61	1.28	1.33	1.31	1.14	1.18	1.16
L ₃₀	0.60	0.66	0.63	1.29	1.35	1.32	1.17	1.21	1.19
L_{60}	0.65	0.68	0.67	1.34	1.40	1.37	1.21	1.23	1.22
L ₁₀₀	0.64	0.70	0.67	1.32	1.38	1.35	1.18	1.20	1.19
SEm±	0.02	0.03	0.02	0.02	0.02	0.01	0.02	0.01	0.01
CD (P=0.05)	NS	NS	NS	0.04	0.05	0.03	0.04	0.03	0.03
Varieties									
\mathbf{V}_1	0.60	0.63	0.61	1.31	1.36	1.33	1.18	1.22	1.20
V_2	0.64	0.69	0.66	1.33	1.39	1.36	1.18	1.21	1.19
V ₃	0.62	0.68	0.65	1.29	1.35	1.32	1.17	1.20	1.18
SEm±	0.02	0.03	0.02	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS

Table 4.3: Leaf area index of maize at different growth stages as affected by soil pH, lime and varieties

4.1.1.4 Effect on cob length, cob girth and cob weight of maize

It is apparent from Table 4.4 that pH was observed to have a significant effect on cob length, cob girth and cob weight of maize. The maximum cob length, cob girth and cob weight was recorded in pH 5.2 (pH₂) soil while the minimum was recorded in pH 4.6 (pH₁) soil during 2016 and 2017. Maize grows best if the soil is maintained at an optimal pH and crop performance is affected indirectly by low soil pH from aluminium and manganese toxicity resulting from overly acidic conditions of the soil (Mallarino, 2011 and Kai *et al.*, 2012).

The cob length, cob girth and cob weight of maize was observed to be significantly influenced by different lime levels. In the year 2016 as well as 2017, the maximum cob length (18.22 and 18.48 cm) was recorded at 60% lime of LR (L_{60}), whereas maximum cob girth (14.91 cm and 14.88 cm) and cob weight (92.39 g and 94.67 g) was recorded at 100% lime of LR (L_{100}). In case of cob length and cob weight, treatment L_{100} was at par with L_{60} and in case of cob girth, treatment L_{100} was at par with L_{30} and L_{60} in both the years. On the other hand, minimum cob length, cob girth and cob weight was observed in control (L_0) during 2016 as well as 2017. Thus, application of lime produced significantly higher yield attributes which could be ascribed to higher photosynthesis and better translocation to the fruiting sink due to liming. Similar lines of findings where application of lime increased these yield attributes have been noted by Brajendra *et al.* (2006); Kumar *et al.* (2012) and Kumar (2015).

From the data presented in Table 4.4, it could be inferred that the maximum cob length, cob girth and cob weight of maize was recorded in variety RCM-76 (V₂) in both the years. Minimum cob length and cob weight was recorded in RCM-1-1 (V₃) and minimum cob girth was recorded in RCM-75 (V₁) during 2016 and 2017. A critical examination of data shows that pooled cob weight of V₂ variety (RCM-76) was 1.02 and 7.2% higher than V₁

64

Treatment	Cob length (cm)			Cob girth (cm)			Cob weight (g)		
Treatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels									
pH_1	17.69	18.04	17.86	14.56	14.53	14.54	81.86	83.03	82.44
pH_2	18.14	18.45	18.29	14.93	14.85	14.89	89.33	91.01	90.17
SEm±	0.05	0.05	0.04	0.08	0.06	0.05	0.61	0.72	0.47
CD (P=0.05)	0.15	0.14	0.10	0.21	0.16	0.13	1.73	2.04	1.32
Lime levels									
L_0	17.35	17.78	17.57	14.33	14.29	14.31	71.61	72.75	72.18
L ₃₀	17.90	18.27	18.08	14.86	14.78	14.82	86.48	88.40	87.44
L_{60}	18.22	18.48	18.35	14.89	14.80	14.85	91.90	92.25	92.07
L_{100}	18.17	18.45	18.31	14.91	14.88	14.90	92.39	94.67	93.53
SEm±	0.08	0.07	0.05	0.11	0.08	0.07	0.86	1.01	0.66
CD (P=0.05)	0.22	0.20	0.14	0.30	0.22	0.19	2.45	2.88	1.86
Varieties									
\mathbf{V}_1	17.91	18.26	18.09	14.57	14.54	14.55	86.71	88.70	87.70
V_2	18.10	18.35	18.23	14.90	14.86	14.88	88.22	88.98	88.60
V_3	17.72	18.12	17.92	14.77	14.68	14.72	81.86	83.38	82.62
SEm±	0.07	0.06	0.04	0.09	0.07	0.06	0.75	0.88	0.58
CD (P=0.05)	0.19	0.17	0.13	0.26	0.19	0.16	2.12	2.49	1.61

Table 4.4: Cob length, cob girth and cob weight of maize as affected by soil pH, lime and varieties

and V_3 varieties, respectively. Varietal characteristics might be the reason for the difference.

4.1.1.5 Effect on number of grains per row, number of rows and grains per cob

From Table 4.5.a it is apparent that effect of pH on number of rows per cob was non-significant during both the years. However, an increase in number of rows per cob was observed in the higher pH of 5.2 (pH₂). The pH had a fairly significant influence on the number of grains per row and a greater significant influence on number of grains per cob. The maximum number of grains per row (23.29) and number of grains per cob (347.57) was recorded in pH 5.2 (pH₂) soil. The minimum number of grains per row (20.89) and number of grains per cob (305.32) was recorded in pH 4.6 (pH₁) soil. Soil reaction accounts for an essential portion of the environmental influence on growth and development of maize (Mallarino, 2011). The pH 4.6 being strongly acidic in reaction results in aluminium toxicity which may have ultimately affected the yield components.

It is apparent from Table 4.5.a that the maximum number of grains per row (22.75) was recorded with application of 60% lime of LR (L_{60}), while number of rows per cob (15.14) and number of grains per cob (355.89) was recorded with application of 100% lime of LR (L_{100}). The minimum number of grains per row, number of rows per cob and number of grains per cob was recorded in control (L_0) in both the years. Application of 60% lime of LR enhanced pooled number of grains per row to the extent of 8.0% over control, while application of 100% lime of LR enhanced number of rows per cob and grains per cob to the extent of 9.0 and 27.0%, respectively over control. A critical examination of data indicates that in case of pooled values of grains per row, 60% lime of LR was at par with 100% lime of LR and in case of pooled values of rows per cob 100% lime of LR was at par with 60% lime of LR. The positive significant effect of lime on these yield attributes can be ascribed to

the higher supply of nutrients particularly NPK due to liming and their subsequent increase in uptake. These results are in accordance with those of Chatterjee *et al.* (2005) and Kumar (2015) who reported increase in yield attributes in various crops with application of lime.

The data (Table 4.5.a) indicated that varieties showed significant influence on number of grains per row, number of rows per cob and number of grains per cob. Among the varieties, the RCM-76 variety produced significantly higher grains per row, rows per cob and grains per cob in comparison to RCM-75 and RCM-1-1. The RCM-76 variety increased pooled grains per row by 2.0% and 5.3%, rows per cob by 1.5 and 4.2% and grains per cob by 4.1 and 10.3% over RCM-75 and RCM-1-1, respectively.

Interaction effect of pH and lime on number of rows per cob

From Table 4.5.b, it can be observed that number of rows per cob enhanced with increase in lime levels in both the pH levels. It was observed that in pH 4.6 soil, maximum number of rows was recorded at 100% lime (pH_1L_{100}) and in pH 5.2 soil, maximum number of rows was recorded with 60% lime of LR (pH_2L_{60}) during both the years. It can thus be observed that in both the pH, lime had a positive effect on number of rows per cob with the maximum at L_{100} in case of pH₁ and at L_{60} in case of pH₂ which however was at par with L_{30} and L_{60} . This indicates that liming even up to 30% lime of LR in case of pH₂ was sufficient to obtain the beneficial effect.

Interaction effect of pH and varieties on number of grains per row

From the data presented in Table 4.5.c, it is apparent that effect of pH and varieties on number of grains per row was significant during 2016. It can be observed from the data that highest number of grains per row was in pH 5.2 with variety RCM-76 (pH_2V_2) and lowest number of grains per row was in pH 4.6 with variety RCM-1-1 (pH_1V_3). Further, it can also be observed that in both the pH, variety RCM-76 (V_2) gave the highest response, while RCM-1-1 gave

Treatment	No of grains per row			No of rows per cob			No of grains per cob		
Treatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels									
pH_1	20.86	20.92	20.89	14.56	14.89	14.72	301.44	309.19	305.32
pH_2	23.25	23.33	23.29	14.69	14.83	14.76	341.92	353.22	347.57
SEm±	0.10	0.12	0.08	0.08	0.09	0.06	4.42	4.19	3.05
CD (P=0.05)	0.27	0.35	0.22	NS	NS	NS	12.57	11.92	8.55
Lime levels									
L_0	21.00	21.11	21.06	13.83	13.94	13.89	276.39	284.00	280.19
L ₃₀	22.17	21.94	22.06	14.83	14.89	14.86	321.22	334.28	327.75
L_{60}	22.67	22.83	22.75	14.89	15.28	15.08	338.33	345.56	341.94
L_{100}	22.39	22.61	22.50	14.94	15.33	15.14	350.78	361.00	355.89
SEm±	0.14	0.18	0.11	0.12	0.13	0.09	6.25	5.93	4.31
CD (P=0.05)	0.39	0.50	0.31	0.34	0.36	0.24	17.78	16.86	12.10
Varieties									
\mathbf{V}_1	22.25	22.08	22.17	14.71	14.88	14.79	323.50	332.50	328.00
\mathbf{V}_2	22.46	22.79	22.63	14.88	15.17	15.02	337.04	346.33	341.69
V_3	21.46	21.50	21.48	14.29	14.54	14.42	304.50	314.79	309.65
SEm±	0.12	0.15	0.10	0.10	0.11	0.08	5.42	5.14	3.73
CD (P=0.05)	0.34	0.43	0.27	0.29	0.31	0.21	15.40	14.60	10.48

Table 4.5.a: Number of grains per row, number of rows per cob and number of grains per cob as affected by soil pH, lime and varieties

		Number of r	ows per cob						
Soil pH levels	Lime levels								
	L_0	L ₃₀	L ₆₀	L ₁₀₀					
		2016							
pH_1	13.67	14.78	14.56	15.22					
pH_2	14.00	14.89	15.22	14.67					
SEm±		0.17							
CD (P=0.05)	0.47								
	2017								
pH_1	13.67	14.78	15.44	15.67					
pH_2	14.22	15.00	15.11	15.00					
SEm±		0.1	18						
CD (P=0.05)		0.4	51						
		Poo	oled						
pH_1	13.67	14.78	15.00	15.44					
pH_2	14.11	14.94	15.17	14.83					
SEm±	0.12								
CD (P=0.05)		0.3	34						

Table 4.5.b: Interaction effect between pH and lime on number of rows per cob

Table 4.5.c: Interaction effect between pH and varieties on number of grains per row

	Number of grains per row						
		Varieties					
Soli pH levels	V1	V_2	V ₃				
		2016					
pH_1	21.25	21.42	19.92				
pH_2	23.25	23.50	23.00				
SEm±		0.17					
CD (P=0.05)		0.47					
		Pooled					
pH_1	21.08	21.58	20.00				
pH_2	23.25	23.67	22.96				
SEm±		0.14					
CD (P=0.05)		0.38					

the lowest response in terms of number of grains per row. Also, all the three varieties performed better in pH 5.2 than in pH 4.6.

4.1.1.6 Effect on test weight, grain yield and stover yield of maize

From Table 4.6 and Fig. 2 it is apparent that the effect of pH on test weight, grain yield as well as stover yield of maize was found to be significant in both the years 2016 and 2017. Irrespective of treatments and years, the test weight varied from 23.32 g to 24.88 g, grain yield of maize varied from 56.79 g pot⁻¹ to 73.10 g pot⁻¹ and stover yield from 108.85 g pot⁻¹ to 134.64 g pot⁻¹. Between the two soil pH, the highest test weight, grain yield and stover yield was observed in pH 5.2 (pH₂). From pooled data, it can be observed that grain and stover yield increased by 13.3% and 9.6%, respectively in pH 5.2 (pH₂) soil as compared to pH 4.6 (pH₁) soil. The reason for lower yields at soil pH 5.2.

It can be observed from the Table 4.6 and Fig. 2 that test weight, grain yield as well as stover yield exhibited an increasing trend with increase in lime application. Lime application significantly increased the yield which is usually associated with significant reduction in exchangeable Al. Maximum test weight, grain yield and stover yield was observed at 100% lime of LR (L_{100}) during 2016 and 2017. A critical examination of data revealed that each increasing level of lime up to L_{60} (60% lime of LR) enhanced the grain and stover yield significantly in comparison to preceding lower level of lime. However, beyond L_{60} level, effect of lime application was at par. Hence, L_{60} (60% lime of LR) proved to be the optimum dose of lime for getting better yield of maize. The L_{60} level of lime enhanced the grain yield by 28.5% and stover yield by 21.3% over control. The lowest values for all the three parameters were recorded in treatment L_0 in both the years. In case of test L_{60} weight, treatments L_{30} , and L_{100} were found to be

Trastmont	Test weight (g)			Grain yield (g pot ⁻¹)			Stover yield (g pot ⁻¹)		
Treatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels									
pH_1	24.02	24.14	24.08	64.09	63.76	63.92	119.03	121.56	120.29
pH_2	24.61	24.78	24.69	71.76	73.13	72.45	130.39	133.40	131.89
SEm±	0.09	0.15	0.09	0.38	0.38	0.27	1.14	1.12	0.80
CD (P=0.05)	0.27	0.43	0.25	1.09	1.08	0.76	3.24	3.17	2.24
Lime levels									
L_0	23.23	23.42	23.32	56.51	57.07	56.79	107.31	110.39	108.85
L ₃₀	24.43	24.69	24.56	70.10	69.58	69.84	127.51	130.14	128.83
L_{60}	24.76	24.80	24.78	72.73	73.30	73.01	130.93	133.19	132.06
L ₁₀₀	24.85	24.91	24.88	72.37	73.82	73.10	133.09	136.19	134.64
SEm±	0.13	0.21	0.13	0.54	0.54	0.38	1.61	1.58	1.13
CD (P=0.05)	0.38	0.61	0.35	1.55	1.53	1.07	4.59	4.49	3.17
Varieties									
\mathbf{V}_1	24.39	24.41	24.40	68.46	69.10	68.78	125.51	128.31	126.91
V_2	24.43	24.63	24.53	70.72	70.93	70.82	127.53	130.89	129.21
V_3	24.14	24.33	24.23	64.61	65.29	64.95	121.10	123.24	122.17
SEm±	0.12	0.19	0.11	0.47	0.46	0.33	1.40	1.37	0.98
CD (P=0.05)	NS	NS	NS	1.34	1.32	0.93	3.97	3.89	2.74

Table 4.6: Test weight, grain yield and stover yield of maize as affected by soil pH, lime and varieties



Fig 2: Effect of soil pH, lime and varieties on grain yield and stover yield of maize

at par with each other. The lowest yield consistently recorded in control treatment (L_0) which may be due to soil acidity in the control pot, which consequently resulted in aluminium toxicity and deficiency of the exchangeable bases or cations which are unavailable in acid soils (Voor, 2012). Liming is an important practice to achieve optimal yields of crops in acid soils. Application of lime at a suitable rate in acid soils brings about several chemical and biological changes in the soils, which is beneficial for improving the yield of crops (Fageria and Baligar, 2008). These results are in congruence with those of Chatterjee *et al.* (2005); Dixit (2006) and Kumar *et al.* (2012).

The effect of variety on test weight was found to be non- significant, while significant response was observed on grain and stover yield. As seen in Table 4.6 and Fig. 2, variety RCM-76 (V₂) gave the highest grain yield as well as stover yield and RCM-1-1 (V₃) gave the lowest grain yield as well as stover yield. It was observed that variety V₂ (RCM-76) gave 3.0% and 9.0% higher grain yield over V₁ and V₃, respectively. It was also observed that V₁ variety (RCM-75) produced 5.9% higher yield over V₃ variety (RCM-1-1). On the basis of pooled grain yield, order of superiority of varieties may be arranged as RCM-76 > RCM-75 > RCM-1-1.

4.1.1.7 Nitrogen content in grain and stover

As can be seen from Table 4.7, effect of pH on nitrogen content in grain as well as stover was in both the years non-significant.

It can be observed from Table 4.7 that effect of lime on nitrogen content in both grain and stover was significant during the two years of experimentation. The highest nitrogen content in both grain and stover during 2016 and 2017 was recorded in the treatment L_{100} and lowest was recorded in L_0 . Pooled data reflected an increase in nitrogen content by 27.4% and 23.0%, respectively in grain and stover in L_{100} over L_0 . The substantial improvement in

Treatment	N co	ontent in grai	n (%)	N content in stover (%)			
	2016	2017	Pooled	2016	2017	Pooled	
Soil pH levels							
pH_1	1.17	1.20	1.19	0.66	0.69	0.67	
pH_2	1.20	1.22	1.21	0.67	0.70	0.69	
SEm±	0.01	0.01	0.01	0.01	0.01	0.01	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	
Lime levels							
L_0	1.02	1.02	1.02	0.60	0.63	0.61	
L_{30}	1.21	1.24	1.23	0.64	0.67	0.65	
L_{60}	1.24	1.26	1.25	0.69	0.72	0.71	
L_{100}	1.28	1.32	1.30	0.74	0.76	0.75	
SEm±	0.02	0.01	0.01	0.01	0.01	0.01	
CD (P=0.05)	0.04	0.04	0.03	0.03	0.03	0.02	
Varieties							
\mathbf{V}_1	1.19	1.21	1.20	0.67	0.69	0.68	
V_2	1.21	1.23	1.22	0.68	0.71	0.69	
V_3	1.16	1.19	1.17	0.65	0.68	0.67	
SEm±	0.01	0.01	0.01	0.01	0.01	0.01	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	

Table 4.7: Nitrogen content in grain and stover of maize as affected by soil pH, lime and varieties

N concentration in plant with liming may be due to their increased solubility in the soil. Enhanced concentration of N as a result of addition of lime in acid soils have been reported by various workers such as Oya and Khondaker (1996); Bhat *et al.* (2007) and Ossom and Rhykerd (2008) in various crops.

From Table 4.7, it is evident that effect of varieties on nitrogen content in grain and stover was non-significant during 2016 as well as 2017.

4.1.1.8 Phosphorus content in grain and stover

It could be inferred from Table 4.8.a that between pH_1 (pH 4.6) and pH_2 (pH 5.2), the highest phosphorus content in both grain and stover was realised in pH₂, *i.e.*, in pH 5.2 during 2016 and 2017. From pooled data, it was observed that phosphorus content in grain increased by 14.7% and in stover by 23.07% in pH₂ over pH₁. Similar findings have been reported by Marquez and Baucas (1990) where higher soil pH (soil pH 5.74) was found to enhance NPK absorption in crops. Low pH soil decreased the phosphorus content in grain and stover of maize. It might be because at low pH, phosphorus is fixed in the soil with aluminium (Adnan *et al.*, 2003) which must have resulted in a decrease in the availability of phosphorus and ultimately reduced phosphorus absorption by plant.

It is evident from Table 4.8.a that the effect of lime on phosphorus content in grain and stover was significant during both the years of experimentation. The maximum phosphorus content in grain was recorded in treatment L_{100} during 2016 as well as 2017, while in stover it was recorded in L_{100} during 2016 and in L_{60} during 2017. From pooled data, P content in both grain and stover was found to increase by 33.3% each in treatment L_{100} in case of grain and in L_{60} in case of stover over L_0 . The minimum phosphorus content in grain and stover was recorded in L_0 . It can be observed that for both grain and stover, treatments L_{60} and L_{100} were at par with each other. The significant increase of P concentrations in plant tissues after lime application can be

Treatment	P co	ntent in grain	n (%)	P content in stover (%)			
	2016	2017	Pooled	2016	2017	Pooled	
Soil pH levels							
pH_1	0.32	0.36	0.34	0.14	0.13	0.13	
pH_2	0.37	0.41	0.39	0.17	0.15	0.16	
SEm±	0.004	0.005	0.003	0.006	0.006	0.004	
CD (P=0.05)	0.01	0.02	0.01	0.02	0.02	0.01	
Lime levels							
L_0	0.29	0.31	0.30	0.13	0.12	0.12	
L ₃₀	0.35	0.39	0.37	0.15	0.14	0.14	
L_{60}	0.37	0.41	0.39	0.16	0.16	0.16	
L_{100}	0.38	0.43	0.40	0.18	0.15	0.16	
SEm±	0.006	0.008	0.005	0.008	0.008	0.006	
CD (P=0.05)	0.02	0.02	0.01	0.02	0.02	0.02	
Varieties							
\mathbf{V}_1	0.35	0.38	0.36	0.14	0.13	0.14	
V_2	0.36	0.41	0.39	0.16	0.15	0.16	
V_3	0.33	0.36	0.35	0.15	0.14	0.15	
SEm±	0.005	0.007	0.004	0.007	0.007	0.005	
CD (P=0.05)	0.02	0.02	0.01	NS	NS	NS	

Table 4.8.a: Phosphorus content in grain and stover of maize as affected by soil pH, lime and varieties

		P content i	n grain (%)					
Soil pH levels	Lime levels							
Son pri levels	L ₀	L ₃₀	L ₆₀	L ₁₀₀				
		20)16					
pH_1	0.26	0.32	0.35	0.37				
pH_2	0.32	0.38	0.39	0.38				
SEm±	0.01							
CD (P=0.05)	0.03							
	2017							
pH_1	0.27	0.35	0.38	0.42				
pH_2	0.34	0.42	0.44	0.44				
SEm±		0.	01					
CD (P=0.05)		0.	03					
		Po	oled					
pH_1	0.26	0.34	0.37	0.40				
pH_2	0.33	0.40	0.42	0.41				
SEm±		0.	01					
CD (P=0.05)		0.	02					

Table 4.8.b: Interaction effect between pH and lime on phosphorus content in grain

as a result of reduced adsorption-precipitation reaction between Al and P at the root surface and in the root free space also known as 'P spring effect' of lime (Fageria *et al.*, 2010). These results are in agreement with those of Jibrin *et al.* (2002); Kovacevic and Rastija (2010) and Muindi *et al.* (2015).

The results revealed that there was no significant difference among the varieties with respect to phosphorus content in stover. However, phosphorus content in grain was observed to show significant difference among the varieties where highest phosphorus content (0.39%) was recorded in the variety RCM-76 (V₂) followed by RCM-75 (V₁) (0.36%) and RCM-1-1 (V₃) (0.35%). However, RCM-75 (V₁) and RCM-1-1 (V₃) were found to be at par with each other.

Interaction effect of pH and lime on phosphorus content in grain

It is evident from Table 4.8.b that maximum phosphorus content in grain was observed in treatment combination pH_2L_{60} and minimum was observed in pH_1L_0 . In both the pH, it was observed that P content increased with increase in lime levels, where at pH_1 , maximum phosphorus content was observed in lime level of L_{100} , while at pH_2 , it was observed in lime level of L_{60} during both the years. Examination of pooled data further revealed that at pH_2 soil, lime levels L_{30} , L_{60} and L_{100} were all at par with each other. This indicates that at pH 5.2, liming up to 30% lime of LR would be ideal for enhancement of phosphorus content in grain. Lime increased the P content in both the soil pH owing to the 'P spring effect' of lime (Fageria *et al.* 2010).

4.1.1.9 Potassium content in grain and stover

It is apparent from Table 4.9 that pH had a significant effect on potassium content where maximum potassium content in both grain and stover was recorded in pH 5.2 (pH₂) and minimum in pH 4.6 (pH₁) during 2016 as well as 2017. From pooled data it can be stated that potassium content in grain and stover increased by 14.9% and 3.5%, respectively in pH₂ over pH₁.

Treatment	K content in grain (%)			K content in stover (%)		
	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	0.46	0.48	0.47	1.14	1.14	1.14
pH_2	0.53	0.55	0.54	1.17	1.18	1.18
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.03	0.03	0.02	0.02	0.02	0.02
Lime levels						
L_0	0.45	0.47	0.46	1.07	1.06	1.06
L ₃₀	0.49	0.51	0.50	1.15	1.16	1.15
L ₆₀	0.51	0.52	0.52	1.20	1.19	1.19
L_{100}	0.55	0.57	0.56	1.21	1.22	1.21
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.04	0.04	0.03	0.03	0.03	0.02
Varieties						
V_1	0.47	0.49	0.48	1.15	1.16	1.15
V_2	0.53	0.55	0.54	1.17	1.17	1.17
V_3	0.50	0.51	0.51	1.15	1.15	1.15
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.03	0.03	0.02	NS	NS	NS

Table 4.9: Potassium content in grain and stover of maize as affected by soil pH, lime and varieties

This is in close conformity with the findings of Otieno *et al.* (2018) who reported positive relationships between pH and N, P, K uptake.

The potassium content in grain and stover was found to increase with increase in lime levels. It is apparent from Table 4.9 that maximum potassium content in both grain (0.56%) and stover (1.21%) was recorded in treatment L_{100} while minimum was recorded in L_0 during both the years of experimentation. Thus, from pooled data it can be stated that L_{100} level of lime enhanced potassium content in grain and stover which increased by 21.7% and 14.2%, respectively over L_0 . Lime application increased the soil pH of acid soils due to which solubility of potassium bearing minerals might have increased which resulted in plants to absorb more potassium from the soil. This is in line with the findings of Ranjit *et al.* (2007) who reported that lime application at 100% lime of LR recorded higher total potassium uptake than other lime levels. Otieno *et al.* (2018) also reported an increase in potassium content in soybean on application of lime.

It can be observed from Table 4.9 that effect of varieties on potassium content was found to be significant only in grain which varied from 0.48 to 0.51%. Maximum potassium content in grain was reported in RCM-76 (V₂) variety which was significantly higher in comparison to other tested varieties which was followed by RCM-1-1 (V₃) and RCM-75 (V₁).

4.1.1.10 Calcium content in grain and stover

The results on calcium content in grain and stover in different treatments have been presented in Table 4.10. It is apparent from the data that maximum calcium content in both grain and stover was recorded in pH 5.2 (pH₂) during 2016 and 2017, whereas the minimum calcium content in grain and stover was recorded in pH 4.6 (pH₁) during both the years. From pooled data, a percent increase of 18.6% in grain and 15.2% in stover can be observed in pH₂ over pH₁. Treatment pH₁ being very strongly acidic in reaction may

Treatment	Ca content in grain (%)			Ca content in stover (%)		
	2016	2017	Pooled	2016	2017	Pooled
Soil pH Levels						
pH_1	0.42	0.43	0.43	0.45	0.48	0.46
pH_2	0.50	0.52	0.51	0.53	0.53	0.53
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.02	0.02	0.02	0.02	0.02	0.01
Lime Levels						
L ₀	0.28	0.28	0.28	0.29	0.32	0.31
L ₃₀	0.50	0.53	0.51	0.53	0.54	0.54
L ₆₀	0.53	0.54	0.53	0.56	0.57	0.56
L_{100}	0.54	0.56	0.55	0.58	0.59	0.58
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.03	0.03	0.02	0.03	0.03	0.02
Varieties						
\mathbf{V}_1	0.46	0.48	0.47	0.49	0.50	0.49
\mathbf{V}_2	0.47	0.48	0.47	0.49	0.51	0.50
V_3	0.46	0.47	0.46	0.49	0.51	0.50
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	NS	NS	NS	NS	NS	NS

Table 4.10: Calcium content in grain and stover of maize as affected by soil pH, lime and varieties

have contained toxic levels of Al which may have affected the uptake of calcium through suppression of root growth. Furthermore, in low pH soils calcium solubility decreases which in turn leads to low calcium absorption by plant. Increase in concentration of H⁺ ions soil in general causes a decrease in the rate of adsorption of cations, which may be due to competition between the similarly charged ions for binding and carrier sites (Alam *et al.*, 1999).

A perusal of the data in Table 4.10 reflects that lime had a significant positive effect on calcium content in grain and stover. The maximum calcium content in both grain and stover was found to be highest in treatment L_{100} during 2016 as well as 2017, whereas the minimum calcium content in grain and stover was recorded in treatment L_0 during 2016 as well as 2017. However, treatment L_{100} was at par with L_{60} in case of calcium content in both grain and stover during both the years. Irrespective of treatments and years, pooled calcium content in grain and stover ranged from 0.28 to 0.55% and 0.31 to 0.58%, respectively. From pooled data, percent increase of 96.4% and 87.1% in grain and stover, respectively in L_{100} over L_0 can be observed. Lime application enhanced the solubility of calcium concentration in soil and resulted in plants to absorb more calcium. These results are in compliance with the findings of Kovacevic and Rastija (2010); Barman *et al.* (2014) and Bhindu *et al.* (2018) who reported a significant increase in Ca concentration in plants with the application of lime.

The calcium content in grain and stover had non-significant difference among the varieties (Table 4.10). Comparatively higher amount of calcium was recorded in stover than grain in case of all varieties.

4.1.1.11 Boron content in grain and stover

From Table 4.11, it is evident that boron content in grain with respect to pH was found to be non-significant. However, it was found to be significant

Treatment	B content in grain (mg kg ⁻¹)			B content in stover (mg kg ⁻¹)		
	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	9.49	9.51	9.50	3.23	3.27	3.25
pH_2	9.51	9.54	9.53	3.27	3.31	3.29
SEm±	0.01	0.02	0.01	0.01	0.01	0.01
CD (P=0.05)	NS	NS	NS	0.02	0.02	0.01
Lime levels						
L_0	9.38	9.40	9.39	3.14	3.17	3.15
L ₃₀	9.60	9.62	9.61	3.33	3.36	3.34
L ₆₀	9.57	9.61	9.59	3.31	3.35	3.33
L_{100}	9.45	9.48	9.46	3.23	3.27	3.25
SEm±	0.02	0.02	0.01	0.01	0.01	0.01
CD (P=0.05)	0.05	0.06	0.04	0.03	0.03	0.02
Varieties						
V_1	9.51	9.54	9.52	3.25	3.28	3.26
V_2	9.50	9.53	9.52	3.26	3.29	3.28
V_3	9.49	9.51	9.50	3.25	3.29	3.27
SEm±	0.01	0.02	0.01	0.01	0.01	0.01
CD (P=0.05)	NS	NS	NS	NS	NS	NS

Table 4.11: Boron content in grain and stover of maize as affected by soil pH, lime and varieties

for boron content in stover, where highest value was recorded in pH 5.2 (pH₂) during 2016 as well as 2017. Thus, lowest B content in stover was recorded in pH 4.6 (pH₁) in both the years. Strongly acid soils (pH less than 5.0) are likely to be low in available boron because of boron sorption to surfaces of iron and aluminium oxides which thus reduces its concentration in plants.

It is apparent from Table 4.11 that maximum boron content in grain and stover was recorded in L_{30} during 2016 as well as 2017. Pooled data reflected an increase in grain and boron content in grain and stover by 2.3% and 6.0%, respectively on application of 30% lime of LR (L_{30}). The minimum boron content in both grain and stover was recorded in L_0 during both the years. It was also observed that application of lime at 30% and 60% lime of LR were statistically at par in influencing the B concentration in plant. At 100% lime of LR significant reduction in plant B concentration could be observed in both grain and stover which may be because B absorption by plants decreases when both pH and Ca concentrations are increased in the soil (Alam *et al.*, 1999). The reduction in B concentration at higher lime level has also been reported by Barman *et al.* (2014).

The results on effect of varieties on boron content in grain and stover was non-significant (Table 4.11).

4.1.1.12 Nitrogen uptake in grain and stover

As can be seen from Table 4.12 and Fig. 3, nitrogen uptake in both grain and stover showed significant difference between the two treatments. It is apparent from the table, maximum nitrogen uptake in both grain and stover was recorded in pH 5.2 (pH₂) and minimum was recorded in pH 4.6 (pH₁) in both the years. From pooled data it can be observed that pH₂ enhanced nitrogen uptake in grain by 14.8% and in stover by 12.5% over pH₁. Results on decrease in N uptake in plants with decrease in soil pH has been reported by Fageria and Zimmermann (1998); Otieno *et al.* (2018) and Pan *et al.* (2020).
Tracturent	N upta	ake in grain (n	ng pot ⁻¹)	N uptak	e in stover (n	ng pot ⁻¹)
Treatment	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	759.34	775.39	767.37	786.83	836.92	811.88
pH_2	865.76	896.05	880.91	884.66	942.69	913.68
SEm±	8.91	7.63	5.87	12.05	14.51	9.43
CD (P=0.05)	25.33	21.70	16.47	34.27	41.26	26.48
Lime levels						
L ₀	575.83	581.49	578.66	641.40	693.09	667.24
L ₃₀	848.67	863.14	855.91	812.47	869.39	840.93
L ₆₀	900.79	921.25	911.02	909.09	965.93	937.51
L ₁₀₀	924.91	977.01	950.96	980.04	1030.82	1005.43
SEm±	12.60	10.79	8.30	17.04	20.52	13.34
CD (P=0.05)	35.82	30.69	23.29	48.46	58.35	37.44
Varieties						
\mathbf{V}_1	821.72	845.94	833.83	843.38	896.92	870.15
V_2	860.26	878.19	869.22	869.09	929.15	899.12
V_3	755.67	783.04	769.36	794.78	843.35	819.07
SEm±	10.91	9.35	7.18	14.76	17.77	11.55
CD (P=0.05)	31.02	26.58	20.17	41.97	50.53	32.43

Table 4.12: Nitrogen uptake in grain and stover of maize as affected by soil pH, lime and varieties



Fig 3: Effect of soil pH, lime and varieties on nitrogen uptake in grain and stover of maize

From Table 4.12 and Fig. 3, it can be observed that lime had a significant positive effect on nitrogen uptake in both grain and stover. From pooled data, it can be observed that during 2016 as well as 2017, maximum nitrogen uptake in both grain and stover was recorded at 100% lime of LR (L_{100}), and the minimum in control (L_0). From pooled data, it can be observed that liming at 100% lime of LR enhanced nitrogen uptake to the extent of 64.3% and 50.6% in grain and stover, respectively over control. Lime application increased the grain and stover yield as well as nitrogen content which ultimately enhanced the nitrogen uptake by crop. Increase in nitrogen uptake in plants due to liming has been reported by Chaterjee *et al.* (2005); Ranjit *et al.* (2007) and Lynrah and Nongmaithem (2017).

The results on effect of varieties on nitrogen uptake revealed that there was a significant difference among the treatments (Table 4.12 and Fig. 3). In both the years, maximum nitrogen uptake in both grain and stover was observed in variety RCM-76 (V₂), while minimum was recorded in RCM-1-1 (V₃). A critical examination of data shows that significantly higher nitrogen uptake was recorded in V₂ in comparison to V₁ and V₃ varieties. The nitrogen uptake in grain of V₂ variety was 4.2% and 13.0% higher than V₁ and V₃, respectively. Variation in nitrogen uptake might be due to variation in grain and stover yield of tested varieties.

4.1.1.13 Phosphorus uptake in grain and stover

The results from Table 4.13.a and Fig. 4 revealed that there was a significant effect of pH levels on phosphorus uptake. During 2016 and 2017, the maximum phosphorus uptake in both grain and stover was recorded in pH 5.2 (pH₂). Thus, minimum P uptake in both grain and stover was recorded in pH 4.6 (pH₁). From pooled data, it can be observed that phosphorus uptake increased by 29.1% in grain and by 31.8% in stover in pH₂ compared to pH₁.

Turadanaant	P uptak	te in grain (m	ng pot ⁻¹)	P uptak	e in stover (n	ng pot ⁻¹)
Treatment	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	210.22	232.15	221.18	162.63	160.48	161.55
pH_2	267.61	303.50	285.55	221.45	204.67	213.06
SEm±	3.50	3.82	2.59	7.35	7.32	5.19
CD (P=0.05)	9.96	10.87	7.28	20.90	20.82	14.56
Lime levels						
L_0	166.02	177.06	171.54	134.82	135.21	135.02
L ₃₀	245.69	269.58	257.63	187.77	175.85	181.81
L ₆₀	271.37	305.04	288.21	204.49	216.20	210.35
L_{100}	272.58	319.62	296.10	241.06	203.04	222.05
SEm±	4.95	5.41	3.67	10.40	10.35	7.34
CD (P=0.05)	14.09	15.38	10.29	29.56	29.44	20.59
Varieties						
V_1	240.87	266.55	253.71	180.89	169.91	175.40
V_2	257.73	296.02	276.87	208.80	199.16	203.98
V_3	218.14	240.90	229.52	186.41	178.66	182.53
SEm±	4.29	4.68	3.18	9.00	8.97	6.35
CD (P=0.05)	12.20	13.32	8.91	NS	NS	NS

Table 4.13.a: Phosphorus uptake in grain and stover of maize as affected by soil pH, lime and varieties



Fig 4: Effect of soil pH, lime and varieties on phosphorus uptake in grain and stover of maize

		P uptake in g	rain (mg pot ⁻¹)						
Soil pH levels		Lime levels							
	L_0	L ₃₀	L ₆₀	L ₁₀₀					
	2017								
pH_1	144.50	230.61	258.29	295.18					
pH ₂	209.61	308.55	351.79	344.06					
SEm±		7.	65						
CD (P=0.05)		21	.75						

Table 4.13.b: Interaction effect between pH and lime on phosphorus uptake in grain

Similar findings have also been reported by Fageria and Zimmermann (1998) where higher pH increased the uptake of phosphorus in corn, wheat and common bean. At high pH, grain and stover yield and phosphorus content increased and ultimately phosphorus uptake was increased because uptake is product of yield and nutrient content.

The data (Table 4.13.a and Fig. 4) showed that during 2016 as well as 2017 maximum phosphorus uptake in grain was recorded at 100% lime of LR (L_{100}) which was however found to be at par with treatment L_{60} . On the other hand, maximum phosphorus uptake in stover during 2016 was recorded in treatment L_{100} , while during 2017 it was recorded in treatment L_{60} which was found to be at par with treatment L_{100} . Minimum phosphorus uptake in both grain and stover was recorded in control (L₀). From pooled data it can be observed that there was 72.6% and 64.5% increase in grain and stover, respectively in L₁₀₀ as compared to L₀. Liming may have increased plant P uptake by relatively decreasing Al than by increasing the availability of phosphorus in soil. Thus, this may have improved the root growth of the plant allowing for a greater volume of soil to be explored (Friesen et al., 1980). Lime application enhanced yield and phosphorus content in grain and stover which resulted in increased phosphorus uptake. These results are in agreement with those of Kovacevic and Rastija (2010); Muindi et al. (2015) and Zhihao et al. (2019).

In case of effect of varieties on phosphorus uptake, it can be observed from Table 4.13.a and Fig. 4 that in both the years, maximum phosphorus uptake in grain was recorded in variety RCM-76 (V₂) followed by RCM-75 (V₁) and minimum was observed in RCM-1-1 (V₃). However, variety did not have any significant effect on phosphorus uptake in stover in both the years. It was also revealed that phosphorus uptake in grain of RCM-76 (V₂) variety was 9.1% and 20.6% higher as compared to V₁ and V₂ varieties, respectively.

Interaction effect of pH and lime on phosphorus uptake in grain

It is evident from Table 4.13.b that during 2017, maximum phosphorus uptake in grain at pH 4.6 (pH₁) was observed at 100% lime of LR (pH₁L₁₀₀), whereas at pH 5.2 (pH₂), maximum phosphorus uptake in grain was observed at 60% lime of LR (pH₂L₆₀). It could also be observed that pH₂L₆₀ was at par with pH₂L₁₀₀. The minimum phosphorus uptake in grain in both the soil pH was observed at L₀. Thus, it could be observed that irrespective of pH levels, lime enhanced phosphorus uptake in grain. Thus, soil pH and lime was observed to exhibit positive interaction which reduced aluminium toxicity in the soils and improved P uptake in acid soils. However pH₂ soil gave greater response in terms of P uptake in grain as compared to pH₁ soil.

4.1.1.14 Potassium uptake in grain and stover

The results on the potassium uptake in different treatments have been presented in Table 4.14 and Fig. 5. Effect of pH on potassium uptake in both grain and stover was found to be highly significant, where during 2016 and 2017, maximum potassium uptake in both grain and stover was recorded in pH 5.2 (pH₂) whereas minimum potassium uptake in both grain and stover was recorded in pH 4.6 (pH₁). An examination of pooled data reflects a 29.9% and 13.3% increase in potassium uptake by grain and stover, respectively in pH₂ over pH₁. Low uptake of K in pH 4.6 could have been due to the detrimental effect of low soil pH. Such low pH hinders normal root growth, development, and absorption of water and nutrients (Lin *et al.*, 2012). Positive relationship between soil pH and NPK uptake has also been reported by Fageria and Zimmermann (1998) and Otieno *et al.* (2018).

The data (Table 4.14 and fig. 5) showed that in 2016 as well as 2017, in both grain and stover, maximum uptake was recorded at 100% lime of LR (L_{100}) and minimum in control (L_0). From pooled data, it can be observed that there was an increase in potassium uptake in grain by 54.7% and in stover by

Turantan	K uptal	ke in grain (m	ng pot ⁻¹)	K uptak	e in stover (n	ng pot ⁻¹)
Treatments	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	299.88	308.86	304.37	1357.53	1389.03	1373.28
pH_2	384.95	406.13	395.54	1536.90	1576.86	1556.88
SEm±	7.23	7.35	5.16	16.73	16.57	11.77
CD (P=0.05)	20.56	20.91	14.48	47.58	47.11	33.05
Lime levels						
L_0	258.11	270.66	264.39	1145.05	1172.92	1158.99
L ₃₀	342.57	352.45	347.51	1471.07	1506.27	1488.67
L_{60}	371.70	385.80	378.75	1566.81	1589.62	1578.21
L_{100}	397.26	421.06	409.16	1605.91	1662.95	1634.43
SEm±	10.22	10.40	7.29	23.66	23.43	16.65
CD (P=0.05)	29.07	29.58	20.47	67.28	66.63	46.74
Varieties						
\mathbf{V}_1	324.82	342.54	333.68	1451.47	1490.14	1470.81
\mathbf{V}_2	378.38	391.19	384.79	1493.02	1539.50	1516.26
V_3	324.04	338.75	331.39	1397.14	1419.19	1408.17
SEm±	8.85	9.01	6.32	20.49	20.29	14.42
CD (P=0.05)	25.18	25.61	17.73	58.27	57.70	40.48

Table 4.14: Potassium uptake in grain and stover of maize as affected by soil pH, lime and varieties



Fig 5: Effect of soil pH, lime and varieties on potassium uptake in grain and stover of maize

41.0% in L_{100} over control. Lime application enhanced the grain and stover yield and potassium content due to which potassium uptake in grain and stover also increased. Increase in potassium uptake due to liming on various crop plants have been reported by Meena and Varma (2016); Lynrah and Nongmaithem (2017) and Han *et al.* (2019).

The data (Table 4.14 and fig. 5) showed that varieties had a significant effect on both potassium uptake in grain and stover. Maximum uptake in both grain and stover in both the years was recorded in the variety RCM-76 (V₂) followed by RCM-75 (V₁) while minimum was observed in RCM-1-1 (V₃). A critical examination of data indicate that pooled potassium uptake in grain of V₂ was 15.3% and 16.1% higher over V₁ and V₃, respectively.

4.1.1.15 Calcium uptake in grain and stover

Data pertaining to calcium uptake is summarized in Table 4.15 and depicted in Fig. 6. The results on effect of pH revealed significant influence on calcium uptake in grain and stover. During 2016 and 2017, maximum calcium uptake was recorded in pH 5.2 (pH₂) in grain as well as stover while minimum calcium uptake was recorded in pH 4.6 (pH₁). Irrespective of treatments and years the calcium uptake in grain and stover varied from 161.73 to 404.16 mg pot⁻¹ in grain and from 338.23 to 784.55 mg pot⁻¹ in stover. From pooled data it can be observed that calcium uptake in grain and stover ca uptake in pH 4.6 (pH₁) may be due to the antagonistic effect between H⁺ and Ca²⁺ ions at the substitution site of the cell wall and/or of the plasma membrane, thus inhibiting Ca²⁺ uptake in plants (Alam *et al.*, 1999).

The data (Table 4.15) showed that the maximum calcium uptake in both grain and stover was recorded at 100% lime of LR (L_{100}) in both the years. However, in case of Ca uptake in grain, treatment L_{100} was at par with L_{60} . The

Turaturant	Ca upta	ke in grain (r	ng pot ⁻¹)	Ca uptak	ke in stover (mg pot ⁻¹)
Ireatment	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	278.53	283.24	280.89	547.98	589.57	568.77
pH_2	367.21	389.56	378.39	702.30	722.92	712.61
SEm±	5.62	5.94	4.09	10.61	10.46	7.45
CD (P=0.05)	15.99	16.88	11.48	30.18	29.74	20.91
Lime levels						
L_0	160.86	162.60	161.73	318.31	358.15	338.23
L ₃₀	352.48	367.63	360.05	681.40	707.89	694.65
L ₆₀	386.79	398.43	392.61	729.72	760.97	745.34
L_{100}	391.35	416.97	404.16	771.13	797.96	784.55
SEm±	7.95	8.40	5.78	15.01	14.79	10.54
CD (P=0.05)	22.61	23.88	16.23	42.68	42.06	29.58
Varieties						
V_1	325.41	340.90	333.16	625.60	655.64	640.62
V_2	339.00	352.15	345.57	638.67	676.54	657.60
V_3	304.20	316.16	310.18	611.16	636.54	623.85
SEm±	6.89	7.27	5.01	13.00	12.81	9.12
CD (P=0.05)	19.58	20.68	14.06	NS	NS	NS

Table 4.15: Calcium uptake in grain and stover of maize as affected by soil pH, lime and varieties



Fig 6: Effect of soil pH, lime and varieties on calcium uptake in grain and stover of maize

minimum calcium uptake in grain as well as stover was observed to be associated with control treatment (L_0) in both the years. Thus, an increase in 149.9% in grain and 131.9% in stover, respectively at 100% lime of LR over control can be observed from the pooled data. Each increasing level of lime resulted in significant increase in calcium uptake by grain and stover in comparison to preceding lower level of lime. Increase in Ca uptake due to liming is recorded in the findings of Singh *et al.* (2009); Moreira and Fageria (2010) and Bhindu *et al.* (2018).

The data (Table 4.15) showed that on the basis of pooled data effect of varieties on calcium uptake in grain was found to be significant where maximum Ca uptake was observed in variety RCM-76 (V₂) followed by RCM-75 (V₁) and the minimum in RCM-1-1 (V₃). Effect of varieties on Ca uptake in stover was observed to be non-significant during both the years.

4.1.1.16 Boron uptake in grain and stover

The data presented in Table 4.16 and depicted in Fig. 7 revealed that the effect of pH on boron uptake in grain and stover was significant. The maximum boron uptake in both the years in both grain and stover was observed in pH 5.2 (pH₂) and the minimum was observed in pH 4.6 (pH_{4.6}). Thus, from pooled data, boron uptake in grain increased by 13.7% and in stover by 10.8% in pH 5.2 over pH 4.6. This may be due to the fact that at a lower soil pH, boron is held more tightly by sesquioxides compared to relatively higher soil pH (Goldberg, 1997) and thus may have led to less availability for plant uptake.

The data (Table 4.16 and Fig. 7) showed that in both the years, maximum B uptake in grain as well as stover was recorded at 60% lime of LR (L_{60}) while minimum B uptake was observed to be associated with control (L_0). A critical examination of the data shows that effect of L_{60} was at par with L_{100} in boron uptake in grain, while L_{60} was at par with L_{100} and L_{30} in boron uptake in stover. The L_{60} level of lime increased pooled boron uptake in grain and

T ()	B uptal	ke in grain (µ	g pot ⁻¹)	B uptake in stover (µg pot ⁻¹) 2016 2017 Pooled			
Treatment	2016	2017	Pooled	2016	2017	Pooled	
Soil pH levels							
pH_1	608.46	606.53	607.50	385.07	398.41	391.74	
pH_2	683.17	698.49	690.83	426.78	441.90	434.34	
SEm±	3.82	3.79	2.69	3.69	4.06	2.74	
CD (P=0.05)	10.85	10.77	7.55	10.49	11.54	7.70	
Lime levels							
L ₀	530.30	536.28	533.29	336.57	350.16	343.36	
L ₃₀	673.37	669.64	671.51	424.30	437.61	430.96	
L ₆₀	695.84	704.29	700.06	433.01	446.64	439.83	
L_{100}	683.75	699.82	691.79	429.81	446.20	438.00	
SEm±	5.40	5.36	3.80	5.21	5.74	3.88	
CD (P=0.05)	15.34	15.23	10.67	14.83	16.32	10.89	
Varieties							
V_1	651.39	659.56	655.48	408.09	422.02	415.06	
V_2	672.44	676.39	674.42	415.88	431.97	423.92	
V ₃	613.61	621.58	617.59	393.80	406.46	400.13	
SEm±	4.67	4.64	3.29	4.52	4.97	3.36	
CD (P=0.05)	13.29	13.19	9.24	12.84	14.14	9.43	

Table 4.16: Boron uptake in grain and stover of maize as affected by soil pH, lime and varieties



Fig 7: Effect of soil pH, lime and varieties on boron uptake in grain and stover of maize

stover to the extent of 31.2% and 28.1% over control, respectively. Similar results where liming increased boron uptake in plants is found in conformity with the findings of Su *et al.* (1994) and Singh *et al.* (2009).

The data presented in table 4.16 and illustrated in fig. 7 showed that the maximum B uptake in grain and stover was recorded in RCM-76 (V₂) and minimum in RCM-1-1 (V₃). In case of B uptake in stover, RCM-75 (V₁) and RCM-76 (V₂) was found to be at par with each other. It was also revealed that pooled boron uptake in grains of V₂ variety was 2.8 and 9.2% higher than V₁ and V₃ varieties, respectively.

4.1.2 Effect of soil pH, lime levels and varieties on soil properties4.1.2.1 Effect on soil pH and electrical conductivity

It is apparent from the data (Table 4.17.a) that during 2016 and 2017 maximum pH in soil after harvest was recorded in pH₂ with pooled data of 5.92 while minimum pH was recorded in pH₁ with pooled data of 5.61. Effect of pH on electrical conductivity was found to be non-significant in both the years. However, it could be observed that electrical conductivity was higher in pH 4.6 (pH₁) compared to pH 5.2 (pH₂). Lower soil pH indicates larger number of hydrogen ions in the soil and higher amount of hydrogen ions in the soil environment can appear in varying amounts which can affect the level of electrical conductivity.

It is apparent from table 4.17.a that lime did not have significant effect on electrical conductivity. However, in case of soil pH, incorporation of increasing doses of lime increased pH across the treatments. From the data, it can be observed that during 2016 as well as 2017, maximum pH was recorded in L_{100} level of lime with a pooled value of 6.42 while minimum pH was observed in L₀ with a pooled value of 4.91. Thus, highest soil pH was recorded LR at 100% lime of which is in congruence with

Tuestaent		Soil pH		Electrical conductivity (dSm ⁻¹) 2016 2017 Pooled 0.19 0.22 0.21 0.18 0.21 0.19 0.008 0.006 0.006 NS NS NS 0.21 0.23 0.22 0.21 0.23 0.22 0.19 0.20 0.19 0.17 0.22 0.19 0.18 0.21 0.20 0.19 0.20 0.19 0.11 0.008 0.20 0.18 0.21 0.20 0.011 0.008 NS NS NS NS 0.20 0.22 0.21		
Ireatment	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	5.61	5.61	5.61	0.19	0.22	0.21
pH_2	5.92	5.91	5.92	0.18	0.21	0.19
SEm±	0.006	0.006	0.004	0.008	0.008	0.006
CD (P=0.05)	0.02	0.02	0.01	NS	NS	NS
Lime levels						
L_0	4.90	4.91	4.91	0.21	0.23	0.22
L ₃₀	5.68	5.73	5.71	0.19	0.20	0.19
L ₆₀	6.06	6.00	6.03	0.17	0.22	0.19
L_{100}	6.43	6.40	6.42	0.18	0.21	0.20
SEm±	0.009	0.009	0.006	0.011	0.011	0.008
CD (P=0.05)	0.02	0.03	0.02	NS	NS	NS
Varieties						
V_1	5.77	5.76	5.77	0.20	0.22	0.21
V_2	5.76	5.77	5.76	0.18	0.22	0.20
V_3	5.77	5.76	5.76	0.19	0.21	0.20
SEm±	0.007	0.008	0.005	0.010	0.010	0.007
CD (P=0.05)	NS	NS	NS	NS	NS	NS

Table 4.17.a: Soil pH and electrical conductivity of post-harvest soil as affected by soil pH, lime and varieties

		Soil	pH				
Soil pH levels		Lime	levels				
	L ₀	L ₃₀	L ₆₀	L ₁₀₀			
		20	16				
pH_1	4.60	5.47	5.96	6.42			
pH ₂	5.20	5.88	6.16	6.44			
SEm±	0.01						
CD (P=0.05)	0.03						
	2017						
pH_1	4.60	5.52	5.94	6.39			
pH ₂	5.21	5.94	6.07	6.42			
SEm±		0.0	01				
CD (P=0.05)		0.0	04				
		Poo	oled				
pH_1	4.60	5.50	5.95	6.41			
pH ₂	5.21	5.91	6.11	6.43			
SEm±		0.0	01				
CD (P=0.05)		0.0	02				

Table 4.17.b: Interaction effect between pH and lime on soil pH of post-harvest soil

the findings of Dasog *et al.* (2010). Liming at 100% lime of LR (L₁₀₀) increased soil pH by 30.7% over the control treatment (L₀) as reflected in pooled data. Liming increased soil pH because of the probable displacement of Al³⁺, H⁺ and Fe³⁺ ions by Ca²⁺ ions present in lime (Kisinyo *et al.*, 2013). The lowest soil pH value obtained in the control (L₀) can be attributed to the lowest values of exchangeable basic cations at the exchange sites of soil due to exhaustive uptake of the exchangeable bases by maize during the growing period which is in conformity with the findings of Lege (2012).

The results revealed that there was no significant difference among the varieties with respect to soil pH and electrical conductivity.

Interaction effect of pH and lime on soil pH

From the data (Table 4.17.b) the maximum pH in soil at harvest during 2016 and 2017 was recorded in pH₂L₁₀₀ with a pooled value of 6.24 while the minimum pH was recorded in pH₁L₀ with a pooled value of 4.44. Soil pH showed a linear increase with increase in lime levels irrespective of pH levels during both the years 2016 and 2017. Lime increases soil pH because of the Ca²⁺ ions present in lime which displaces Al³⁺, H⁺ and Fe³⁺ ions in the soil exchange complex (Kisinyo *et al.*, 2013).

4.1.2.2 Effect on percent base saturation and organic carbon

From Table 4.18.a it is evident that effect of pH was non-significant on base saturation and organic carbon content in soil at harvest. But, a slightly higher value of base saturation and organic carbon was recorded in pH₂ soils.

It can be observed from Table 4.18.a that there was no significant effect of lime on soil organic carbon during both the years of experimentation. The base saturation of soil was affected significantly with lime application during both the years of experimentation. The pooled base saturation increased from 24.33% in control to 33.47% in 100% lime of LR. Each increasing level of

Turnet	Ba	se saturation	(%)	Org	ganic carbon	nic carbon (%) 2017 Pooled 1.55 1.55 1.57 1.56 0.01 0.01 NS NS		
I reatment	2016	2017	Pooled	2016	2017	Pooled		
Soil pH levels								
pH ₁	29.57	29.16	29.36	1.55	1.55	1.55		
pH ₂	30.25	29.60	29.92	1.56	1.57	1.56		
SEm±	0.40	0.34	0.26	0.01	0.01	0.01		
CD (P=0.05)	NS	NS	NS	NS	NS	NS		
Lime levels								
L_0	24.17	24.48	24.33	1.55	1.56	1.55		
L ₃₀	28.82	27.97	28.39	1.55	1.56	1.56		
L_{60}	32.88	31.90	32.39	1.56	1.55	1.56		
L_{100}	33.76	33.17	33.47	1.57	1.57	1.57		
SEm±	0.56	0.48	0.37	0.01	0.01	0.01		
CD (P=0.05)	1.61	1.38	1.04	NS	NS	NS		
Varieties								
\mathbf{V}_1	30.05	29.33	29.69	1.55	1.56	1.56		
V_2	29.72	29.34	29.53	1.56	1.57	1.56		
V_3	29.95	29.47	29.71	1.56	1.55	1.56		
SEm±	0.49	0.42	0.32	0.01	0.01	0.01		
CD (P=0.05)	NS	NS	NS	NS	NS	NS		

Table 4.18.a: Base saturation and organic carbon of post-harvest soil as affected by soil pH, lime and varieties

Table 4.18.b: Interaction effect between pH and lime on percent base saturation of post-harvest soil

	Base saturation (%)						
Soil pH levels		Lime	levels				
	L ₀	L ₃₀	L ₆₀	L ₁₀₀			
	2017						
pH_1	24.69	28.88	30.19	32.87			
pH_2	24.27	27.05	33.62	33.48			
SEm±	0.69						
CD (P=0.05)		1.	95				
		Poo	bled				
pH_1	24.35	28.82	30.71	33.57			
pH_2	24.30	27.96	34.07	33.36			
SEm±	0.53						
CD (P=0.05)		1.	48				

lime enhanced pooled base saturation significantly over preceding lower level of lime. Application of L_{100} level of lime increased base saturation by 37.5% as compared to control. The increase in percent base saturation due to liming can be attributed to the replacement of acidic cations by basic cations. These results are in agreement with those of Sethi (2015).

As apparent from Table 4.18.a there was no significant effect of varieties on percent base saturation and organic carbon in both the years.

Interaction effect of pH and lime on percent base saturation

The effect of pH and lime on percent base saturation in soil at harvest was found to be significant during the second year of experimentation. From Table 4.18.b it can be seen that during 2017, the maximum percent base saturation in soil at harvest was recorded in pH_2L_{60} with pooled data as 34.07%. It can also be observed that increase in lime levels led to an increase in the percent base saturation under both the soil pH. This is because liming increases the soil pH, and acidic cations such as Al and H are replaced by basic cations such as Ca, Mg and K which then increases the percent base saturation.

4.1.2.3 Effect on available nitrogen, phosphorus and potassium

From Table 4.19 it can be observed that pH did not have any effect on available nitrogen and potassium whereas effect of pH on phosphorus was significant during both the years. The maximum available phosphorus in soil at harvest during 2016 as well as 2017 was recorded in pH₂ with a pooled value of 12.13 kg ha⁻¹, whereas the minimum available phosphorus was recorded in pH₁ with pooled value of 8.97 kg ha⁻¹. The pH₂ level enhanced available phosphorus by 10.6% over pH₁. Thus, available P significantly increased with increasing pH, suggesting that the relative abundance of available P decreased with increasing acidity. Soil acidification can result in decreased available P value (Awani, 2012). Concentration of P in soil depends mainly on soil pH and

Trastmont	Availa	able nitrogen (k	kg ha ⁻¹)	Availab	le phosphorus	(kg ha ⁻¹)	Available potassium (kg ha ⁻¹)			
Treatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled	
Soil pH levels										
pH_1	262.14	263.66	262.90	10.65	9.82	10.23	150.95	151.49	151.22	
pH_2	265.01	265.35	265.18	11.69	10.94	11.32	151.98	152.06	152.02	
SEm±	1.54	1.56	1.10	0.18	0.27	0.16	0.54	0.54	0.38	
CD (P=0.05)	NS	NS	NS	0.51	0.76	0.45	NS	NS	NS	
Lime levels										
L ₀	240.18	241.80	240.99	9.13	8.82	8.97	149.52	148.89	149.21	
L ₃₀	259.02	258.10	258.56	10.74	10.35	10.54	151.83	151.37	151.60	
L ₆₀	277.77	279.99	278.88	12.27	11.03	11.65	151.68	152.83	152.25	
L ₁₀₀	277.33	278.13	277.73	12.53	11.33	11.93	152.82	154.01	153.41	
SEm±	2.18	2.21	1.55	0.25	0.38	0.23	0.76	0.76	0.54	
CD (P=0.05)	6.20	6.29	4.36	0.72	1.08	0.64	2.17	2.16	1.51	
Varieties										
\mathbf{V}_1	262.96	263.30	263.13	11.18	10.21	10.70	151.84	151.88	151.86	
V_2	263.16	264.75	263.95	11.09	10.42	10.76	151.30	151.66	151.48	
V ₃	264.59	265.47	265.03	11.24	10.51	10.87	151.25	151.78	151.51	
SEm±	1.89	1.92	1.34	0.22	0.33	0.20	0.66	0.66	0.47	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS	

Table 4.19: Available nitrogen, phosphorus and potassium of post-harvest soil as affected by soil pH, lime and varieties

therefore, a decrease in the soil pH level reduces the concentration of P in soil due to the precipitation of P as Al or Fe phosphate which are amorphous polynuclear complexes with high surface area causing fixation to occur (Alam *et al.*, 1999).

It is apparent from table 4.19 that maximum and minimum nitrogen content in soil at harvest was recorded in L₆₀ and L₀, respectively during 2016 and 2017. However, L_{60} was found to be at par with L_{100} . Phosphorus and potassium content in soil at harvest was recorded maximum in L₁₀₀ during 2016 and 2017 which was at par with L_{30} and L_{60} in case of phosphorus content, while L_{100} was at par with L_{60} in case of potassium content in soil. The minimum phosphorus and potassium content was recorded in treatment L₀ during 2016 and 2017. A critical examination of data showed that L_{60} level of lime enhanced available nitrogen by 15.7% over control while L₁₀₀ level of lime enhanced availability of phosphorus and potassium by 33% and 2.8% over control. Higher availability of N with liming may be due to increase in soil pH where the rate of organic matter mineralization and microbial activity increases. Higher availability of P in soil under liming might be attributed to increased solubility of native and applied P due to increase in soil pH as reported by Rahman et al. (2005). Liming increased K availability, likely through the displacement of exchangeable K by Ca. The increase in potassium availability in soil due to liming coincides with the findings of Basak (2010) and Han et al. (2019).

It is apparent from Table 4.19 that effect of varieties on available nitrogen, phosphorus and potassium was non-significant during both the experimentation years.

4.1.2.4 Effect on exchangeable calcium and available boron

Effect of pH was non-significant on exchangeable calcium whereas it was significant with respect to available boron as can be observed in Table 4.20. During 2016 as well as 2017 maximum available boron content in soil at

Tractmont	Ez	xchangeable [cmol (p ⁺) kg	Ca ²⁺ g ⁻¹]	Ava	ilable B (mg	kg ⁻¹)
Treatment	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels						
pH_1	4.92	4.91	4.92	0.47	0.45	0.46
pH_2	4.94	4.95	4.95	0.55	0.53	0.54
SEm±	0.06	0.06	0.04	0.005	0.005	0.004
CD (P=0.05)	NS	NS	NS	0.01	0.02	0.01
Lime levels						
L ₀	2.98	2.95	2.96	0.49	0.47	0.48
L ₃₀	4.24	4.23	4.24	0.53	0.51	0.52
L ₆₀	6.01	6.02	6.02	0.51	0.49	0.50
L_{100}	6.50	6.51	6.51	0.50	0.49	0.49
SEm±	0.08	0.08	0.06	0.007	0.008	0.005
CD (P=0.05)	0.24	0.24	0.17	0.02	0.02	0.01
Varieties						
\mathbf{V}_1	4.93	4.92	4.92	0.51	0.50	0.51
V_2	4.93	4.93	4.93	0.50	0.48	0.49
V ₃	4.94	4.94	4.94	0.51	0.49	0.50
SEm±	0.07	0.07	0.05	0.006	0.007	0.004
CD (P=0.05)	NS	NS	NS	NS	NS	NS

Table 4.20: Exchangeable calcium and available boron of post-harvest soil as affected by soil pH, lime and varieties

harvest was recorded in pH 5.2 (pH₂), whereas, the minimum was recorded in pH 4.6 (pH₁). Thus, higher pH level recorded higher available boron. Availability of boron was improved by 17.4% in pH₂ soil over pH₁. The positive relationship of B desorption with soil pH may be ascribed to the fact that both free Fe and Al oxide contents in soil decreased with an increase in soil pH. It can also be inferred that the B adsorbed on sesquioxides at lower pH was held more tightly as compared to that adsorbed on soil constituents other than sesquioxides at relatively higher pH (Goldberg, 1997).

From Table 4.20 it can be observed that increasing doses of lime increased exchangeable calcium significantly during both the years of experimentation. During 2016 as well as 2017 the highest exchangeable calcium was observed in treatment L_{100} and lowest in control treatment (L_0). The data further revealed that each increasing level of lime resulted in a significant enhancement in pooled exchangeable calcium in comparison to preceding lower levels of lime. The L_{100} level of lime enhanced exchangeable calcium to the extent of 119.9% over control. On the other hand, highest available boron was observed in treatment L_{30} and lowest in control (L_0). Available boron @ 30% lime of LR (L₃₀) was observed to increase by 8.3% over control (L_0) . Increasing doses of lime was observed to reduce the soil available boron. Liming generally increases the retention capacity of boron in soil due to formation of insoluble metaborate (Mikko, 1972) which may have been the reason for its reduction in higher lime levels. The result on soil available boron is in line with the findings of Barman et al. (2014). The status of available Ca on soils was positively correlated with the rate of lime application. This result is in close conformity with the findings of Garica (1975) who reported that the pH of acid soils becomes raised due to liming, and higher rate of lime application leads to higher adsorption of Ca and thus ameliorates calcium deficiency.

There was no significant effect of varieties on soil exchangeable calcium and available boron content at harvest.

4.1.2.5 Effect on exchangeable hydrogen, aluminium and total potential acidity

From Table 4.21.a it can be observed that the effect of pH on exchangeable hydrogen, exchangeable aluminium and total potential acidity was found to be significant where highest values during 2016 and 2017 was observed in pH 4.6 (pH₁) soil and lowest values were observed in pH 5.2 (pH₂) soil. Thus, from pooled data it can be stated that exchangeable hydrogen, exchangeable aluminium and total potential acidity decreased by 32.6%, 9.2% and 6.3%, respectively in pH₂ as compared to pH₁. At low pH (<5) aluminium becomes soluble and is adsorbed by silicate clays or is tightly bound by organic matter. The Al³⁺ ions are then hydrolysed in the soil solution releasing H⁺ ions which contribute to soil acidity. However, an increase in the pH of soil leads to the precipitation of exchangeable and soluble Al to insoluble Al hydroxides which reduces the concentration of Al in the soil solution (Kifuko *et al.*, 2007).

From Table 4.21.a it is evident that the exchangeable hydrogen, aluminium and total potential acidity in soil at harvest was highest during 2016 as well as 2017 in control (L₀). Decreasing trend in exchangeable hydrogen, aluminium and total potential acidity was observed with increasing doses of lime where the lowest values were observed in L₁₀₀. From pooled data it can be observed that exchangeable hydrogen, exchangeable aluminium and total potential acidity registered a decline of 65.8%, 43.6% and 28.8%, respectively in L₁₀₀ when compared to L₀. Similar findings have also been reported by Badole *et al* (2015) where liming decreased all forms of soil acidity and the effect was greater in full dose of lime application than in half dose of liming

Treatment	Exchangeable H ⁺ [cmol (p ⁺) kg ⁻¹]			Ez	kchangeable A [cmol (p ⁺) kg ⁻¹	1 ³⁺]	Total potential acidity [cmol (p ⁺) kg ⁻¹]		
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Soil pH levels									
pH_1	0.48	0.50	0.49	1.18	1.21	1.19	13.03	13.00	13.02
pH_2	0.36	0.30	0.33	1.08	1.09	1.08	12.35	12.05	12.20
SEm±	0.03	0.03	0.02	0.02	0.01	0.01	0.17	0.17	0.12
CD (P=0.05)	0.07	0.09	0.06	0.05	0.04	0.03	0.47	0.47	0.33
Lime levels									
L_0	0.79	0.72	0.76	1.52	1.59	1.56	15.01	14.83	14.92
L ₃₀	0.31	0.30	0.31	1.13	1.13	1.13	13.16	12.98	13.07
L_{60}	0.30	0.32	0.31	0.98	0.99	0.98	11.90	11.76	11.83
L_{100}	0.26	0.25	0.26	0.88	0.89	0.88	10.70	10.54	10.62
SEm±	0.04	0.04	0.03	0.02	0.02	0.01	0.23	0.23	0.17
CD (P=0.05)	0.10	0.13	0.08	0.07	0.05	0.04	0.67	0.67	0.46
Varieties									
\mathbf{V}_1	0.44	0.41	0.42	1.11	1.14	1.13	12.68	12.52	12.60
V_2	0.40	0.40	0.40	1.15	1.15	1.15	12.71	12.54	12.62
V ₃	0.41	0.39	0.40	1.13	1.15	1.14	12.69	12.53	12.61
SEm±	0.03	0.04	0.02	0.02	0.02	0.01	0.20	0.20	0.14
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS

Table 4.21.a: Exchangeable H⁺, Al³⁺ and total potential acidity of post-harvest soil as affected by soil pH, lime and varieties

Soil pH levels		Exchangeable H^+ [cmol (p ⁺) kg ⁻¹]												
	Lime levels				Lime levels				Lime levels					
	L ₀	L ₃₀	L ₆₀	L ₁₀₀	L ₀	L ₃₀	L ₆₀	L ₁₀₀	L ₀	L ₃₀	L ₆₀	L ₁₀₀		
	2016			2017				Pooled						
pH_1	0.96	0.34	0.31	0.30	0.93	0.38	0.35	0.32	0.95	0.36	0.33	0.31		
pH_2	0.62	0.29	0.29	0.22	0.51	0.23	0.28	0.18	0.57	0.26	0.29	0.20		
SEm±	0.05				0.06			0.04						
CD (P=0.05)	0.15				0.18			0.11						
	Exchangeable Al^{3+} [cmol (p ⁺) kg ⁻¹]													
	2016				2017				Pooled					
pH_1	1.64	1.12	1.04	0.92	1.72	1.13	1.06	0.93	1.68	1.12	1.05	0.92		
pH_2	1.40	1.13	0.93	0.85	1.46	1.13	0.91	0.85	1.43	1.13	0.92	0.85		
SEm±	0.03				0.03			0.02						
CD (P=0.05)	0.09				0.07			0.06						

Table 4.21.b: Interaction effect between pH and lime on exchangeable hydrogen and aluminium of post-harvest soil

might be due to the precipitation of exchangeable Al^+ and neutralization of exchangeable H^+ in the soil.

It can be observed from Table 4.21.a that varieties did not have any significant influence on exchangeable hydrogen, aluminium and total potential acidity in the post harvest soil.

Interaction effect of pH and lime on exchangeable hydrogen and aluminium

It can be observed from Table 4.21.b that exchangeable hydrogen and aluminium in soil at harvest in both the years were found to be highest in the treatment combination pH_1L_0 . On the other hand, lowest exchangeable hydrogen and aluminium was observed in pH_2L_{100} treatment combination. On further examination of the data it can be observed that irrespective of soil pH, exchangeable hydrogen and aluminium decreased as lime levels increased under each level of soil pH. This is because of the increase in soil pH due to liming and lime is known to precipitate the Al ions to hydroxyl compounds such as Al(OH)₃ (Caires *et al.*, 2002 and Hue, 2004). This has the effect of reducing exchangeable acidity which comprises Al³⁺ and H⁺.

4.2. EFFECT OF LIME, PHOSPHORUS AND BORON ON PERFORMANCE OF MAIZE AND SOIL PROPERTIES (EXPT-II)

4.2.1 Effect of lime, phosphorus and boron on performance of maize4.2.1.1 Effect on plant height

The results on the plant height in different treatments have been presented in Table 4.22.a and Fig. 8. There was an appreciable increase in the height of the plant with the advancement of days and also significant difference among various treatments. It is apparent from the data the maximum plant height was recorded in the treatment L_{25} (25% lime of LR) at 30, 60 DAS and at harvest, while the minimum plant height was recorded in control (L₀) during 2016 and 2017. This positive response in growth to lime can be ascribed to reduction of Al-toxicity and/ increase in availability of P (Kisinyo *et al.*, 2014). The significant increase in maize growth after lime application has been reported by Brajendra *et al.* (2006); Muindi *et al.* (2015) and Opala (2017).

Plant height of maize was significantly affected by different phosphorus levels (Table 4.22.a and Fig. 8). Data regarding phosphorus levels at all crop stages during 2016 and 2017 revealed that taller plants were produced when phosphorus was applied at 90 kg ha⁻¹ which was statistically at par with phosphorus applied at 60 kg ha⁻¹. The minimum plant height at 30, 60 DAS and at harvest was recorded in the treatment P₀ (control) during 2016 and 2017. A critical examination of data revealed that application of 90 kg P₂O₅ ha⁻¹ increased pooled plant height at 30, 60 DAS and at harvest by 12.04%, 21.2% and 21.7%, respectively over control. The probable reason for the increase in plant height due to phosphorus might be that it improved the root development and nutrient absorption which had a great effect on overall growth performance which resulted in taller plants. Similar findings have been reported by Hussain *et al.* (2006); Masood *et al.* (2011); Rashid and Iqbal (2012).

From the data depicted in Table 4.22.a and illustrated in Fig. 8, the results revealed that there was a significant difference among the treatments and the maximum plant height was recorded in the treatment B_2 *i.e.*, at 2 kg B ha⁻¹ while the minimum plant height was recorded in the treatment B_0 *i.e.*, at 0 kg B ha⁻¹ during 2016 and 2017. The data further revealed that each increment in boron application resulted in significant enhancement in plant height in comparison to preceding lower levels of boron at all three growth stages during both the years of experimentation. It was also observed that B_2 level of boron increased plant height to the extent of 12.0%, 7.5% and 4.9%, respectively at 30, 60 DAS and at harvest over control. Increase in plant height due to boron application may be due to its vital role in cell wall synthesis, division, elongation and nucleic acid metabolism. Similar results have also been reported by Ceyhan *et al.* (2007) and Ahmed *et al.* (2008).

	Plant height (cm)										
Treatment	At 30 DAS			At 60 DAS			At harvest				
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled		
Lime levels											
L_0	41.96	40.18	41.07	159.13	161.79	160.46	195.00	195.82	195.41		
L ₂₅	45.48	43.14	44.31	180.56	182.25	181.41	220.73	221.34	221.04		
SEm±	0.59	0.63	0.43	0.71	0.71	0.50	0.52	0.66	0.42		
CD (P=0.05)	1.68	1.79	1.21	2.03	2.02	1.41	1.49	1.86	1.18		
Phosphorus levels											
\mathbf{P}_0	42.01	38.37	40.19	147.66	151.16	149.41	182.07	181.84	181.95		
P ₃₀	43.07	40.32	41.69	171.87	173.27	172.57	209.25	208.66	208.95		
P ₆₀	44.48	43.24	43.86	179.71	181.53	180.62	218.98	221.99	220.48		
P ₉₀	45.33	44.73	45.03	180.16	182.12	181.14	221.18	221.84	221.51		
SEm±	0.84	0.89	0.61	1.01	1.00	0.71	0.74	0.93	0.59		
CD (P=0.05)	2.38	2.53	1.71	2.86	2.85	1.99	2.11	2.64	1.67		
Boron levels											
\mathbf{B}_0	41.60	39.01	40.31	163.09	165.93	164.51	202.07	203.74	202.91		
B_1	43.71	41.49	42.60	170.08	172.89	171.49	208.02	209.59	208.81		
B_2	45.85	44.49	45.17	176.37	177.24	176.80	213.51	212.41	212.96		
SEm±	0.72	0.77	0.53	0.87	0.87	0.62	0.64	0.80	0.51		
CD (P=0.05)	2.06	2.19	1.48	2.48	2.47	1.73	1.82	2.28	1.44		

Table 4.22.a: Plant height of maize at different growth stages as affected by lime, phosphorus and boron



Fig 8: Effect of lime, phosphorus and boron on plant height of maize at 30, 60 DAS and at harvest

Interaction effect of lime and phosphorus on plant height

The interaction effect of lime and phosphorus was found to be significant at 60 DAS during 2016 and pooled (Table 4.22.b). Without lime application, plant height increased with increasing P rates, but with lime application plant height increased from 0 to 60 kg P_2O_5 ha⁻¹ and declined at 90 kg P_2O_5 ha⁻¹. The observed increase in plant height with increasing P rate in treatments with no lime application shows that P might be a limiting factor to maize growth in this soil. The observed positive effect of lime on maize growth was likely due to its effect in ameliorating aluminium toxicity. Maximum plant height was recorded in treatment combination $L_{25}P_{60}$. Significant interaction between lime and phosphorus on plant height of maize has also been reported by Opala (2017).

Interaction effect of lime, phosphorus and boron on plant height at harvest

The data indicated that the plant height at harvest was found to be significantly affected by the combined application of lime, phosphorus and boron during 2017 (Table 4.22.c). A critical examination of the data revealed that plant height was observed to increase with increase in phosphorus and boron levels irrespective of lime application. However, higher plant heights were observed with lime application and among the treatment combinations, maximum plant height was recorded in $L_{25}P_{60}B_2$ (240.60 cm) during 2017, while in pooled data maximum plant height was recorded in $L_{25}P_{90}B_2$ (239.87 cm). Treatment combinations $L_{25}P_{60}B_2$ and $L_{25}P_{90}B_2$ were found to be at par with each other during 2017 as well as in the pooled data. Minimum plant height in 2017 and pooled data was recorded in $L_{0}P_{0}B_{0}$. The results revealed that $L_{25}P_{60}B_2$ enhanced plant growth significantly compared to the other treatment combinations, except $L_{25}P_{60}B_1$ and $L_{25}P_{90}B_2$ during 2017. Application of lime along with increased nutrient input may have led to an

	Plant height at 60 DAS (cm)							
Lime levels	Phosphorus levels							
	Po	P ₃₀	P ₆₀	P100				
	2016							
L_0	138.16	161.37	166.32	170.68				
L_{25}	157.16	182.38	193.09	189.63				
SEm±	1.42							
CD (P=0.05)	4.05							

Table 4.22.b: Interaction effect between lime and phosphorus on plant height at 60 DAS

Table 4.22.c: Interaction effect of lime, phosphorus and boron on plant height at harvest

	Plant height at harvest (cm)									
	Lime levels									
Phosphorus levels		L ₀			L ₂₅					
	H	Boron level	S]	Boron levels					
	B ₀	B_1	B ₂	B ₀	B_1	B ₂				
			20)17						
P_0	159.77	170.83	172.83	192.37	197.57	197.67				
P ₃₀	189.77	195.40	203.13	217.57	222.73	223.33				
P ₆₀	209.60	209.87	209.23	228.33	234.30	240.60				
P ₉₀	205.87	210.10	213.40	226.67	235.93	239.07				
SEm±	2.27									
CD (P=0.05)	6.45									
	Pooled									
\mathbf{P}_0	160.47	170.45	175.38	190.83	196.13	198.45				
P ₃₀	192.23	195.22	199.87	216.82	223.85	225.73				
P ₆₀	205.85	207.98	209.63	227.02	232.58	239.83				
P ₉₀	203.72	209.20	214.92	226.32	235.03	239.87				
SEm±	1.45									
CD (P=0.05)	4.08									
increase in availability of nutrients which may have aided in favourable conditions for crop growth resulting in increased plant height.

4.2.1.2 Effect on number of leaves

The data on the number of leaves in different treatments have been presented in table 4.23. It is apparent from the data, that during 2016 and 2017, maximum number of leaves was recorded in the treatment L_{25} at 30, 60 DAS and at harvest while minimum number of leaves was recorded in the treatment L_0 . In general the application of lime was observed to boost the growth of the crop due to increase in the soil pH towards neutrality (Opala, 2017). The increase in number of leaves in maize due to application of lime at higher rate has been observed by Muindi *et al.* (2015).

It is apparent from Table 4.23 that during 2016 and 2017, maximum number of leaves was recorded in treatment P_{90} at all crop stages, while minimum number of leaves was recorded in the treatment P_0 . The reason for higher number of leaves at all the growth stages of crop with higher levels of phosphorus can be ascribed to the important role of phosphorus in cell division and cell enlargement (Assuero *et al.*, 2004). At harvest, application of 90 kg P_2O_5 ha⁻¹ increased pooled number of leaves by 11.2% over control (P_0). Significant increase in number of leaves due to phosphorus application has also been reported by various workers (Ayub *et al.*, 2002 and Timlin *et al*, 2017).

From the data (Table 4.23) it can be inferred that effect of boron on number of leaves at 30 DAS was non-significant. However, at 60 DAS and at harvest there was a significant difference among the treatments and maximum number of leaves was recorded in treatment B_2 , while, minimum number of leaves was recorded in treatment B_0 during both the years of experimentation. The B_2 level of boron was observed to improve the number of leaves to the extent of 6.1% in comparison to control (B_0) during harvest. Favourable conditions for crop growth due to boron application may be due to its role in

					Number of lea	ves			
Treatment		At 30 DAS			At 60 DAS			At harvest	
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Lime levels									
L_0	6.42	6.50	6.46	10.14	10.17	10.15	12.81	13.44	13.13
L ₂₅	6.69	6.75	6.72	10.83	10.86	10.85	14.17	14.53	14.35
SEm±	0.08	0.09	0.06	0.10	0.09	0.07	0.11	0.13	0.08
CD (P=0.05)	0.24	0.25	0.17	0.27	0.25	0.18	0.30	0.37	0.24
Phosphorus levels									
\mathbf{P}_0	6.28	6.33	6.31	9.72	9.83	9.78	12.67	13.11	12.89
P ₃₀	6.50	6.56	6.53	10.33	10.50	10.42	13.50	14.28	13.89
P ₆₀	6.67	6.78	6.72	10.78	10.83	10.81	13.67	14.00	13.83
P ₉₀	6.78	6.83	6.81	11.11	10.89	11.00	14.11	14.56	14.33
SEm±	0.12	0.12	0.09	0.14	0.12	0.09	0.15	0.18	0.12
CD (P=0.05)	0.34	0.35	0.24	0.39	0.35	0.26	0.43	0.52	0.33
Boron levels									
\mathbf{B}_0	6.42	6.50	6.46	10.08	10.25	10.17	13.04	13.63	13.33
\mathbf{B}_1	6.54	6.67	6.60	10.46	10.54	10.50	13.50	13.96	13.73
B ₂	6.71	6.71	6.71	10.92	10.75	10.83	13.92	14.38	14.15
SEm±	0.10	0.11	0.07	0.12	0.11	0.08	0.13	0.16	0.10
CD (P=0.05)	NS	NS	NS	0.34	0.31	0.22	0.37	0.45	0.29

Table 4.23: Number of leaves per plant of maize at different growth stages as affected by lime, phosphorus and boron

forming and strengthening cell walls leading to growth of growing tissues and hence plant development. These results are in accordance with those of Soomro *et al.* (2011) and Borase *et al.* (2018).

4.2.1.3 Effect on leaf area index

The results on leaf area index in different treatments have been presented in table 4.24. It is apparent from the data that the leaf area index was greater in the treatment with lime (L_{25}) which may be attributed to reduction in aluminium toxicity which led to an overall improvement in plant growth. Lime application significantly increased leaf area index at all growth stages during both the years of experimentation over control. These results are in line with the findings of Muindi *et al.*, 2015 and Bekele *et al.*, 2018.

The leaf area index at all the stages of crop growth was found higher in those treatments where higher amount of phosphorus was applied which can be attributed to its major role in the growth of new tissue and division of cells. It is apparent from table 4.24, the maximum leaf area index was recorded in the treatment P_{90} which was at par with P_{60} , while the minimum leaf area index was recorded in the treatment P_0 at 30, 60 DAS and at harvest during 2016 and 2017. Positive association of leaf area index with increase in P levels have been reported by Hamdi and Woodard (1995); Amanullah *et al.* (2009) and Amanullah *et al.* (2010).

The effect of boron on leaf area index was found to be non-significant (Table 4.24).

4.2.1.4 Effect on cob length, cob girth and cob weight

The results on cob length, cob girth and cob weight in different treatments have been presented in Table 4.25.a. There was a significant increase in cob length, cob girth and cob weight with application of lime @ 25% lime of LR (L_{25}) which recorded maximum cob length, cob girth as well

					Leaf area inde	Х			
Treatment		At 30 DAS			At 60 DAS			At harvest	
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Lime levels									
L_0	0.60	0.63	0.61	1.24	1.27	1.25	1.16	1.17	1.17
L ₂₅	0.64	0.67	0.66	1.29	1.32	1.31	1.18	1.19	1.19
SEm±	0.01	0.01	0.01	0.005	0.006	0.004	0.004	0.005	0.003
CD (P=0.05)	0.04	0.03	0.02	0.02	0.02	0.01	0.01	0.01	0.01
Phosphorus levels									
\mathbf{P}_0	0.56	0.59	0.57	1.24	1.26	1.25	1.16	1.16	1.16
P ₃₀	0.61	0.62	0.62	1.26	1.28	1.27	1.16	1.18	1.17
P ₆₀	0.64	0.68	0.66	1.28	1.30	1.29	1.17	1.19	1.18
P ₉₀	0.67	0.71	0.69	1.29	1.33	1.31	1.18	1.20	1.19
SEm±	0.02	0.01	0.01	0.008	0.009	0.006	0.006	0.006	0.004
CD (P=0.05)	0.05	0.04	0.03	0.02	0.03	0.02	0.02	0.02	0.01
Boron levels									
\mathbf{B}_0	0.60	0.65	0.63	1.27	1.29	1.28	1.17	1.18	1.17
B_1	0.63	0.65	0.64	1.27	1.30	1.28	1.16	1.18	1.17
B_2	0.63	0.66	0.64	1.27	1.30	1.28	1.18	1.19	1.18
SEm±	0.02	0.01	0.01	0.007	0.008	0.005	0.005	0.006	0.004
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS

Table 4.24: Leaf area index of maize at different growth stages as affected by lime, phosphorus and boron

as cob weight during 2016 and 2017. Application of 25% lime of LR increased cob length, cob girth and cob weight by 2.3%, 3.0% and 21.1%, respectively over control. The beneficial effect of lime in crops is well known, which is reduction of aluminium toxicity by bringing the pH towards neutrality, hence improvement in nutrient solubility which improves yield. Similar findings have been reported by Brajendra *et al.* (2006) and Kumar *et al.* (2012).

The cob length, cob girth and cob weight in both the years was found higher in those treatments with higher level of phosphorus. A critical examination of data indicated that each increasing level of phosphorus enhanced cob length, cob girth and cob weight significantly as compared to preceding lower levels in case of pooled values. It is apparent from table 4.25.a that the maximum cob length, cob girth as well as cob weight was recorded in the treatment P₉₀. The P₉₀ level of phosphorus increased pooled cob length, cob girth, cob weight by 12.2%, 8.8% and 22.5%, respectively over control which can be ascribed to the important role of phosphorus in cell division and cell enlargement (Assuero *et al.*, 2004). However, treatment P₉₀ was at par with treatment P₆₀ in case of cob length. As phosphorus is responsible for good root growth which directly affects the overall plant performance, phosphorus at 0 kg ha⁻¹ (P₀) resulted in minimum cob length, cob girth and cob weight during 2016 as well as 2017. Similar findings have been reported by Khan *et al.* (2017); Pal *et al.* (2017) and Sharma *et al.* (2018).

The effect of boron was observed to have a significant difference among the treatments. The highest cob length, cob girth and cob weight was recorded in treatment B_2 while the lowest cob length, cob girth and cob weight was recorded in the treatment B_0 during 2016 and 2017. From pooled data, it can be observed that B_2 level of boron enhanced cob length, cob girth and cob weight by 3.2%, 2.2% and 5.2%, respectively over control. The significant effect of boron application might be due to its role in pollen tube formation, increasing

Treatment		Cob length (cr	n)		Cob girth (cm))	Cob weight (g)		
Ireatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Lime levels									
L_0	17.99	18.39	18.19	14.40	14.45	14.43	82.44	82.27	82.35
L ₂₅	18.49	18.73	18.61	14.86	14.87	14.87	99.55	99.86	99.71
SEm±	0.05	0.05	0.04	0.06	0.06	0.04	0.32	0.26	0.21
CD (P=0.05)	0.14	0.14	0.10	0.18	0.17	0.12	0.91	0.74	0.58
Phosphorus levels									
\mathbf{P}_0	17.01	17.42	17.21	14.02	14.06	14.04	79.51	79.23	79.37
P ₃₀	18.00	18.48	18.24	14.44	14.46	14.45	90.10	90.72	90.41
P ₆₀	18.71	18.99	18.85	14.80	14.82	14.81	97.15	97.00	97.08
P ₉₀	19.26	19.36	19.31	15.25	15.31	15.28	97.21	97.31	97.26
SEm±	0.07	0.07	0.05	0.09	0.09	0.06	0.45	0.37	0.29
CD (P=0.05)	0.20	0.20	0.14	0.25	0.24	0.17	1.29	1.04	0.82
Boron levels									
\mathbf{B}_0	17.91	18.28	18.10	14.48	14.49	14.49	88.59	88.43	88.51
B_1	18.25	18.59	18.42	14.62	14.67	14.64	91.71	91.19	91.45
B_2	18.57	18.81	18.69	14.79	14.83	14.81	92.67	93.57	93.12
SEm±	0.06	0.06	0.04	0.08	0.07	0.05	0.39	0.32	0.25
CD (P=0.05)	0.17	0.18	0.12	0.22	0.21	0.15	1.12	0.90	0.71

Table 4.25.a: Cob length, cob girth and cob weight of maize as affected by lime, phosphorus and boron

the efficiency of fertilization process. This is in congruence with the findings of Tahir *et al.* (2012) and Arunkumar and Srinivasa (2018).

Interaction effect of lime and phosphorus on cob weight

From table 4.25.b, it can be observed that in both the years, irrespective of lime application, cob weight increased with increase in P levels. Without lime application (L₀), cob weight was found to increase up to 90 kg ha⁻¹ (P₉₀), while with the application of lime, cob weight increased up to 60 kg P₂O₅ ha⁻¹ (P₆₀) and declined at 90 kg P₂O₅ ha⁻¹ (P₉₀). The observed increase in cob weight with increasing P rate in treatments with no lime application shows that P might be a limiting factor to maize growth in this soil. Highest cob weight was recorded under L₂₅P₆₀ treatment combination and lowest cob weight was observed under L₀P₀ treatment combination. Significant reduction of Al in the soil due to liming along with the effect of P on cell division and enlargement (Assuero *et al.*, 2004) in plants may have enhanced the cob parameter.

4.2.1.5 Effect on number of grains per row, number of rows and grains per cob

The results obtained on the number of rows per cob, number of grains per row, and number of grains per cob in different treatments has been presented in table 4.26.a. Significant response to application of lime with respect to these parameters were observed. Maximum values of all the three parameters were observed in L_{25} while minimum was recorded in the treatment L_0 during 2016 and 2017. The increase in these parameters due to liming might be attributed to the reduction in acidity and increase in nutrient availability. Similar results have also been reported by Sierra *et al.* (2003); Adikuru *et al.* (2019) and Devkota *et al.* (2019).

The number of rows per cob was found to be non-significant, while number of grains per row, and number of grains per cob was found to be significant with phosphorus application. It is apparent from Table 4.26.a that

	Cob weight (g)								
x· 1 1		Phosphor	rus levels						
Lime levels	P ₀	P ₃₀	P ₆₀	P ₉₀					
		20	16						
L ₀	70.50	80.53	87.74	90.97					
L ₂₅	88.51	99.67	106.56	103.44					
SEm±		0.0	64						
CD (P=0.05)		1.82							
		2017							
L ₀	70.07	80.99	87.86	90.14					
L ₂₅	88.40	100.46	106.13	104.47					
SEm±		0.5	52						
CD (P=0.05)		1.4	48						
		Poc	oled						
L ₀	70.28	80.76	87.80	90.56					
L ₂₅	88.46	100.06	106.35	103.96					
SEm±		0.4	41						
CD (P=0.05)		1.1	16						

Table 4.25.b: Interaction effect between lime and phosphorus on cob weight

Tractment	No	of grains per	row	No	No of rows per cob			No of grains per cob		
Ireatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled	
Lime levels										
L_0	20.06	20.25	20.15	13.33	13.47	13.40	283.17	287.36	285.26	
L ₂₅	22.81	23.36	23.08	14.92	15.00	14.96	331.69	340.97	336.33	
SEm±	0.08	0.10	0.07	0.15	0.10	0.09	6.52	3.49	3.70	
CD (P=0.05)	0.24	0.30	0.19	0.42	0.29	0.25	18.54	9.92	10.38	
Phosphorus levels										
\mathbf{P}_0	19.56	19.61	19.58	13.72	13.94	13.83	264.83	270.61	267.72	
P ₃₀	21.50	21.44	21.47	14.17	14.28	14.22	318.94	307.78	313.36	
P ₆₀	21.83	22.72	22.28	14.28	14.44	14.36	321.06	338.50	329.78	
P ₉₀	22.83	23.44	23.14	14.33	14.28	14.31	324.89	339.78	332.33	
SEm±	0.12	0.15	0.09	0.21	0.14	0.13	9.22	4.93	5.23	
CD (P=0.05)	0.34	0.42	0.26	NS	NS	NS	26.23	14.02	14.68	
Boron levels										
\mathbf{B}_0	20.92	21.25	21.08	13.96	14.04	14.00	290.50	298.96	294.73	
B_1	21.50	21.83	21.67	14.08	14.38	14.23	308.71	315.00	311.85	
B ₂	21.88	22.33	22.10	14.33	14.29	14.31	323.08	328.54	325.81	
SEm±	0.10	0.13	0.08	0.18	0.13	0.11	7.99	4.27	4.53	
CD (P=0.05)	0.29	0.36	0.23	NS	NS	NS	22.71	12.14	12.71	

Table 4.26.a: Number of grains per row, number of rows and number of grains per cob as affected by lime, phosphorus and boron

the maximum number of grains per row and grains per cob was recorded in treatment P_{90} while the minimum was recorded in the treatment P_0 during 2016 and 2017. From pooled data, P_{90} level of phosphorus increased number of grains per row and number of grains per cob to the extent of 18.8% and 24.1%, respectively, over control. However, from pooled data it was observed that treatment P_{90} was at par with treatment P_{60} in case of number of grains per cob. The increase in phosphorus level might have partitioned greater amount of assimilates to ears which may have resulted in an increase in these cob characteristics. Significant effect of phosphorus on these yield attributes have also been reported by Alias *et al.* (2003); Khan *et al.* (2005) and Sadiq *et al.* (2017).

The data (Table 4.26.a) showed that there was no significant difference in the number of rows per cob with respect to application of boron while significant difference was observed in case of number of grains per row and per cob. The maximum number of grains per row and number of grains per cob was observed in the treatment B_2 which elucidates the role of boron in pollen viability and seed production of crops. Minimum reading was observed in treatment B_0 . The present findings are well in agreement with that of Rahim *et al.* (2004) and Tahir *et al.* (2012).

Interaction effect of lime and phosphorus on number of grains per row and number of grains per cob

From the data presented in Table 4.26.b, it can be observed that irrespective of liming, number of grains per row and number of grains per cob increased with application of increasing phosphorus levels. The maximum number of grains per row and number of grains per cob was recorded in $L_{25}P_{90}$ and $L_{25}P_{60}$ treatment combination, respectively, while minimum was recorded in L_0P_0 during 2016 as well as 2017. In case of number of grains per row, treatment combination $L_{25}P_{90}$ and $L_{25}P_{60}$ were found to be at par with each

					N	umber of g	rains per rov	W				
Lime levels		Phosphor	us levels			Phospho	rus levels			Phosphor	rus levels	
	P ₀	P ₃₀	P ₆₀	P ₉₀	\mathbf{P}_0	P ₃₀	P ₆₀	P ₉₀	P ₀	P ₃₀	P ₆₀	P ₉₀
		20	16			20)17			Poo	oled	
L ₀	18.22	20.00	20.11	21.89	18.33	19.67	20.89	22.11	18.28	19.83	20.50	22.00
L ₂₅	20.89	23.00	23.56	23.78	20.89	23.22	24.56	24.78	20.89	23.11	24.06	24.28
SEm±		0.1	17			0.	21			0.	13	
CD (P=0.05)	0.47				0.	59			0.	37		
					N	umber of g	rains per co	b				
Lime levels			Phosphor	us levels		Phosphorus levels						
	\mathbf{P}_0		P ₃₀	P ₆₀		P ₉₀	P ₀ P ₃₀ P ₆₀				P ₉₀	
			20	16					Poo	oled		
L_0	238.89	2	71.11	303.78	3	318.89	240.78	2	276.44	307.6	1	316.22
L ₂₅	290.78	290.78 366.78 338.33			3	330.89	294.67 350.28 351.94				1	348.44
SEm±			13.	04					7.	40		
CD (P=0.05)			37.	09					20	.76		

Table 4.26.b: Interaction effect between lime and phosphorus on number of grains per row and number of grains per cob

other. In case of number of grains per cob, treatment combination $L_{25}P_{60}$ was found to be at par with $L_{25}P_{30}$ and $L_{25}P_{90}$.

4.2.1.6 Effect on test weight, grain yield and stover yield of maize

The results on the test weight, grain yield and stover yield of maize in different treatments have been presented in Table 4.27.a and Fig 9. Effect of lime was found to be significant with respect to test weight, grain and stover yield, which was found to be highest in the treatment where lime was applied i.e., at L_{25} , while the lowest was observed in the control treatment (L_0) during 2016 and 2017. The L_{25} level of lime increased pooled grain and stover yield by 24.6% and 23.0%, respectively over control (L_0). Liming is essential in order to achieve optimum yields of crops that are grown on acid soils, because it increases pH and reduces acidity-related constraints (Fageria and Baligar, 2008). Furthermore, lime application enhanced yield attributes which resulted in increased grain yield.

From the Table 4.27.a and Fig 9, it can be observed that effect of phosphorus was significant with respect to grain and stover yield during both the years. The maximum grain and stover yield was recorded in treatment P_{90} , whereas the minimum grain and stover yield was recorded in the treatment P_0 during 2016 as well as 2017. However, P_{90} was at par with P_{60} in case of both grain and stover yield during 2016 as well as 2017. Irrespective of treatment and year, from pooled, the grain yield ranged from 59.93 to 77.47 g pot⁻¹ and stover yield ranged from 101.01 to 125.57 g pot⁻¹. Each increasing level of phosphorus significantly enhanced pooled grain and stover yield as compared to preceding lower level of phosphorus. The P_{90} level of phosphorus increased grain and stover yield by 24.3% and 22.1%, respectively over control. The increase in grain and stover yield due to phosphorus application may be due to higher yield components like number of grains per cob and higher growth with the addition of phosphorus as P plays important role in plant metabolism,

Tuestueset		Test weight ((g)	Gr	ain yield (g po	ot ⁻¹)	Stover yield (g pot ⁻¹)		
I reatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled
Lime levels									
L_0	22.41	22.53	22.47	61.93	62.43	62.18	101.66	102.47	102.06
L ₂₅	24.93	24.83	24.88	77.63	77.31	77.47	126.21	124.93	125.57
SEm±	0.17	0.20	0.13	0.26	0.38	0.23	1.00	0.80	0.64
CD (P=0.05)	0.48	0.57	0.37	0.73	1.08	0.64	2.83	2.28	1.79
Phosphorus levels									
\mathbf{P}_0	23.45	23.66	23.56	59.70	60.15	59.93	101.46	100.56	101.01
P ₃₀	23.66	23.70	23.68	70.97	71.09	71.03	110.37	109.30	109.83
P ₆₀	23.86	23.66	23.76	73.78	73.92	73.85	121.45	120.64	121.04
P ₉₀	23.70	23.70	23.70	74.66	74.30	74.48	122.46	124.28	123.37
SEm±	0.24	0.28	0.19	0.36	0.54	0.32	1.41	1.13	0.90
CD (P=0.05)	NS	NS	NS	1.03	1.53	0.91	4.01	3.22	2.54
Boron levels									
\mathbf{B}_0	23.57	23.61	23.59	66.19	64.79	65.49	108.57	105.85	107.21
\mathbf{B}_1	23.68	23.70	23.69	71.19	71.52	71.35	113.88	117.36	115.62
B ₂	23.75	23.73	23.74	71.96	73.30	72.63	119.35	117.88	118.61
SEm±	0.21	0.25	0.16	0.31	0.47	0.28	1.22	0.98	0.78
CD (P=0.05)	NS	NS	NS	0.89	1.33	0.79	3.47	2.79	2.20

Table 4.27.a: Test weight, grain yield and stover yield of maize as affected by lime, phosphorus and boron



Fig 9: Effect of lime, phosphorus and boron on grain yield and stover yield of maize

finally leading to enhanced yield. Similar results have also been reported by Kumar *et al.* (2017). Results on higher grain and straw yields with higher levels of phosphorus according to the present investigation have also been reported by Arya and Singh (2000).

The effect of boron levels on test weight was non-significant. However, it can be observed that the treatment with higher levels of boron exhibited higher test weight. Significantly highest grain and stover yield was recorded in B_0 during 2016 and 2017. It was also observed that each increasing level of boron significantly enhanced grain and stover yield in comparison to preceding lower level of boron. From pooled data it can be observed that B_2 level of boron increased pooled grain and stover yield by 10.9% and 10.6% over control. The improvement in maize grain and stover yield may be attributed to the complementary role of boron in the reproduction and vegetative stage of plants. The present finding is in agreement with that of Akhter and Mahmud (2009).

Interaction effect of lime and phosphorus on grain yield

From the Table 4.27.b it is evident that effect of lime and phosphorus was found to be significant on grain yield in 2016 and 2017. It could be observed that increasing levels of lime as well as phosphorus significantly increased the grain yield in both the years. However, in non-limed pots (L₀), highest grain yield was recorded in P₉₀, whereas in limed pots (L₂₅), highest grain yield was observed in P₆₀. Thus, with lime application, phosphorus application up to P₆₀ was optimum to achieve the maximum grain yield. Overall, maximum grain yield was observed in L₀P₀ during both the years. These results are in line with the findings of Venkatesh (2002); Muindi *et al.* (2015) who reported a significant positive effect of lime and phosphorus on grain yields of crops.

		Grain yield (g pot ⁻¹)							
Lime levels		Phosphor	rus levels						
	P ₀	P ₃₀	P ₆₀	P ₉₀					
		20	16						
L ₀	52.20	62.31	64.83	68.37					
L ₂₅	67.21	79.64	82.73	80.96					
SEm±		0.:	51						
CD (P=0.05)		1.4	46						
	2017								
L_0	52.52	63.41	65.61	68.16					
L ₂₅	67.79	78.78	82.23	80.44					
SEm±		0.	76						
CD (P=0.05)		2.	16						
		Poo	oled						
L_0	52.36	62.86	65.22	68.27					
L ₂₅	67.50	79.21	82.48	80.70					
SEm±	0.46								
CD (P=0.05)		1.2	29						

Table 4.27.b: Interaction effect between lime and phosphorus on grain yield

4.2.1.7 Nitrogen content in grain and stover

It could be inferred from Table 4.28 that lime had a significant effect on nitrogen content in both grain and stover during 2016 and 2017. From pooled data, it can be seen that irrespective of treatment and year, nitrogen content in grain and stover ranged from 1.00 to 1.20% and from 0.56 to 0.68%, respectively. The nitrogen content in both grain and stover was found to be higher in the treatment where lime was applied (L_{25}) as compared to control (L_0). Pooled data reflected an increase by 20.0% and 21.4% in the nitrogen content in grain and stover, respectively when lime was applied. Quaggio *et al.* (1991) reported that liming increased root growth and nutrient absorption, mainly nitrogen in maize. Similar findings where application of lime increased the nitrogen content of crop have also been reported by Rosolem and Caires (1998) and Bhat *et al.* (2007).

Effect of phosphorus on nitrogen content in both grain and stover was found to be significant where highest phosphorus content was observed in the treatment P₉₀ and the lowest in control (P₀). On the basis of pooled data it could be observed that there was an increase in nitrogen content in grain by 10.7% and in stover by 17.5% in the P₉₀ treatment when compared to control. However, treatment P₉₀ was at par with treatment P₆₀ in case of both grain and stover during both the years. The maximum nitrogen content was recorded at higher dose of phosphorus because of the increase in nutrient supply which increased the nutrient contents in both grain and stover. Increase in nutrient supply due to phosphorus application is due to its positive effect on better root system (Sharma *et al.*, 2008) which helps the plant to effectively absorb nutrients from the soil. Similar findings where phosphorus increased the nitrogen content in plants have also been reported by Prajapati *et al.* (2013); Kumar *et al.* (2015); Snehlata (2015) and Etabo *et al.* (2018).

Turoturout	N co	ntent in grai	n (%)	N coi	ntent in stov	er (%)
Treatment	2016	2017	Pooled	2016	2017	Pooled
Lime levels						
L_0	0.98	1.01	1.00	0.55	0.57	0.56
L ₂₅	1.19	1.21	1.20	0.69	0.68	0.68
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.03	0.03	0.02	0.04	0.04	0.03
Phosphorus levels						
\mathbf{P}_0	1.02	1.04	1.03	0.58	0.56	0.57
P ₃₀	1.10	1.11	1.10	0.61	0.59	0.60
P ₆₀	1.12	1.13	1.13	0.63	0.66	0.64
P ₉₀	1.11	1.17	1.14	0.66	0.68	0.67
SEm±	0.02	0.01	0.01	0.02	0.02	0.01
CD (P=0.05)	0.04	0.04	0.03	0.06	0.05	0.04
Boron levels						
\mathbf{B}_0	1.02	1.05	1.04	0.58	0.58	0.58
B_1	1.11	1.13	1.12	0.62	0.62	0.62
B_2	1.12	1.14	1.13	0.66	0.67	0.66
SEm±	0.01	0.01	0.01	0.02	0.02	0.01
CD (P=0.05)	0.04	0.04	0.03	0.05	0.05	0.03

Table 4.28: Nitrogen content in grain and stover of maize as affected by lime, phosphorus and boron

Effect of boron on nitrogen content in grain and stover was significant as evident from Table 4.28. Highest boron content in both grain and stover was observed in the treatment B_2 , whereas lowest boron content was observed in the treatment B_0 during both the years. Boron in B_2 treatment increased the nitrogen content by 15% in grain and by 22.2% in stover over B_0 treatment. It could also be observed that in case of nitrogen content in grain, B_2 was at par with B_1 . Positive effect of boron on nitrogen content in maize has been reported by Adem *et al.* (2011) and Günes *et al.* (2011).

4.2.1.8 Phosphorus content in grain and stover

As evident from Table 4.29, it can be observed that lime had a significant effect on the phosphorus content in both grain and stover, where highest value was observed at 25% lime of LR (L_{25}) compared to 0% lime of LR (L_0). Irrespective of treatment and year, it can be observed from pooled data that phosphorus content in grain and stover ranged from 0.35 to 0.45% and from 0.12 to 0.16%, respectively. On the basis of pooled data there was an increase by 17.9 % and 13.3% in the phosphorus content in grain and stover, respectively when lime was applied. Verde (2013) reported that lime improved soil acidity preventing P fixation in the soil, enhancing the P solubility which make the nutrient more available and thus increasing its content. Similar findings where lime increased the phosphorus content in plants have been reported by Bhat *et al.* (2007); Busari *et al.* (2008) and Kovacevic and Rastija (2010).

Data pertaining to phosphorus content as affected by phosphorus (Table 4.29) revealed that there was a significant difference among the treatments during 2016 and 2017. The maximum phosphorus content was recorded in P_{90} in grain during both the years, while in stover it was recorded in treatments P_{60} and P_{90} during 2016 and in P_{60} during 2017. Minimum phosphorus content in both grain and stover was recorded in control (P_0) during 2016 as well as 2017. On the basis of pooled data there was an increase by 28.6% in case of

	P co	ntent in grain	n (%)	P cor	ntent in stove	er (%)
Ireatment	2016	2017	Pooled	2016	2017	Pooled
Lime levels						
L_0	0.37	0.39	0.38	0.12	0.15	0.13
L ₂₅	0.43	0.46	0.45	0.15	0.16	0.16
SEm±	0.008	0.009	0.006	0.007	0.005	0.005
CD (P=0.05)	0.02	0.03	0.02	0.02	0.02	0.01
Phosphorus levels						
\mathbf{P}_0	0.34	0.36	0.35	0.11	0.14	0.12
P ₃₀	0.40	0.44	0.42	0.13	0.16	0.15
P ₆₀	0.43	0.45	0.44	0.15	0.17	0.16
P ₉₀	0.44	0.45	0.45	0.15	0.16	0.16
SEm±	0.011	0.013	0.008	0.010	0.008	0.006
CD (P=0.05)	0.03	0.04	0.02	0.03	0.02	0.02
Boron levels						
\mathbf{B}_0	0.38	0.39	0.38	0.12	0.13	0.13
B_1	0.40	0.44	0.42	0.13	0.17	0.15
B ₂	0.43	0.45	0.44	0.15	0.17	0.16
SEm±	0.010	0.011	0.007	0.009	0.007	0.006
CD (P=0.05)	0.03	0.03	0.02	0.03	0.02	0.02

Table 4.29: Phosphorus content in grain and stover of maize as affected by lime, phosphorus and boron

phosphorus content in grain and an increase by 33.3% in case of phosphorus content in stover due to P_{90} over control. From pooled data, in case of P content in grain, treatment P_{90} was at par with P_{60} , while in case of P content in stover treatment P_{90} was at par with P_{30} and P_{60} . Increase in phosphorus content with increasing P application rates have been reported by Rashid and Iqbal (2012) and Saeed *et al.* (2017).

It can be observed from Table 4.29 that there was significant effect of boron on phosphorus content in grain as well as stover during 2016 and 2017. Maximum P content in both grain and stover was observed in treatment B_2 and minimum was observed in B_0 during both the years of experimentation. On the basis of pooled data there was an increase by 15.8% in boron content in grain and an increase by 23.1% in boron content in stover due to B_2 over control. Also from pooled data, it can be observed that treatment B_2 was at par with treatment B_1 . Positive effect of boron on P content in maize has been reported by Adiloglu and Adiloglu (2006); Adem *et al.* (2011) and Gunes *et al.* (2011).

4.2.1.9 Potassium content in grain and stover

The data indicated that the potassium content in both grain and stover significantly increased with the application of lime when compared to control (Table 4.30). Thus, maximum potassium content in grain and stover was observed in L_{25} , while, minimum was observed in L_0 during 2016 and 2017. Potassium content in grain and stover, irrespective of treatments and years varied from 0.53 to 0.61% and from 1.07 to 1.16%, respectively. It was also observed that stover contained more potassium content than grain. Pooled data reflected an increase by 10.9% in case of potassium content in grain and 4.6% in case of potassium content in stover due to liming. The affirmative influence of liming on K concentration in plants may be due to the synergistic relation between K and Ca. This is in congruence with the findings of Busari *et al.* (2008); Barman *et al.* (2014) and Bhindhu *et al.* (2018).

T ()	Ксо	ntent in grai	n (%)	К со	ntent in stov	er (%)
Ireatment	2016	2017	Pooled	2016	2017	Pooled
Lime levels						
L_0	0.53	0.57	0.55	1.10	1.08	1.09
L ₂₅	0.59	0.62	0.61	1.15	1.14	1.14
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.03	0.03	0.02	0.03	0.03	0.02
Phosphorus levels						
\mathbf{P}_0	0.52	0.54	0.53	1.08	1.05	1.07
P ₃₀	0.55	0.60	0.57	1.10	1.09	1.10
P ₆₀	0.58	0.60	0.59	1.15	1.13	1.14
P ₉₀	0.59	0.63	0.61	1.17	1.16	1.16
SEm±	0.02	0.02	0.01	0.01	0.01	0.01
CD (P=0.05)	0.05	0.05	0.03	0.04	0.04	0.03
Boron levels						
\mathbf{B}_0	0.55	0.59	0.57	1.12	1.10	1.11
B_1	0.57	0.60	0.58	1.13	1.11	1.12
B_2	0.56	0.59	0.58	1.13	1.10	1.12
SEm±	0.01	0.02	0.01	0.01	0.01	0.01
CD (P=0.05)	NS	NS	NS	NS	NS	NS

Table 4.30: Potassium content in grain and stover of maize as affected by lime, phosphorus and boron

Effect of phosphorus on potassium content in grain and stover was also found to be significant. Highest potassium content in grain and stover was observed in treatment P₉₀ and the lowest in control (P₀) during 2016 and 2017. On the basis of pooled data it could be observed that there was an increase in potassium content in grain by 15.1% and in stover by 8.4% in the treatment P₉₀ when compared to control. However, treatment P₉₀ was at par with treatment P₆₀ in case of potassium content in both grain and stover in 2016 as well as 2017. Increase in potassium content due to increasing doses of phosphorus application has been reported by Kumar *et al.* (2015) and Snehlata (2015).

Effect of boron was found to be non- significant in case of potassium content in grain and stover (Table 4.30).

4.2.1.10 Calcium content in grain and stover

The data presented in Table 4.31 shows that there was a significant influence of lime on calcium content in grain and stover over control. Thus, highest calcium content in grain and stover was recorded in L_{25} level and lowest calcium content in grain and stover was recorded in L_0 during 2016 and 2017. A critical examination of the pooled data indicated that irrespective of treatments and year, calcium content in grain and stover ranged from 0.32 to 0.53% and from 0.35 to 0.49%, respectively. Pooled data observation showed that due to liming there was an increase by 60.0% in case of calcium content in grain and 58.3% in case of calcium content in stover over L_0 . Enhanced concentration of calcium in plant as a result of liming is related to the addition of significant amount of Ca through lime (CaCO₃) which is in line with the findings of Kovacevic and Rastija (2010); Barman *et al.* (2014) and Bhindhu *et al.* (2018).

Effect of phosphorus on calcium content in grain and stover was found to be significant (Table 4.31). Highest calcium content was observed in

Treatment	Ca co	ontent in gra	in (%)	Ca co	Ca content in stover (%)			
Treatment	2016	2017	Pooled	2016	2017	Pooled		
Lime levels								
L_0	0.35	0.34	0.35	0.36	0.35	0.36		
L ₂₅	0.56	0.55	0.56	0.57	0.57	0.57		
SEm±	0.02	0.02	0.01	0.01	0.01	0.01		
CD (P=0.05)	0.05	0.05	0.04	0.03	0.04	0.02		
Phosphorus levels								
\mathbf{P}_0	0.38	0.38	0.38	0.40	0.41	0.40		
P ₃₀	0.47	0.44	0.46	0.47	0.48	0.47		
P ₆₀	0.49	0.48	0.48	0.46	0.48	0.47		
P ₉₀	0.49	0.49	0.49	0.52	0.49	0.50		
SEm±	0.03	0.03	0.02	0.02	0.02	0.01		
CD (P=0.05)	0.07	0.08	0.05	0.05	0.05	0.04		
Boron levels								
\mathbf{B}_0	0.47	0.46	0.47	0.47	0.47	0.47		
B_1	0.46	0.44	0.45	0.47	0.47	0.47		
B ₂	0.45	0.44	0.45	0.46	0.46	0.46		
SEm±	0.02	0.02	0.02	0.01	0.02	0.01		
CD (P=0.05)	NS	NS	NS	NS	NS	NS		

Table 4.31: Calcium content in grain and stover of maize as affected by lime, phosphorus and boron

treatment P₉₀ while lowest was observed in control (P₀) in case of both grain and stover in both the years. Pooled data reflected an increase in calcium content by 28.9% and 24.9% in grain and stover, respectively in treatment P₉₀ over P₀. Treatment P₉₀ was found to be at par with treatments P₃₀ and P₆₀ in Ca content in both grain and stover. This result is in line with the findings of Kumar *et al.* (2017) and Wafula *et al.* (2018) where they observed an increase in plant calcium contents with increase in rate of phosphorus application.

The effect of boron on calcium content in grain and stover was found to be non-significant as can be observed from Table 4.31.

4.2.1.11 Boron content in grain and stover

The data (Table 4.32.a) indicated that B content in grain and stover was recorded highest in the treatment L_{25} and lowest in treatment L_0 during 2016 and 2017, where on the basis of pooled data, liming (L_{25}) increased the boron content in grain by 2.1% and in stover by 5.0%. Irrespective of treatments and year, from pooled data it can be observed that boron content in grain and stover ranged from 9.04 to 12.57 mg kg⁻¹ and from 3.18 to 4.19 mg kg⁻¹, respectively. The increase in boron content in the plant due to liming may be due to increase in its availability in the soil as a result of neutralization of soil acidity which releases boron in the soil solution (Sarkar *et al.*, 2015). Similar findings have also been reported by Barman *et al.* (2014).

The data pertaining to effect of phosphorus on boron content (Table 4.32.a) revealed that there was no significant difference among the treatments during 2016 and 2017.

The effect of boron on boron content in grain and stover was significant where there was an increase in the content with increasing levels of boron. The highest boron content for both grain and stover was recorded in B_2 while the minimum boron content in grain and stover was observed in B_0 during 2016 and 2017. The B_2 level of boron enhanced pooled B content in grain and stover

Treatment	B conte	ent in grain (mg kg ⁻¹)	B content in stover (mg kg ⁻¹)		
	2016	2017	Pooled	2016	2017	Pooled
Lime levels						
L_0	10.81	10.86	10.83	3.73	3.76	3.75
L ₂₅	11.03	11.08	11.06	3.93	3.95	3.94
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.02	0.02	0.02	0.03	0.03	0.02
Phosphorus levels						
\mathbf{P}_0	10.91	10.97	10.94	3.82	3.84	3.83
P ₃₀	10.93	10.96	10.94	3.83	3.85	3.84
P ₆₀	10.92	10.98	10.95	3.85	3.87	3.86
P ₉₀	10.91	10.96	10.94	3.84	3.87	3.85
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	NS	NS	NS	NS	NS	NS
Boron levels						
\mathbf{B}_0	9.01	9.07	9.04	3.16	3.20	3.18
B_1	11.20	11.25	11.22	4.16	4.18	4.17
B_2	12.55	12.60	12.57	4.18	4.21	4.19
SEm±	0.01	0.01	0.01	0.01	0.01	0.01
CD (P=0.05)	0.03	0.03	0.02	0.04	0.03	0.02

Table 4.32.a: Boron content in grain and stover of maize as affected by lime, phosphorus and boron

	B content in stover (%)						
Lime levels	Boron levels						
	B ₀ B ₁		B ₂				
	2016						
L ₀	3.09	4.04	4.07				
L ₂₅	3.24	4.27	4.29				
SEm±	0.02						
CD (P=0.05)		0.05					
	2017						
L ₀	3.14	4.07	4.09				
L ₂₅	3.26	4.28	4.32				
SEm±	0.02						
CD (P=0.05)	0.04						
	Pooled						
L ₀	3.12	4.05	4.08				
L ₂₅	3.25	4.28	4.31				
SEm±	0.01						
CD (P=0.05)		0.03					

Table 4.32.b: Interaction effect between lime and boron on boron content in stover

by 39.0% and 31.8%, respectively over control. However, in case of B content in stover treatment B_2 was at par with treatment B_1 . The present finding where B content in the plant increased with application of boron is in line with the findings of Barman *et al.* (2014) and Sahin (2014) who reported that where B was supplemented through external sources, B concentration in plant increased.

Interaction effect of lime and boron on boron content in stover

From Table 4.32.b, it is evident that interaction effect of lime and boron had a significant positive effect on boron content in stover during 2016 and 2017. The highest B content in both lime levels of L_0 (control) and L_{25} (25% lime of LR) was observed in B_2 (2 kg B ha⁻¹) level during both the years. Also, in both the years in both lime levels, treatment B_2 was observed to be at par with B_1 . Thus, in both the treatments, L_0 (control) and L_{25} (25% lime of LR), boron level of B_1 was found to be optimum for increasing B content in stover.

4.2.1.12 Nitrogen uptake in grain and stover

The results on nitrogen uptake in grain and stover due to different treatments have been presented in Table 4.33.a and Fig 10. The nitrogen uptake in grain and stover showed significant response to lime. It is apparent from the data that maximum nitrogen uptake was recorded in L_{25} while the minimum was recorded in L_0 for grain and stover during 2016 and 2017. Pooled data reflected that there was an increase in nitrogen uptake by 49.8% and 50.3% in grain and stover, respectively due to liming (L_{25}) over control (L_0). The B content in the plant increased with application of boron is in line with the findings of Barman *et al.* (2014) and Sahin (2014) who reported that where B was supplemented through external sources, B concentration in plant increased. increase in nitrogen uptake might be due to the increase in the available soil nitrogen content. Increase in nitrogen uptake due to liming has been reported in earlier works, where maize recorded maximum uptake of N, P

Treatment	N uptak	te in grain (n	ng pot ⁻¹)	N uptake in stover (mg pot ⁻¹)		
	2016	2017	Pooled	2016	2017	Pooled
Lime levels						
L ₀	611.59	634.12	622.86	561.71	586.36	574.04
L ₂₅	927.10	939.50	933.30	872.27	853.79	863.03
SEm±	7.24	7.28	5.13	15.31	14.91	10.69
CD (P=0.05)	20.58	20.71	14.41	43.54	42.40	30.00
Phosphorus levels						
\mathbf{P}_0	614.93	633.70	624.32	591.81	569.63	580.72
P ₃₀	787.23	794.95	791.09	679.58	657.49	668.53
P ₆₀	839.74	841.95	840.84	775.89	797.13	786.51
P ₉₀	835.47	876.64	856.06	820.69	856.05	838.37
SEm±	10.24	10.30	7.26	21.66	21.09	15.11
CD (P=0.05)	29.10	29.29	20.38	61.58	59.96	42.43
Boron levels						
\mathbf{B}_0	687.89	691.33	689.61	637.35	619.29	628.32
B_1	803.40	821.22	812.31	718.38	743.08	730.73
B_2	816.74	847.89	832.31	795.25	797.85	796.55
SEm±	8.86	8.92	6.29	18.75	18.26	13.09
CD (P=0.05)	25.20	25.37	17.65	53.33	51.93	36.74

Table 4.33.a: Nitrogen uptake in grain and stover of maize as affected by lime, phosphorus and boron



Fig 10: Effect of lime, phosphorus and boron on nitrogen uptake in grain and stover of maize

and K next to soybean in a study conducted by Gupta *et al.* (1989). Increased N uptake with lime on various crops have also been reported by Kanwar and Paliyal (2002); Busari *et al.* (2005); Ranjit *et al.* (2007) and Lynrah and Nongmaithem (2017).

The maximum nitrogen uptake was recorded in P₉₀ while the minimum was recorded in control (P₀) in both grain as well as stover during 2016 and 2017. Pooled data reflected that there was an increase in nitrogen uptake by 37.1% and 44.4% in grain and stover, respectively due to P₉₀ level over control (P₀). The results are in congruence with the findings of Kumar *et al.* (2006); Awomi *et al.* (2012) and Etabo *et al.* (2018). Positive response in nitrogen uptake due to application of phosphorus may be as a result of the role of phosphorus in cell division of shoot and extended growth of meristematic tissues or foliage (Jaetzold *et al.*, 2006 and Etabo *et al.*, 2018).

The results on effect of boron on nitrogen uptake revealed that there was a significant difference among the treatments in case of nitrogen uptake in grain, and stover, where maximum uptake was recorded in treatment B_2 while the minimum uptake was recorded in B_0 during 2016 and 2017. On the basis of pooled data, B_2 level increased nitrogen uptake in grain by 20.7% and in stover by 26.7% over control. It was thus observed that the presence of nutritional element such as boron may have increased the nitrogen uptake in plants. Similar result where the effect of boron on N uptake and metabolism was found to be positive was reported by Inal and Tarakcioglu (2001); Aref (2011) and Sahin (2014).

Interaction effect of lime and phosphorus on nitrogen uptake in grain

From Table 4.33.b, it is apparent that interaction effect of lime and phosphorus on nitrogen uptake in grain was found to be significant in the year 2016 along with pooled data. Highest nitrogen uptake was observed in treatment combination $L_{25}P_{60}$ and minimum was observed in treatment

Table 4.33.b: Interaction effect between lime and phosphorus on nitrogen uptake in grain

	N uptake in grain (mg pot ⁻¹)				
Lime levels	Phosphorus levels				
	Po	P ₃₀	P ₆₀	P ₉₀	
	2016				
L_0	480.35	617.65	664.22	684.16	
L ₂₅	749.52	956.81	1015.26	986.79	
SEm±		14	.48		
CD (P=0.05)		41	.16		

combination L_0P_0 . It could also be observed that at lime level L_0 , N uptake was highest at phosphorus level P_{90} and at lime level L_{25} , it was highest at P_{60} . Thus, lime and phosphorus together had a significant positive impact on nitrogen uptake.

4.2.1.13 Phosphorus uptake in grain and stover

The results on effect of lime revealed that there was a significant effect on the treatments (Table 4.34.a and Fig 11). During 2016 and 2017, the maximum phosphorus uptake in both grain and stover was recorded in L_{25} while the minimum phosphorus uptake in both grain and stover was recorded in L₀. On the basis of pooled data, phosphorus uptake in grain and stover increased by 45.9% and 44.4%, respectively due to liming (L₂₅) over control (L₀). The increase in phosphorus uptake due to liming may be because it breaks the Al and Fe phosphates in soil, thereby making it available to plant. Also, available phosphorus may increase due to increase in mineralization of organic phosphorus as affected by lime (Haynes, 1982). Increased P uptake due to liming in maize has been reported by Busari *et al.* (2005); Singh *et al.* (2009) and Yadesa *et al.* (2019).

The phosphorus uptake by grain and stover was found to be higher in treatments with higher levels of phosphorus as can be seen from table 4.34.a. The maximum phosphorus uptake in grain was recorded in treatment P₉₀ during 2016 as well as 2017, while in stover it was recorded in P₉₀ in 2016 and P₆₀ in 2017. On the basis of pooled data there was an increase in phosphorus uptake by 58.5% and 55.7%, respectively in grain and stover due to application of 90 kg P₂O₅ ha⁻¹ (P₉₀) and 60 kg P₂O₅ ha⁻¹ (P₆₀), respectively over control. Treatments P₆₀ and P₉₀ were found to be at par with each other in case of P uptake in grain and stover. The minimum phosphorus uptake in both grain and stover was observed in the treatment P₀ during 2016 and 2017. Increase in uptake of nutrients was attributed to higher concentration of nutrients as a result of increased availability of nutrients from soil and fertilizer

Treatment	P uptak	e in grain (n	ng pot ⁻¹)	P uptake in stover (mg pot ⁻¹)		
	2016	2017	Pooled	2016	2017	Pooled
Lime levels						
L ₀	230.08	247.77	238.93	123.16	152.73	137.94
L ₂₅	338.52	358.87	348.69	190.53	208.03	199.28
SEm±	5.29	5.86	3.95	8.14	5.95	5.04
CD (P=0.05)	15.04	16.66	11.08	23.16	16.91	14.15
Phosphorus levels						
\mathbf{P}_0	204.73	218.84	211.79	111.25	137.53	124.39
P ₃₀	283.86	316.39	300.13	150.10	177.42	163.76
P ₆₀	317.77	337.42	327.59	181.16	206.15	193.66
P ₉₀	330.84	340.63	335.73	184.87	200.42	192.64
SEm±	7.48	8.29	5.58	11.52	8.41	7.13
CD (P=0.05)	21.27	23.56	15.67	32.75	23.91	20.01
Boron levels						
\mathbf{B}_0	253.34	256.33	254.83	133.99	140.52	137.26
B_1	287.16	318.38	302.77	149.48	199.77	174.62
B_2	312.40	335.24	323.82	187.06	200.85	193.96
SEm±	6.48	7.18	4.83	9.97	7.28	6.17
CD (P=0.05)	18.42	20.41	13.57	28.36	20.70	17.33

Table 4.34.a: Phosphorus uptake in grain and stover of maize as affected by lime, phosphorus and boron



Fig 11: Effect of lime, phosphorus and boron on phosphorus uptake in grain and stover of maize

(Shankarlingappa *et al.*, 2000). Nutrient uptake was recorded maximum with highest dose which was significantly superior to its lower level and control. This was because under these treatments, there was higher biological production which must have increased the nutrient uptake. Affirmative effect of P on phosphorus uptake was observed by Khan *et al.* (2002); Kumar *et al.* (2006); Singh and Singh (2012) and Kumar *et al.* (2015).

Effect of boron on phosphorus uptake in both grain and stover was found to be significant during both the years. Maximum P uptake was recorded in treatment B_2 and minimum was recorded in treatment B_0 in both grain and stover during 2016 as well as 2017. On the basis of pooled data there was an increase in phosphorus uptake by 27.1% and 41.3%, respectively in grain and stover due to application of 2 kg B ha⁻¹ (B₂) over control (B₀). Similar findings have been reported by Aref (2011); Sentimenla *et al.* (2013) and Singh *et al.* (2012), where boron application enhanced the P uptake in plant.

Interaction effect of lime and phosphorus on phosphorus uptake in grain

Interaction effect of lime and phosphorus on phosphorus uptake in grain was significant during 2016 along with pooled data as can be seen from Table 4.34.b. It can be observed that P uptake increased with increase in levels of P application with maximum uptake in P₉₀ level in both lime levels of L₀ (control) and L₂₅ (25% lime of LR). The highest P uptake in grain was observed in treatment combination L₂₅P₉₀ which was at par with L₂₅P₆₀. Treatments with lime application gave higher P uptake compared to treatments without lime application at all levels of phosphorus application. Also, in L₂₅ lime level, treatment P₉₀ was at par with P₆₀. Thus, in treatment L₀ (control), phosphorus level of P₉₀ was found to be the optimum level, whereas in the treatment L₂₅ (25% lime of LR), phosphorus level of P₆₀ was found to be the optimum level for increasing phosphorus uptake in grain.
Table 4.34.b: Interaction effect between lime and phosphorus on phosphorus uptake in grain

	P uptake in grain (mg pot ⁻¹)							
Lime levels	Phosphorus levels							
	Po	P ₃₀	P ₆₀	P ₉₀				
		20	16					
L ₀	171.59	224.03	250.36	274.36				
L ₂₅	237.88	343.69	385.18	387.33				
SEm±		10.	58					
CD (P=0.05)		30.	08					

4.2.1.14 Potassium uptake in grain and stover

It is evident from Table 4.35 and Fig 12 that effect of lime on potassium uptake in grain and stover showed significant difference. The maximum potassium uptake in both grain and stover was recorded in L_{25} while the minimum potassium uptake was recorded in L_0 during 2016 and 2017. Percent increase by 37.7% and 29.5%, respectively in grain and stover was observed in pooled data due to application of L_{25} level of lime over control. Increase in potassium uptake due to liming has been reported in earlier works, where maize recorded maximum uptake of N, P and K next to soybean in a study conducted by Gupta *et al.* (1989). Significant effect of liming on potassium uptake in various crops including maize has been reported by Ranjit *et al.* (2007); Beukes *et al.* (2012); Otieno *et al.* (2018) and Zhihao *et al.* (2019).

Effect of phosphorus on potassium uptake in grain and stover was found to be significant in both the years. It is apparent from table 4.35 that maximum potassium uptake in grain and stover was observed in P₉₀ and minimum uptake in both grain and stover was observed in treatment P₀. Pooled data reflected an increase in potassium uptake in grain by 42.7% and in stover by 33.1% due to application of 90 kg P₂O₅ ha⁻¹ over control. However, potassium uptake in stover in treatment P₉₀ was at par with treatment P₆₀. Increase in potassium uptake due to phosphorus fertilizer application may be explained on the ground that the addition of phosphorus increased the nutrient content of plant by enhancing the root growth and increasing the root hair length, besides increasing the root surface area (Darwesh *et al.*, 2013). Significant increase in potassium uptake due to increasing doses of phosphorus application was also reported by Shankarlingappa *et al.* (2000); Kumar *et al.* (2006) and Kumar *et al.* (2015).

Effect of boron on potassium uptake in both grain and stover was found to be significant in both the years. Maximum potassium uptake in both grain and stover was recorded in the treatment B_2 and minimum was recorded in B_0

	K uptak	te in grain (n	ng pot ⁻¹)	K uptak	K uptake in stover (mg pot ⁻¹)			
Treatment	2016	2017	Pooled	2016	2017	Pooled		
Lime levels								
L_0	329.24	355.39	342.32	1120.73	1105.61	1113.17		
L ₂₅	459.03	483.68	471.36	1459.08	1423.81	1441.45		
SEm±	7.34	8.26	5.53	15.45	13.78	10.35		
CD (P=0.05)	20.88	23.50	15.52	43.94	39.18	29.06		
Phosphorus levels								
\mathbf{P}_0	312.60	329.72	321.16	1100.57	1058.12	1079.34		
P ₃₀	389.78	429.03	409.41	1219.78	1195.08	1207.43		
P ₆₀	430.07	446.93	438.50	1404.56	1366.05	1385.31		
P ₉₀	444.10	472.46	458.28	1434.71	1439.58	1437.15		
SEm±	10.39	11.69	7.82	21.85	19.49	14.64		
CD (P=0.05)	29.53	33.23	21.94	62.14	55.41	41.10		
Boron levels								
\mathbf{B}_0	367.48	386.98	377.23	1221.20	1172.84	1197.02		
B_1	407.00	432.68	419.84	1293.72	1312.76	1303.24		
B_2	407.93	438.95	423.44	1354.79	1308.51	1331.65		
SEm±	9.00	10.12	6.77	18.92	16.88	12.68		
CD (P=0.05)	25.58	28.78	19.00	53.81	47.99	35.59		

Table 4.35: Potassium uptake in grain and stover of maize as affected by lime, phosphorus and boron



Fig 12: Effect of lime, phosphorus and boron on potassium uptake in grain and stover of maize

in both the years. From pooled data, it can be observed that K uptake in grain and stover increased by 12.2% and 11.2%, respectively from treatment B_0 to B_2 . Similar observations where B enhanced potassium uptake in plant has been reported by Günes *et al.* (2011) and Sentimenla *et al.* (2013).

4.2.1.15 Calcium uptake in grain and stover

From Table 4.36 and Fig 13, it can be seen that effect of lime on calcium uptake was significant, where maximum calcium uptake was recorded in treatment L_{25} while minimum uptake was recorded in L_0 during 2016 as well as 2017. According to pooled data, an increase by 98.6% and 97.4% in calcium uptake by grain and stover, respectively could be observed with application of L_{25} over L_0 . Liming might have contributed to Ca uptake by improving the Ca nutrition of crop in addition to neutralizing the Al toxicity. Significant increase in uptake of calcium with increasing doses of lime has been reported by Prasad (1992); Ranjit *et al.* (2007); Singh *et al.* (2009) and Sultana *et al.* (2009).

Effect of phosphorus on calcium uptake in grain and stover was found to be significant in both the years. It is apparent from table 4.36.a that maximum calcium uptake in grain and stover was observed in treatment P₉₀ and minimum in treatment P₀. Pooled data reflected an increase by 60.8% and 50.3% on calcium uptake in grain and stover, respectively due to 90 kg P₂O₅ ha⁻¹ (P₉₀) over control (P₀). In case of Ca uptake in grain treatment P₉₀ was at par with P₆₀. This result is in line with the findings of Kumar *et al.* (2017) and Wafula *et al.* (2018) where they observed an increase in calcium contents with increase in rate of phosphorus.

It is apparent from table 4.36 that effect of phosphorus and boron levels was found to be non-significant in case of Ca uptake in both grain and stover during 2016 as well as 2017.

Turadanant	Ca uptal	ke in grain (r	ng pot ⁻¹)	Ca uptake in stover (mg pot ⁻¹)			
Treatment	2016	2017	Pooled	2016	2017	Pooled	
Lime levels							
L_0	220.70	216.06	218.38	366.79	364.43	365.61	
L ₂₅	440.23	427.27	433.75	723.49	719.83	721.66	
SEm±	13.15	13.61	3.61 9.46		13.86	10.16	
CD (P=0.05)	37.41	38.70	26.57	42.21	39.42	28.51	
Phosphorus levels							
\mathbf{P}_0	232.06	231.86	231.96	419.43	424.03	421.73	
P ₃₀	345.65	322.54	334.09	532.50	534.48	533.49	
P ₆₀	369.50	360.63	365.07	579.40	591.15	585.27	
P ₉₀	374.64	371.63	373.13	649.25	618.87	634.06	
SEm±	18.60	19.25	13.38	21.00	19.60	14.36	
CD (P=0.05)	52.90	54.73	37.57	59.70	55.74	40.32	
Boron levels							
\mathbf{B}_0	324.02	305.97	314.99	523.51	509.49	516.50	
B_1	335.83	324.01	329.92	551.50	562.26	556.88	
B_2	331.54	335.01	333.28	560.42	554.64	557.53	
SEm±	16.11	16.67	11.59	18.18	16.98	12.44	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	

Table 4.36: Calcium uptake in grain and stover of maize as affected by lime, phosphorus and boron



Fig 13: Effect of lime, phosphorus and boron on calcium uptake in grain and stover of maize

4.2.1.16 Boron uptake in grain and stover

The data indicates that boron uptake in grain and stover was found to be significantly affected by the application of lime (Table 4.37.a and Fig 14). The maximum boron uptake in both grain and stover was recorded in L_{25} while minimum boron uptake in both grain and stover was recorded in L_0 in both the years 2016 and 2017. Due to L_{25} level of lime, an increase in boron uptake by 27.0% and 29.3% in grain and stover, respectively was observed from the pooled data. This is parallel with the findings of Singh *et al.* (2009) and Barman *et al.* (2014) who stated a significantly higher uptake of boron by maize crop with application of lime.

B uptake in grain and stover showed significant difference among different levels of phosphorus. It is apparent from the data that maximum B uptake in both grain and stover was recorded with P₉₀ while minimum B uptake in both grain and stover was observed in P₀ during 2016 and 2017. Due to P₉₀ level of phosphorus application, an increase in boron uptake by 23.9% and 22.7% in grain and stover, respectively could be observed from pooled data. In both the years, P₉₀ was found to be at par with P₆₀ in case of B uptake in both grain and stover. Muhlbachova *et al.* (2017) also reported a positive effect of phosphorus fertilization on boron uptake by plants.

It is clear from the data presented in Table 4.37.a that boron application enhanced boron uptake significantly during both the years. The maximum B uptake in both grain and stover was recorded in B_2 , while the minimum uptake was recorded in B_0 in both grain and stover during 2016 and 2017. An increase in boron uptake by 54.2% and 45.9% in grain and stover, respectively due to B_2 level of boron application could be observed from the pooled data. Various workers (Sahin, 2014 and Barman *et al.*, 2014) have also reported a significant increase in plant's boron uptake due to boron fertilization.

Treatment	B uptak	ke in grain (μ	g pot ⁻¹)	B uptake in stover (µg pot ⁻¹)			
	2016	2017	Pooled	2016	2017	Pooled	
Lime levels							
L_0	672.92	682.83	677.87	381.32	388.26	384.79	
L ₂₅	859.82	862.05	860.93	498.46	496.66	497.56	
SEm±	2.84	3.99	3.99 2.45		3.11	2.53	
CD (P=0.05)	8.07	11.34	11.34 6.87		8.84	7.12	
Phosphorus levels							
\mathbf{P}_0	656.42	666.23	661.32	390.00	390.09	390.04	
P ₃₀	779.68	785.76	782.72	425.79	424.28	425.04	
P ₆₀	810.61	817.64	814.12	470.81	470.92	470.87	
P ₉₀	818.77	820.13	819.45	472.95	484.54	478.74	
SEm±	4.01	5.64	3.46	5.67	4.39	3.58	
CD (P=0.05)	11.41	16.04	9.72	16.11	12.49	10.06	
Boron levels							
\mathbf{B}_0	597.10	588.13	592.62	344.57	339.00	341.79	
\mathbf{B}_1	798.03	805.10	801.56	474.77	491.32	483.05	
B_2	903.98	924.09	914.04	500.32	497.05	498.69	
SEm±	3.47	4.89	3.00	4.91	3.81	3.10	
CD (P=0.05)	9.88	13.89	8.41	13.95	10.82	8.72	

Table 4.37.a: Boron uptake in grain and stover of maize as affected by lime, phosphorus and boron



Fig 14: Effect of lime, phosphorus and boron on boron uptake in grain and stover of maize

Interaction effect of lime and phosphorus on boron uptake in grain

It is evident from Table 4.37.b that boron uptake in grain due to effect of lime and phosphorus was found to be significant. In case of treatments with lime, phosphorus level up to 60 kg ha⁻¹ showed maximum boron uptake, while in case of treatments without lime, phosphorus level up to 90 kg ha⁻¹ showed maximum boron uptake. Irrespective of liming, it can also be observed that the boron uptake in grain increased with increase in phosphorus levels and vice versa. Highest uptake was observed in the treatment combination $L_{25}P_{60}$ and lowest uptake in L_0P_0 during 2016 as well as 2017.

Interaction effect of phosphorus and boron on boron uptake in grain

From Table 4.37.c, it can be observed that interaction effect of phosphorus and boron on boron uptake in grain was significant during 2016 along with pooled data. The boron uptake in grain was observed to increase linearly with increase in phosphorus levels as well as boron levels, the maximum of which was observed in the treatment combination $P_{90}B_2$ and minimum was observed in P_0B_0 . Significant increase in B uptake with increase in phosphorus and boron levels has been reported by Sentimenla *et al.* (2013) in soybean.

Interaction effect of lime and boron on boron uptake in grain and stover

As observed from Table 4.37.d, interaction effect of lime and boron on boron uptake in grain and stover was significant during 2016 and 2017. It can be observed that B uptake in grain and stover increased with lime application and also increased with increasing levels of boron in both the years of experimentation. Maximum boron uptake in both grain and stover was observed in treatment combination $L_{25}B_2$ and minimum was observed in treatment combination $L_{0}B_0$ during 2016 as well as 2017.

grain								
	B uptake in grain (µg pot ⁻¹)							
Lime levels	Phosphorus levels							
	P ₀	P ₃₀	P ₆₀	P ₁₀₀				
	2016							
L_0	568.04	677.11	704.33	742.20				

882.25

693.69

877.84

685.40

880.05

5.67

16.13

2017

7.98

22.69

Pooled

4.89

13.74

916.89

718.16

917.13

711.24

917.01

895.35

744.05

896.21

743.12

895.78

744.79

575.43

757.03

571.73

750.91

L₂₅

SEm±

CD (P=0.05)

 L_0

L25

SEm±

CD (P=0.05)

 L_0

L25

 $SEm \pm$

CD (P=0.05)

Table 4.37.b: Interaction effect between lime and phosphorus on boron uptake in grain

	B uptake in grain (μ g pot ⁻¹)									
Phosphorus levels		Boron levels								
	B1	B ₂	B ₃							
		2016								
\mathbf{P}_0	503.45	686.56	779.24							
P ₃₀	612.09	813.89	913.07							
P ₆₀	631.56	842.10	958.17							
P_{100}	641.31	849.56	965.45							
SEm±		6.95								
CD (P=0.05)		19.76								

Table 4.37.c: Interaction effect between phosphorus and boron on boron uptake in grain

	B uptake in grain (µg pot ⁻¹)									
Lime levels		Boron levels	5		Boron levels	5		Boron levels		
	B ₀	B_1	B ₂	B ₀	B_1	B ₂	B ₀	B_1	B ₂	
		2016			2017			Pooled		
L_0	517.93	701.38	799.45	514.41	711.80	822.27	516.17	706.59	810.86	
L ₂₅	676.27	894.67	1008.51	661.86	898.39	1025.91	669.06	896.53	1017.21	
SEm±		4.91			6.91			4.24		
CD (P=0.05)		13.97			19.65			11.90		
				B uptak	e in stover (µ	ug pot ⁻¹)				
		2016			2017			Pooled		
L ₀	298.44	410.24	435.27	297.49	431.01	436.28	297.96	420.63	435.77	
L ₂₅	390.71	539.29	565.38	380.51	551.64	557.82	385.61	545.46	561.60	
SEm±		6.94			5.38			4.39		
CD (P=0.05)		19.73			15.30			12.33		

Table 4.37.d: Interaction effect between lime and boron on boron uptake in grain and stover

4.2.2 Effect of lime, phosphorus and boron on soil properties4.2.2.1 Effect on soil pH and electrical conductivity

The results on soil pH and electrical conductivity after crop harvest in different treatments have been presented in Table 4.38. It is apparent from the data that lime had a significant effect on soil pH, whereas it had no significant effect on electrical conductivity. The maximum pH was recorded in the treatment L_{25} , while the minimum was recorded in the treatment L_0 during 2016 and 2017. Pooled data reflected an increase by 8.8% in the soil pH due to liming over control (L_0). Lime increased soil pH because of the release of Ca²⁺ ions which displaces Al³⁺, H⁺ and Fe³⁺ ions in the soil (Kisinyo *et al.*, 2013).

The treatment effect of phosphorus as well as boron on soil pH and electrical conductivity after crop harvest was found to be non- significant as can be observed from Table 4.38.

4.2.2.2 Effect on percent base saturation and organic carbon

From Table 4.39 it is apparent that effect of lime on soil organic carbon in post harvest soil was non-significant, whereas significant effect of lime application on percent base saturation of post harvest soil was observed during both the years. Maximum percent base saturation was recorded in treatment L_{25} and minimum was recorded in L_0 . A critical examination of data indicated that L_{25} level of lime increased percent base saturation from 26.32 to 31.30%. Increase in percent base saturation due to liming is because liming releases calcium into the soil solution which replaces a large part of the acidic cations in the exchange complex, thereby increasing the percent base saturation.

The results on effect of phosphorus levels and boron levels on percent base saturation and organic carbon were found to be non-significant (Table 4.39).

Traatmont		Soil pH		Electrical conductivity (dSm ⁻¹)			
Treatment	2016	2017	Pooled	2016	2017	Pooled	
Lime levels							
L_0	5.22	5.20	5.21	0.20	0.19	0.20	
L ₂₅	5.65	5.69	5.67	0.18	0.18	0.18	
SEm±	0.02	0.02	0.02	0.01	0.01	0.01	
CD (P=0.05)	0.07	0.06	0.04	NS	NS	NS	
Phosphorus levels							
\mathbf{P}_0	5.40	5.41	5.41	0.21	0.19	0.20	
P ₃₀	5.49	5.48	5.49	0.19	0.18	0.19	
P ₆₀	5.44	5.45	5.45	0.19	0.19	0.19	
P ₉₀	5.41	5.43	5.42	0.18	0.18	0.18	
SEm±	0.03	0.03	0.02	0.01	0.01	0.01	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	
Boron levels							
\mathbf{B}_0	5.44	5.44	5.44	0.20	0.19	0.19	
B_1	5.43	5.44	5.43	0.19	0.18	0.19	
B_2	5.44	5.45	5.44	0.19	0.19	0.19	
SEm±	0.03	0.03	0.02	0.01	0.01	0.01	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	

Table 4.38: Soil pH and electrical conductivity of post-harvest soil as affected by lime, phosphorus and boron

Turaturant	Percent	base satura	tion (%)	Organic carbon (%)			
I reatment	2016	2017	Pooled	2016	2017	Pooled	
Lime levels							
L ₀	26.00	26.65	26.32	1.56	1.58	1.57	
L ₂₅	30.95	31.64	31.30	1.57	1.59	1.58	
SEm±	0.30	0.30	0.21	0.02	0.02	0.01	
CD (P=0.05)	0.84	0.86	0.59	NS	NS	NS	
Phosphorus levels							
\mathbf{P}_0	28.56	29.29	28.93	1.55	1.57	1.56	
P ₃₀	28.66	29.33	28.99	1.57	1.59	1.58	
P ₆₀	28.46	29.10	28.78	1.58	1.60	1.59	
P ₉₀	28.21	28.87	28.54	1.57	1.59	1.58	
SEm±	0.42	0.43	0.30	0.02	0.02	0.02	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	
Boron levels							
\mathbf{B}_0	28.45	29.12	28.79	1.55	1.58	1.57	
B1	28.50	29.18	28.84	1.57	1.58	1.58	
B ₂	28.46	29.14	28.80	1.58	1.59	1.58	
SEm±	0.36	0.37	0.26	0.02	0.02	0.01	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	

Table 4.39: Percent base saturation and soil organic carbon of post-harvest soil as affected by lime, phosphorus and boron

4.2.2.3 Effect on available nitrogen, phosphorus and potassium

It is apparent from the data (Table 4.40) that effect of lime on available nitrogen, phosphorus and potassium was significant during both the years of experimentation. Maximum available nitrogen, phosphorus and potassium in soil were recorded in L_{25} while the minimum was recorded in L_0 during 2016 and 2017. The amount of these nutrients were found to be more in the limed treatment because of the mineralization of organic matter, a process which is known to be enhanced by liming and whereby inorganic nutrients such as N and P are released into the soil solution (Andersson and Nilsson (2001). In case of phosphorus, it may also be due to release of phosphate from Fe and Al complex. The increase in potassium availability may be due to the displacement of exchangeable K by Ca as reported by Haynes and Naidu (1998). Significant increase in potassium content in soil due to liming has also been reported by Barman *et al.* (2014) and Kamaruzzaman *et al.* (2014).

The effect of phosphorus on phosphorus content in soil at harvest was found to be significant, while its effect on available nitrogen and potassium was found to be non-significant. Soil available phosphorus was found to be higher where higher rate of phosphorus was applied. It is apparent from Table 4.40.a that maximum available phosphorus was recorded in P₉₀, while the minimum was recorded in treatment P₀ during both the years. However, treatment P₉₀ was found to be at par with P₆₀. Thus, increasing levels of phosphorus significantly increased phosphorus content in soil which must be primarily due to the release of P from fertilizer. Similar results have also been reported on P deficient acid soils (Weisz *et al.*, 2003; Kisinyo *et al.*, 2014).

The data presented in table 4.40.a revealed that there was no significant difference among the treatments with respect to boron application on soil available nitrogen, phosphorus and potassium which is in congruence to the findings of Barman *et al.* (2014).

161

Turaturat	Available nitrogen (kg ha ⁻¹)			Availab	Available phosphorus (kg ha ⁻¹)			Available potassium (kg ha ⁻¹)		
Ireatment	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled	
Lime levels										
L_0	238.65	239.85	239.25	10.61	10.40	10.51	149.85	150.00	149.92	
L ₂₅	248.73	248.99	248.86	12.23	12.46	12.35	150.95	151.60	151.27	
SEm±	0.31	0.24	0.19	0.15	0.19	0.12	0.24	0.28	0.18	
CD (P=0.05)	0.88	0.68	0.55	0.41	0.54	0.34	0.68	0.78	0.51	
Phosphorus levels										
\mathbf{P}_0	242.97	243.81	243.39	10.07	10.00	10.04	150.67	150.73	150.70	
P ₃₀	243.51	244.37	243.94	11.19	11.21	11.20	150.41	150.63	150.52	
P ₆₀	244.12	244.54	244.33	12.12	12.17	12.15	150.33	150.57	150.45	
P90	244.15	244.97	244.56	12.30	12.36	12.33	150.19	151.26	150.72	
SEm±	0.44	0.34	0.28	0.21	0.27	0.17	0.34	0.39	0.26	
CD (P=0.05)	NS	NS	NS	0.58	0.77	0.48	NS	NS	NS	
Boron levels										
\mathbf{B}_0	243.34	243.97	243.65	11.36	11.28	11.32	150.48	150.83	150.65	
B_1	243.89	244.65	244.27	11.41	11.47	11.44	150.16	150.54	150.35	
B_2	243.84	244.65	244.25	11.49	11.55	11.52	150.57	151.03	150.80	
SEm±	0.38	0.29	0.24	0.18	0.23	0.15	0.29	0.34	0.22	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS	

Table 4.40: Available nitrogen, phosphorus and potassium of post-harvest soil as affected by lime, phosphorus and boron

4.2.2.4 Effect on exchangeable calcium and available boron

It is evident from table 4.41 that liming had a significant effect on available boron and exchangeable calcium content in soil at harvest where maximum was recorded in L_{25} and minimum was recorded in L_0 during 2016 and 2017. Liming supplies Ca to the soil and also increases the CEC by raising the soil pH. This condition makes it ideal for Ca to freely occupy the exchange complex and hence its availability increases. Similar findings have been reported by Rahman *et al.* (2005); Beukes *et al.* (2012) and Han *et al.* (2019). Increase in boron availability due to liming may be due to neutralization of soil acidity which may have released boron into the soil solution (Sarkar *et al.*, 2015).

Effect of phosphorus on exchangeable calcium content in soil was significant. It can be observed that exchangeable calcium increased with increase in P levels and maximum value was recorded in treatments P_{60} and P_{90} , while minimum was recorded in treatment P_0 . Similar findings where P application increased Ca content in soil have been reported by Venkatesh *et al.* (2002). Result on the effect of boron on exchangeable calcium revealed that there was no significant difference among the treatments.

From Table 4.41 it can be observed that there was no significant effect of boron on exchangeable calcium, whereas there was significant effect of boron on available boron. The application of boron @ 2 kg ha⁻¹ (B₂) reflected the highest value while minimum soil available boron was observed in the control treatment *i.e.*, B₀ during 2016 and 2017. The increase in extractable B in soil with increasing rate of B application has been reported by Barman *et al.* (2014).

Treatment	Exc [c	zhangeable (zmol (p+) kg	Ca^{2+}	Available boron (mg kg ⁻¹)			
Ireatment	2016	2017	Pooled	2016	2017	Pooled	
Lime levels							
L ₀	3.04	3.06	3.05	0.51	0.54	0.52	
L ₂₅	4.23	4.27	4.25	0.56	0.58	0.57	
SEm±	0.01	0.01	0.01	0.02	0.01	0.01	
CD (P=0.05)	0.04	0.04	0.03	0.04	0.04	0.03	
Phosphorus levels							
\mathbf{P}_0	2.89	2.93	2.91	0.54	0.55	0.54	
P ₃₀	3.80	3.83	3.82	0.52	0.57	0.54	
P ₆₀	3.92	3.94	3.93	0.53	0.56	0.55	
P ₉₀	3.92	3.95	3.93	0.54	0.57	0.55	
SEm±	0.02	0.02	0.01	0.02	0.02	0.01	
CD (P=0.05)	0.06	0.05	0.04	NS	NS	NS	
Boron levels							
\mathbf{B}_0	3.64	3.66	3.65	0.47	0.52	0.49	
B_1	3.64	3.67	3.65	0.54	0.56	0.55	
B ₂	3.63	3.66	3.64	0.59	0.61	0.60	
SEm±	0.02	0.02	0.01	0.02	0.02	0.01	
CD (P=0.05)	NS	NS	NS	0.05	0.05	0.04	

Table 4.41: Exchangeable calcium and available boron of post-harvest soil as affected by lime, phosphorus and boron

4.2.2.5 Effect on exchangeable hydrogen, aluminium and total potential acidity

The data on exchangeable hydrogen, aluminium and total potential acidity in soil at harvest (Table 4.42) showed a decreasing trend with application of lime. Thus, maximum exchangeable hydrogen, aluminium and total potential acidity in post harvest soil were recorded in L_0 and minimum in L_{25} during 2016 and 2017. From pooled data, a percent decrease in exchangeable hydrogen, aluminium and total potential acidity by 37.9%, 8.3% and 11.3%, respectively was observed with application of L_{25} level. A decrease in all forms of soil acidity due to liming has also been reported by Badole *et al.* (2015). Chatterjee *et al.* (2005) also reported that addition of lime shows significant decrease in exchangeable H, Al and total potential acidity.

There was no significant effect of phosphorus and boron on exchangeable hydrogen, aluminium and total potential acidity in soil at harvest as can be observed in Table 4.42.

4.3. Agronomic efficiency and nutrient use efficiency

4.3.1 Agronomic efficiency

From Table 4.43 it can be observed that agronomic efficiency (AE) of P decreased with increase in the dose of phosphorus. Agronomic efficiency of P was highest at 30 kg P_2O_5 ha⁻¹ (P_{30}), which declined with further increase in P levels. Minimum was observed in the highest level at 90 kg P_2O_5 ha⁻¹ (P_{90}). Similar findings where AE of P decreased with increasing levels of phosphorus doses have been reported by Kumar *et al.* (2006) in maize and Sentimenla *et al.* (2013) in soybean. Agronomic efficiency of applied boron was highest at 1 kg B ha⁻¹ (B₁) and minimum at 2 kg B ha⁻¹ B₂. This is also in line with the findings of Khurana and Arora (2012) who recorded a decrease in AE of B applied through borax with increase in B levels. Higher AE values reflect the optimum nutrient management practice (Chuan *et al.*, 2013).

Treatments	Exchangeable H ⁺ [cmol (p ⁺) kg ⁻¹]			Ex	Exchangeable Al^{3+} [cmol (p ⁺) kg ⁻¹]			Total potential acidity [cmol (p ⁺) kg ⁻¹]		
	2016	2017	Pooled	2016	2017	Pooled	2016	2017	Pooled	
Lime levels										
L_0	0.99	0.92	0.95	1.32	1.34	1.33	14.90	14.83	14.87	
L ₂₅	0.55	0.64	0.59	1.22	1.21	1.22	13.20	13.17	13.19	
SEm±	0.01	0.01	0.01	0.01	0.01	0.01	0.02	0.01	0.01	
CD (P=0.05)	0.03	0.02	0.02	0.03	0.02	0.02	0.05	0.03	0.03	
Phosphorus levels										
\mathbf{P}_0	0.78	0.79	0.78	1.27	1.27	1.27	14.07	14.02	14.05	
P ₃₀	0.75	0.76	0.76	1.28	1.29	1.28	14.04	14.00	14.02	
P ₆₀	0.78	0.79	0.78	1.26	1.28	1.27	14.06	13.99	14.03	
P ₉₀	0.76	0.78	0.77	1.27	1.28	1.27	14.04	13.98	14.01	
SEm±	0.02	0.01	0.01	0.01	0.01	0.01	0.02	0.02	0.01	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS	
Boron levels										
\mathbf{B}_0	0.77	0.78	0.78	1.26	1.27	1.27	14.04	13.99	14.02	
B_1	0.77	0.78	0.78	1.27	1.28	1.28	14.05	14.00	14.03	
B_2	0.77	0.77	0.77	1.28	1.29	1.28	14.07	14.01	14.04	
SEm±	0.01	0.01	0.01	0.01	0.01	0.01	0.02	0.01	0.01	
CD (P=0.05)	NS	NS	NS	NS	NS	NS	NS	NS	NS	

Table 4.42: Exchangeable H⁺, Al³⁺ and total potential acidity of post-harvest soil as affected by lime, phosphorus and boron

4.3.2 Nutrient use efficiency

As shown in Table 4.44, during 2016 and 2017, the phosphorus use efficiency (PUE) by maize was observed to decrease with increase in level of phosphorus. This decreasing trend is in accordance with von Leibig's Law of the Minimum which states that the most limiting factor determines the yield potentials (Giller *et al.*, 2002). Lower the P application rate better was the use efficiency. Higher PUE among treatments was obtained at 30 kg P_2O_5 ha⁻¹ (P_{30}), thereafter it significantly reduced, where lowest PUE was recorded at 90 kg P_2O_5 ha⁻¹ (P_{90}). PUE decreases at higher doses of P due to the fact that plants grown in P deficient soil exhibit greater P sorption at lower doses of P (Dubey, 2000). Similar findings have also been reported by Hussein (2009); Nekesa (2007) and Kisinyo *et al.* (2014).

From Table 4.44, the boron use efficiency (BUE) by maize was maximum at 1 kg B ha⁻¹ (B₁) and minimum at 2 kg B ha⁻¹ (B₂) during both the years. Thus, the BUE decreased with increment of B levels applied which can be attributed to loss of boron. Boron may encounter greater losses and less utilization by the crops when applied in excess, since plant nutritional demand is limited. The excess of applied B may not be absorbed by plant and can be lost decreasing the efficiency of fertilization with higher B rates. This result is in accordance with the findings of Byju *et al.* (2007) where it was found that application of B at 1.0 kg ha⁻¹ gave the highest BUE. The efficiency of boron fertilization was low when compared to phosphorus. It is mainly due to the complex dynamics of this nutrient with low mobility within the phloem (Mantovani *et al.*, 2013), but with high mobility in soil, high solubility in water and low reactivity with the soil making it susceptible to leaching (Trautmann *et al.*, 2014).

Treatment	Agronomic efficiency (g g ⁻¹)			
	2016	2017	Pooled	
P ₀	-	-	-	
P ₃₀	56.07	54.43	55.22	
P ₆₀	35.02	34.25	34.63	
P90	24.81	23.47	24.13	
B_0	-	-	-	
B1	746.27	1004.48	874.63	
B ₂	430.60	635.07	532.84	

Table 4.43: Agronomic efficiency of phosphorus and boron

Table 4.44: Nutrient use efficiency of phosphorus and boron

Treatment	Phosphorus use efficiency (%)			
	2016	2017	Pooled	
Po	-	_	-	
P ₃₀	39.37	48.53	43.95	
P ₆₀	28.12	29.50	28.81	
P ₉₀	20.91	20.20	20.55	
	Boron use efficiency (%)			
B_0	-	_	-	
B_1	3.00	3.24	3.12	
B_2	2.29	2.51	2.40	

CHAPTER V

SUMMARY AND CONCLUSION

SUMMARY AND CONCLUSION

Two pot experiments were conducted in the greenhouse, Department of Agricultural Chemistry and Soil Science, School of Agricultural Sciences and Rural Development (SASRD), Nagaland University, Medziphema, Nagaland during the *kharif* season of 2016 and 2017 to carry out the investigation entitled, "Study on Acidity Tolerance of Maize and its Response to Phosphorus and Boron". The main findings of the investigations are summarized below:

EFFECT OF SOIL pH, LIME AND VARIETIES ON THE PERFORMANCE OF MAIZE AND SOIL PROPERTIES (EXPT-I)

- Maximum plant height at all growth stages was recorded in pH₂ (pH 5.2) over pH₁ (pH 4.6). Effect of liming on plant height was recorded maximum at 100% lime of LR (L₁₀₀) at 30, 60 DAS and at harvest. At harvest, application of 100% lime enhanced the pooled plant height by 22% over control. Treatments L₁₀₀ and L₆₀ were found to be at par with each other at all growth stages. At harvest, superiority of varieties in terms of plant height was recorded in the order RCM-76 > RCM-1-1 > RCM-75. Significant interaction effect of pH and lime on plant height at 30 DAS was observed. Higher soil pH level along with higher lime levels yielded in higher plant height. Maximum plant height was recorded in treatment combination pH₂L₆₀ and minimum was recorded in treatment combination pH₁L₀.
- 2. The effect of pH on number of leaves was non-significant. The effect of lime on the number of leaves was significant and at 30 and 60 DAS, highest value was recorded at 100% lime of LR (L₁₀₀) and at harvest highest value was recorded at 60% lime of LR (L₆₀). Lowest value was observed in control (L₀) at all growth stages. Treatments L₆₀ and L₁₀₀ were at par at all growth stages. Varietal effect was also significant and

maximum number of leaves was recorded in the variety RCM-76 (V_2) and minimum in the variety RCM-75 (V_1) at all growth stages.

- **3.** The effect of pH on leaf area index was non-significant at all growth stages. Significant effect of lime on leaf area index was observed at 60 DAS and at harvest, where maximum leaf area index was recorded at 60% lime of LR (L₆₀) and minimum leaf area index was recorded in control (L₀) in both crop stages. At 60 DAS, treatment L₆₀ was at par with L₁₀₀ and at harvest, treatment L₆₀ was at par with L₃₀ and L₁₀₀. Varieties did not have any significant effect on the leaf area index at any growth stages.
- 4. Effect of pH on cob length, cob girth and cob weight was significant and maximum value was recorded in pH 5.2 (pH₂) and minimum in pH 4.6 (pH₁). Effect of lime was significant and maximum cob length was recorded at 60% lime of LR (L₆₀), whereas, maximum cob girth and cob weight was recorded at 100% lime of LR (L₁₀₀). Minimum cob length, cob girth and cob weight was observed in control (L₀). In case of cob length and cob weight, treatment L₁₀₀ was at par with L₆₀ and in case of cob girth, treatment L₁₀₀ was at par with L₃₀ and L₆₀. Varieties also showed significant response and maximum cob length, cob girth and cob weight was recorded in variety RCM-76 (V₂), whereas, minimum cob length and cob weight was recorded in RCM-1-1 (V₃) and minimum cob girth was recorded in RCM-75 (V₁).
- 5. Effect of pH was significant on the number of grains per row and number of grains per cob, where highest value was recorded in pH 5.2 (pH₂) and minimum was recorded in pH 4.6 (pH₁). Effect of lime was significant on number of grains per row, number of rows per cob and number of grains per cob where maximum number of grains per row was recorded in L₆₀, while number of rows per cob and number of grains per cob was recorded in L₁₀₀. L₆₀ enhanced pooled number of

grains per row to the extent of 8.0% over control, while L_{100} enhanced number of rows per cob and grains per cob to the extent of 9.0 and 27.0%, respectively over control. For number of grains per row and number of rows per cob, treatments L_{60} and L_{100} were found to be at par with each other. Effect of varieties was significant and variety RCM-76 (V_2) recorded maximum number of grains per row, number of rows per cob and number of grains per cob followed by RCM-75 (V1) and RCM-1-1 (V₃). The RCM-76 variety increased pooled grains per row by 2.0% and 5.3%, rows per cob by 1.5 and 4.2% and grains per cob by 4.1 and 10.3% over RCM-75 and RCM-1-1, respectively. Interaction effect of pH and lime on number of rows per cob was significant, where maximum rows per cob were recorded in treatment combination pH₁L₁₀₀. However, treatment combination pH₁L₁₀₀ was at par with pH₂L₆₀. Interaction effect of pH and varieties on number of grains per row was found to be significant during 2016, where highest value was obtained in pH 5.2 with variety RCM-76 (pH₂V₂) and lowest value in pH 4.6 with variety RCM-75 (pH_1V_1).

6. Effect of pH on test weight, grain yield and stover yield of maize was significant, where highest response was observed in pH 5.2 (pH₂) and lowest in pH 4.6 (pH₁). Grain and stover yield increased by 13.3% and 9.6%, respectively in pH 5.2 (pH₂) soil as compared to pH 4.6 (pH₁) soil. Effect of lime was also significant and maximum test weight, grain yield and stover yield was observed @ 100% lime of LR (L₁₀₀) and minimum in control treatment (L₀). Grain yield increased by 28.7% and stover yield by 23.7% in treatment L₁₀₀ over L₀. However, L₁₀₀ was at par with L₆₀ for all the three parameters. Significant response of varieties was also observed and variety RCM-76 (V₂) gave the highest grain as well as stover yield, while RCM-1-1 (V₃) gave the lowest grain

and stover yield. RCM-76 (V₂) gave 3.0% and 9.0% higher grain yield over RCM-75 (V₁) and RCM-1-1 (V₃), respectively

- 7. Effect of pH on N content in grain as well as stover was non-significant in both the years. Effect of lime on N content in grain and stover was significant with highest value in the treatment L₁₀₀ and lowest in L₀. N content increased by 27.4% and 23.0%, respectively in grain and stover in L₁₀₀ over L₀. Effect of varieties on N content in grain and stover was non-significant.
- 8. Effect of pH on P content in grain and stover was significant and highest value was observed in pH 5.2 (pH₂). P content in grain increased by 14.7% and in stover by 23.07% in pH₂ over pH₁. Effect of lime on P content in grain and stover was significant where maximum was recorded in treatment L_{100} in grain and in L_{60} in stover. Minimum P content for both grain and stover was observed in L₀. P content in both grain and stover was found to increase by 33.3% each in treatment L_{100} in grain and in L_{60} in stover over L_0 . In both P content in grain and stover, treatments L_{60} and L_{100} were at par with each other. Significant effect of varieties on P content was observed only in grain, where variety RCM-76 showed highest P content followed by RCM-75 and RCM-1-1. Interaction effect of pH and lime on P content in grain was significant and maximum P content was recorded in treatment combination pH₂L₆₀ and minimum P content in treatment combination pH_1L_0 . Treatment combination pH_2L_{60} was at par with pH_1L_{100} , pH_2L_{30} , and pH_2L_{100} .
- **9.** Effect of pH on K content in grain and stover was significant and recorded highest in pH 5.2 (pH₂) and lowest in pH 4.6 (pH₁). K content in grain and stover increased by 14.9% and 3.5%, respectively in pH₂ over pH₁. Effect of lime on K content in grain and stover was significant and recorded highest in treatment L₁₀₀ and lowest in L₀. L₁₀₀ level of

lime enhanced K content in grain and stover which increased by 21.7% and 14.2%, respectively over L₀.In case of K content in stover, L₁₀₀ was at par with L₆₀. Effect of varieties on K content was found to be significant only in grain. Maximum K content in grain was reported in the variety RCM-76 (V₂) and minimum in variety RCM-75 (V₁).

- **10.** Effect of pH on Ca content in grain and stover was significant and highest value was recorded in pH 5.2 (pH₂) and lowest in pH 4.6 (pH₁). A percent increase of 18.6% in grain and 15.2% in stover was observed in pH₂ over pH₁. Lime had a significant effect on Ca content in grain and stover, where treatment L_{100} recorded highest Ca content and L_0 recorded lowest Ca content in both grain and stover. A percent increase of 96.4% and 87.1% in grain and stover, respectively in L_{100} over L_0 was observed. However, treatment L_{100} was at par with L_{60} in case of calcium content in both grain and stover. Effect of varieties on Ca content was non-significant.
- 11. Effect of pH was non-significant on B content in grain. It was significant on B content in stover and highest value was recorded in pH 5.2 (pH₂) and lowest in pH 4.6 (pH₁). Effect of lime was significant and maximum B content in both grain and stover was recorded in L₃₀ which was at par with L₆₀. An increase in grain and boron content in grain and stover by 2.3% and 6.0%, respectively at L₃₀ over L₀ was observed. Minimum B content in grain and stover was recorded in L₀. The effect of varieties on B content in grain and stover was non-significant.
- 12. Effect of pH on N uptake in grain and stover was significant and highest value was recorded in pH 5.2 (pH₂) and minimum value in pH 4.6 (pH₁). pH₂ enhanced nitrogen uptake in grain by 14.8% and in stover by 12.5% over pH₁. Lime effect was significant and maximum N uptake in both grain and stover was recorded at 100% lime of LR (L₁₀₀), and minimum in control (L₀). Treatment L₁₀₀ enhanced nitrogen uptake to

the extent of 64.3% and 50.6% in grain and stover, respectively over control. Among the varieties, significant difference was also observed, where maximum N uptake in grain and stover was observed in variety RCM-76 (V₂), and minimum in RCM-1-1 (V₃). N uptake in V₂ was 4.2% and 13.0% higher than V₁ and V₃, respectively. Variety RCM-75 (V₁) was at par with variety RCM-76 (V₂) with respect to N uptake in stover.

- 13. Effect of pH on P uptake in grain and stover was significant and recorded highest value in pH 5.2 (pH₂) over pH 4.6 (pH₁). P uptake increased by 29.1% in grain and by 31.8% in stover in pH₂ compared to pH₁. Effect of lime was significant on P uptake in grain and stover and highest value was recorded @ 100% lime of LR (L100), and minimum in control (L_0). An increase by 72.6% and 64.5% in grain and stover, respectively in L_{100} as compared to L_0 was observed. Treatment L_{100} was found to be at par with treatment L_{60} in case of P uptake in both grain and stover. Variety exhibited significant effect on P uptake in grain and non-significant effect on stover. In grain, variety RCM-76 (V₂) exhibited highest P uptake followed by RCM-75 (V1) and RCM-1-1 (V₃). P uptake in grain of RCM-76 (V₂) variety was 9.1% and 20.6% higher as compared to V_1 and V_2 varieties, respectively. Interaction effect of pH and lime on P uptake in grain was significant during 2017, where maximum P uptake in grain was observed in pH 5.2 at 60% lime of LR (pH_2L_{60}) and minimum in pH 4.6 at 0% lime of LR (pH_1L_0). It was observed that irrespective of pH levels, lime enhanced P uptake in grain.
- 14. Significant effect of pH on K uptake in grain and stover was recorded highest in pH 5.2 (pH₂) and minimum in pH 4.6 (pH₁). There was an increase by 29.9% and 13.3% in potassium uptake in grain and stover, respectively in pH₂ over pH₁. Effect of lime was significant, where

maximum K uptake in grain and stover was recorded @ 100% lime of LR (L_{100}). Minimum K uptake in both grain and stover was recorded in control treatment (L_0). There was an increase in K uptake in grain by 54.7% and in stover by 41.0% in L_{100} over control. Variety showed significant effect, where maximum K uptake in grain and stover was recorded in variety RCM-76 (V_1) and minimum was recorded in variety RCM-1-1 (V_3). K uptake in grain of V_2 was 15.3% and 16.1% higher over V_1 and V_3 , respectively.

- **15.** Effect of pH was significant on Ca uptake in grain and stover, where highest value was recorded in pH 5.2 (pH₂) and lowest value in pH 4.6 (pH₁). Ca uptake in grain and stover increased by 34.7% and 25.3%, respectively in pH₂ over pH₁. Effect of lime on Ca uptake in grain and stover was significant which recorded highest value @ 100% lime of LR (L₁₀₀) and lowest value in control treatment (L₀). An increase by 149.9% in grain and 131.9% in stover, respectively at 100% lime of LR over control was observed. In case of Ca uptake in grain, treatment L₁₀₀ was at par with L₆₀. Effect of varieties was significant on Ca uptake in grain and non-significant on Ca uptake in stover. Maximum Ca uptake in grain was observed in variety RCM-76 (V₂) followed by RCM-75 (V₁) and RCM-1-1 (V₃).
- 16. Significant effect of pH on B uptake in grain and stover was observed and maximum value was recorded in pH 5.2 (pH₂) and minimum value in pH 4.6 (pH_{4.6}). B uptake in grain increased by 13.7% and in stover by 10.8% in pH 5.2 over pH 4.6. Effect of lime was significant on B uptake in grain and stover. Maximum value of B uptake in both grain and stover was recorded @ 60% lime of LR (L₆₀) and minimum in control treatment (L₀). The L₆₀ level of lime increased boron uptake in grain and stover to the extent of 31.2% and 28.1% over control. Treatment L₆₀ was at par with L₁₀₀ in case of B uptake in grain, whereas treatment L₆₀

was at par with L_{30} , and L_{100} in case of B uptake in stover. Variety had a significant effect where maximum B uptake in grain and stover was recorded in RCM-76 (V₂) and minimum was recorded in RCM-1-1 (V₃). B uptake in grains of V₂ variety was 2.8 and 9.2% higher than V₁ and V₃ varieties, respectively. In case of B uptake in stover, RCM-75 (V₁) and RCM-76 (V₂) were found to be at par with each other.

- 17. Significant effect of pH on soil pH of post harvest soil was observed where highest pH value was recorded in pH 5.2 (pH₂) over pH 4.6 (pH₁). Lime had a significant effect on soil pH and maximum pH value was recorded @ 100% lime of LR (L₁₀₀), while, minimum pH value was observed in control (L₀). On the other hand, effect of pH and lime on electrical conductivity of post harvest soil was non-significant. Effect of varieties was non-significant on both soil pH and electrical conductivity. Interaction effect of pH and lime on soil pH was significant, where maximum soil pH value was recorded in pH₂L₁₀₀ which was at par with pH₁L₁₀₀, and minimum was recorded in pH₁L₀ treatment combination.
- 18. Effect of pH was non-significant on base saturation and organic carbon content in post harvest soil. Effect of lime was non-significant on soil organic carbon content and significant on base saturation of post harvest soil. Highest base saturation value was observed in treatment of 100% lime of LR (L₁₀₀) and lowest in control treatment (L₀). No significant effect of varieties on percent base saturation and organic carbon in post harvest soil was observed. Interaction effect of pH and lime on percent base saturation was significant during 2017, where maximum value was recorded in pH₂L₆₀. It was observed that increase in lime levels led to an increase in the percent base saturation under both the soil pH.
- 19. Effect of pH on available N and K of post harvest soil was non-significant, whereas it was significant on P content of post harvest soil. Maximum available P content in post harvest soil was recorded in pH₂

and minimum was recorded in pH₁. The pH₂ level enhanced available phosphorus by 10.6% over pH₁. Effect of liming on NPK content of post harvest soil was significant where maximum N content was recorded in L_{60} and maximum P and K content was recorded in L_{100} . L_{60} level of lime enhanced available nitrogen by 15.7% over control, while L_{100} level of lime enhanced availability of phosphorus and potassium by 33% and 2.8% over control. Treatments L_{60} and L_{100} were found to be at par with each other for all the three parameters. Minimum N, P and K were recorded in the control treatment (L_0). Effect of varieties on available NPK content in post harvest soil was non-significant.

- 20. Effect of pH was non-significant on exchangeable Ca and significant on available B. Maximum available B content in soil at harvest was recorded in pH 5.2 (pH₂) soil, whereas minimum was recorded in pH 4.6 (pH₁) soil. Availability of boron was improved by 17.4% in pH₂ soil over pH₁. Effect of lime on exchangeable Ca and available B of post harvest soil was significant and maximum value of Ca was observed in 100% lime of LR (L₁₀₀) and maximum value of B was observed in 30% lime of LR (L₃₀). Lowest Ca and B content were observed in control treatment (L₀). The L₁₀₀ level of lime enhanced exchangeable calcium to the extent of 119.9%, while L₃₀ level of lime was observed to enhance available B by 8.3% over control. No significant effect of varieties was recorded on Ca and B content in soil.
- **21.** Significant effect of pH on exchangeable hydrogen, exchangeable aluminium and total potential acidity was observed, where highest values were observed in pH 4.6 (p_{H1}) soil and lowest values were observed in pH 5.2 (pH₂) soil. Exchangeable hydrogen, exchangeable aluminium and total potential acidity decreased by 32.6%, 9.2% and 6.3%, respectively in pH₂ as compared to pH₁. Effect of lime on exchangeable hydrogen, aluminium and total potential acidity in post-

harvest soil was significant where highest values were observed in control (L_0) and lowest in treatment of 100% lime of lime requirement (L_{100}). Exchangeable hydrogen, exchangeable aluminium and total potential acidity registered a decline of 65.8%, 43.6% and 28.8%, respectively in L_{100} when compared to L_0 . Varieties did not have any significant effect on exchangeable hydrogen, exchangeable aluminium and total potential acidity in post harvest soil. Interaction effect of pH and lime on exchangeable hydrogen and aluminium was significant where highest value was observed in the treatment combination pH_1L_0 and lowest in pH_2L_{100} . Irrespective of soil pH, exchangeable hydrogen and aluminium decreased as lime levels increased under each level of soil pH.

EFFECT OF LIME, PHOSPHORUS AND BORON ON PERFORMANCE OF MAIZE AND SOIL PROPERTIES (EXPT-II)

 Effect of lime was significant on plant height and maximum plant height was recorded in the treatment L₂₅ (25% lime of LR) and minimum was recorded in control (L₀) at 30, 60 DAS and at harvest. Effect of phosphorus on plant height was significant where 90 kg P₂O₅ ha⁻¹ (P₉₀) produced taller plants at all crop stages. P₉₀ was statistically at par with P₆₀ at 60 DAS and at harvest. Application of 90 kg P₂O₅ ha⁻¹ increased pooled plant height at 30, 60 DAS and at harvest by 12.04%, 21.2% and 21.7%, respectively over control. Minimum plant height at 30, 60 DAS and at harvest was recorded in control (P₀). Effect of boron was significant and maximum plant height was recorded at 2 kg B ha⁻¹ (B₂) while minimum plant height was recorded in control (B₀) at all the growth stages. B₂ level of boron increased plant height to the extent of 12.0%, 7.5% and 4.9%, respectively at 30, 60 DAS and at harvest over control. Interaction effect of lime and P on plant height was significant during 2016, where maximum was observed in L₂₅P₆₀ and minimum in
control (L_0P_0). It was observed that in treatments with lime, plant height increased up to 60 kg P_2O_5 ha⁻¹, while in treatments without lime, plant height increased at P level of 90 kg P_2O_5 ha⁻¹. Interaction effect of lime, P and B on plant height was significant at harvest during 2017, where maximum plant height was recorded in $L_{25}P_{60}B_2$ and minimum plant height was recorded in $L_0P_0B_0$. Thus, application of lime along with increased nutrient input led to better plant height. However, treatment combination $L_{25}P_{60}B_2$ was at par with $L_{25}P_{30}B_2$, and $L_{25}P_{90}B_2$.

- 2. Significant effect of lime on number of leaves was recorded and maximum value was found in the treatment L₂₅ at 30, 60 DAS and at harvest, while minimum number of leaves was recorded in control (L₀). Effect of phosphorus was significant and maximum number of leaves was recorded in the treatment P₉₀ at 30, 60 DAS and at harvest, while minimum was recorded in the treatment P₀ at all growth stages. At harvest, application of 90 kg P₂O₅ ha⁻¹ increased pooled number of leaves at 60 DAS and at harvest was significant where maximum number of leaves was recorded in the treatment B₂, while minimum was recorded in the treatment B₂, while minimum was recorded in the treatment B₂, while minimum was recorded in control (B₀). At harvest, B₂ level of boron was observed to improve the number of leaves to the extent of 6.1% in comparison to control (B₀).
- 3. Effect of lime on leaf area index was significant and maximum leaf area index was recorded in L_{25} , while minimum was recorded in L_0 at 30, 60 DAS and at harvest. Effect of phosphorus was significant and maximum leaf area index at all growth stages was recorded in the treatment P₉₀ which was at par with L_{60} , while minimum was recorded in the treatment P₀. Effect of boron on leaf area index was non-significant.
- **4.** Effect of lime on cob length, cob girth and cob weight was significant and maximum value was recorded at 25% lime of LR (L₂₅) over control

(L₀). Application of 25% lime of LR increased cob length, cob girth and cob weight by 2.3%, 3.0% and 21.1%, respectively over control. Effect of phosphorus was significant on cob length, cob girth and cob weight and was recorded maximum in treatment P_{90} and minimum in P_0 . The P₉₀ level of phosphorus increased pooled cob length, cob girth, cob weight by 12.2%, 8.8% and 22.5%, respectively over control. Treatment P₉₀ was at par with P₆₀ in case of cob weight. Effect of boron on cob length, cob girth and cob weight was also significant and recorded maximum in treatment B_2 and minimum in control (B_0). The B_2 level of boron enhanced cob length, cob girth and cob weight by 3.2%, 2.2% and 5.2%, respectively over control. Interaction effect of lime and phosphorus on cob weight was significant and highest cob weight was recorded in L₂₅P₆₀ treatment combination and minimum in L₀P₀. Cob weight increased up to the maximum P level (P_{90}) in the treatments without lime, whereas cob weight increased up to 60 kg P_2O_5 ha⁻¹ and declined at 90 kg P_2O_5 ha⁻¹ in the treatments with lime.

5. Significant effect of lime on number of grains per row, number of rows per cob and number of grains per cob was observed which recorded highest values in L₂₅ and lowest values in L₀. Phosphorus effect on number of rows per cob was non-significant, while effect on number of grains per row, and number of grains per cob were significant. Maximum number of grains per row and grains per cob were recorded in treatment P₉₀, while minimum was recorded in control (P₀). The P₉₀ level of phosphorus increased number of grains per row and number of grains per cob to the extent of 18.8% and 24.1%, respectively, over control. Treatment P₉₀ was at par with treatment P₆₀ in case of number of grains per cob, while significant effect was observed on number of grains per row and per cob. Maximum number of grains per row and number of grains per cob.

of grains per cob was observed in the treatment B_2 and minimum was observed in control (B_0). Interaction effect of lime and phosphorus on number of grains per row was significant during both the years, while on number of grains per cob, it was significant during 2016. Maximum value was recorded in $L_{25}P_{90}$ treatment combination in case of number of grains per row and in $L_{25}P_{60}$ treatment combination in case of number of grains per cob. Treatment combination $L_{25}P_{90}$ was found to be at par with $L_{25}P_{60}$ in case of number of grains per row, while treatment combination $L_{25}P_{60}$ was found to be at par with $L_{25}P_{30}$ and $L_{25}P_{90}$. It was observed that irrespective of lime treatments, number of grains per row increased with increase in P levels and vice versa.

6. Effect of lime on test weight of maize was non-significant, whereas effect of lime on grain and stover yield was significant, where highest grain and stover yield was observed in the treatment L₂₅ and lowest in control (L₀). The L₂₅ level of lime increased pooled grain and stover yield by 24.6% and 23.0%, respectively over control (L₀). Effect of phosphorus on test weight of maize was non-significant, whereas significant effect was observed in grain and stover yield. Maximum grain and stover yield was recorded in the treatment P₉₀, whereas minimum was recorded in control (P_0). The P_{90} level of phosphorus increased grain and stover yield by 24.3% and 22.1%, respectively over control. Treatment P₉₀ was at par with treatment P₆₀ in case of both grain and stover yield. Effect of boron on test weight was nonsignificant, whereas significant effect on grain and stover yield was observed. Highest grain and stover yield was recorded in B₂, while lowest was recorded in B₀. The B₂ level of boron increased pooled grain and stover yield by 10.9% and 10.6% over control. Interaction effect of lime and phosphorus on grain yield was significant, where maximum grain yield was recorded in the treatment combination $L_{25}P_{60}$, whereas minimum was recorded in L_0P_0 .

- 7. Significant effect of lime on N content in grain and stover was significant where 25% lime of LR (L₂₅) recorded the highest values over control (L₀). An increase by 20.0% and 21.4% in the nitrogen content in grain and stover, respectively was observed when lime was applied. Effect of phosphorus on N content in grain and stover was also significant, where highest N content in grain and stover was observed @ 90 kg P₂O₅ ha⁻¹ (P₉₀) and lowest in control (P₀). An increase in nitrogen content in grain by 10.7% and in stover by 17.5% in the P₉₀ treatment over control was observed. Treatment P₉₀ was at par with treatment P₆₀ in case of both grain and stover. Effect of boron on N content in grain and stover was significant, where highest N content in grain and stover was observed @ 2 kg B ha⁻¹ (B₂) and lowest in control (B₀). B₂ treatment increased the nitrogen content by 15% in grain and by 22.2% in stover over B₀ treatment. Treatment B₂ was at par with treatment B₁ in case of N content in grain.
- 8. Lime had a significant effect on the P content in grain and stover, where highest value was observed @ 25% lime of LR (L₂₅) over control (L₀). There was an increase by 17.9 % and 13.3% in the phosphorus content in grain and stover, respectively when lime was applied. Significant effect of phosphorus was observed, where maximum P content in both grain and stover was recorded in P₉₀, while minimum was recorded in control (P₀). There was an increase by 28.6% in case of phosphorus content in stover due to P₉₀ over control. Treatment P₉₀ was at par with treatment P₆₀ in case of P content in both grain and stover. Effect of boron on P content in grain and stover was observed @ 2 kg B ha⁻¹ (B₂) which was

at par with treatment B_1 , while lowest P content was observed in control (B_0). There was an increase by 15.8% in boron content in grain and an increase by 23.1% in boron content in stover due to B_2 over control.

- **9.** Significant effect of lime on K content in grain and stover was observed to be highest in the treatment L₂₅ over L₀. There was an increase by 10.9% in case of potassium content in grain and 4.6% in case of potassium content in stover due to liming. Highest K content in both grain and stover was recorded in the treatment P₉₀ and lowest K content was recorded in control (P₀) in both grain and stover. There was an increase in potassium content in grain by 15.1% and in stover by 8.4% in the treatment P₉₀ when compared to control. Treatment P₉₀ was at par with treatment P₆₀ in case of K content in both grain and stover. Effect of boron was non-significant on K content in grain and stover.
- **10.** Effect of lime on Ca content in grain and stover was significant where highest value was recorded in L₂₅. There was an increase by 60.0% in case of calcium content in grain and 58.3% in case of calcium content in stover in L₂₅ over L₀. Effect of phosphorus was significant and highest Ca content in grain and stover was observed in treatment P₉₀, while lowest was observed in control (P₀). An increase in calcium content by 28.9% and 24.9% in grain and stover, respectively in treatment P₉₀ over P₀ was observed. Effect of boron on Ca content in grain and stover was non-significant.
- **11.** Significant effect of lime on B content in grain and stover was recorded highest in the treatment L₂₅ and lowest in control (L₀). Liming (L₂₅) increased the boron content in grain by 2.1% and in stover by 5.0%. Effect of phosphorus on B content was non-significant. Effect of boron on B content in grain and stover was significant, where highest B content for both grain and stover was recorded in B₂, while minimum was recorded in B₀. The B₂ level of boron enhanced pooled B content in

grain and stover by 39.0% and 31.8%, respectively over control. Treatment B_2 was at par with treatment B_1 in case of B content in stover. Interaction effect of lime and boron on boron content in stover was significant, where highest B content was observed in treatment combination $L_{25}B_2$ which was at par with $L_{25}B_1$ and lowest B content was observed in treatment combination L_0B_0 .

- **12.** Significant effect of lime on N uptake was recorded and highest value was recorded in L₂₅ in both grain and stover. There was an increase in nitrogen uptake by 49.8% and 50.3% in grain and stover, respectively due to liming (L_{25}) over control (L_0) . Significant effect of phosphorus on N uptake was recorded in P₉₀, while minimum was recorded in control (P_0) in both grain and stover. There was an increase in nitrogen uptake by 37.1% and 44.4% in grain and stover, respectively due to P₉₀ level over control (P_0). Treatment P_{90} was at par with treatment P_{60} in case of N uptake in grain. Effect of boron on N uptake in grain as well as stover was significant and maximum uptake was recorded in the treatment B₂, while minimum was recorded in control (B₀). B₂ level increased nitrogen uptake in grain by 20.7% and in stover by 26.7% over control. Interaction effect of lime and phosphorus on N uptake in grain was significant during 2016, where highest N uptake was observed in treatment combination L₂₅P₆₀ and minimum was observed in treatment combination L_0P_0 .
- 13. Effect of lime on P uptake in grain and stover was significant which recorded a maximum in L₂₅. Phosphorus uptake in grain and stover increased by 45.9% and 44.4%, respectively due to liming (L₂₅) over control (L₀). Effect of phosphorus on P uptake in grain and stover was significant with maximum P uptake in P₉₀ in both grain and stover. There was an increase in phosphorus uptake by 58.5% and 55.7%, respectively in grain and stover due to application of 90 kg P₂O₅ ha⁻¹

(P₉₀) and 60 kg P₂O₅ ha⁻¹ (P₆₀), respectively over control. P uptake in stover in treatment P₉₀ was at par with treatment P₆₀. Minimum P uptake was observed in the treatment P₀ in both grain and stover. Effect of boron on P uptake in grain and stover was significant, where maximum uptake was recorded in the treatment B₂, while minimum was recorded in control (B₀). There was an increase in phosphorus uptake by 27.1% and 41.3%, respectively in grain and stover due to application of 2 kg B ha⁻¹ (B₂) over control (B₀). Interaction effect between lime and phosphorus on P uptake in grain was significant during 2016, where maximum uptake was recorded in the treatment combination L₂₅P₉₀ was at par with L₂₅P₆₀.

- 14. Effect of lime on K uptake in grain and stover was significant which recorded the highest uptake in limed treatment (L₂₅) over control (L₀). Percent increase by 37.7% and 29.5%, respectively in grain and stover was observed in pooled data due to application of L₂₅ level of lime over control. Phosphorus effect was significant and highest K uptake in grain and stover was recorded in P₉₀, while minimum K uptake was recorded in treatment P₀. An increase in potassium uptake in grain by 42.7% and in stover by 33.1% due to application of 90 kg P₂O₅ ha⁻¹ over control was observed. However, in case of K uptake in stover, treatment P₉₀ was at par with treatment P₆₀. Effect of boron on K uptake in grain and stover was significant, where maximum uptake was recorded in the treatment B₂, while minimum was recorded in control (B₀). K uptake in grain and stover increased by 12.2% and 11.2%, respectively from treatment B₀ to B₂. Treatment B₂ was at par with B₁ in case of K uptake in both grain and stover.
- **15.** Effect of lime on Ca uptake was significant, where maximum Ca uptake was recorded in treatment L₂₅ over L₀. An increase by 98.6% and 97.4%

in calcium uptake by grain and stover, respectively could be observed with application of L_{25} over L_0 . Phosphorus effect was significant on Ca uptake and maximum value was observed in the treatment P_{90} and minimum in the treatment P_0 in both grain and stover. There was an increase by 60.8% and 50.3% on calcium uptake in grain and stover, respectively due to 90 kg P_2O_5 ha⁻¹ (P_{90}) application over control. Effect of boron was non-significant for Ca uptake in both grain and stover.

16. Effect of lime was significant on B uptake in grain and stover and highest uptake was recorded in lime treatment (L_{25}) over control (L_0) . Due to L_{25} level of lime, an increase in boron uptake by 27.0% and 29.3% in grain and stover, respectively was observed. Effect of phosphorus was significant and treatment P₉₀ recorded maximum B uptake in both grain and stover which was however found to be at par with P_{60} . Control treatment (P_0) recorded minimum B uptake in both grain and stover. Due to P₉₀ level of phosphorus, an increase in boron uptake by 23.9% and 22.7% in grain and stover, respectively could be observed. Effect of boron was also significant and treatment B₂ recorded maximum B uptake, while treatment B₀ recorded minimum in both grain and stover. An increase in boron uptake by 54.2% and 45.9% in grain and stover, respectively due to B₂ level of boron application could be observed. Interaction effect of lime and phosphorus on boron uptake was significant in case of grain and highest value was observed in the treatment combination $L_{25}P_{60}$ and lowest in L_0P_0 . It was observed that in lime applied treatments, 60 kg P2O5 ha-1 (P60) showed maximum B uptake, while in treatments without lime, 90 kg P₂O₅ ha⁻¹ (P₉₀) showed maximum B uptake. Irrespective of liming, B uptake in grain increased with increase in P levels and vice versa. Interaction effect of lime and boron on boron uptake was significant in case of both grain and stover, where maximum boron uptake in both grain and stover was observed in treatment combination $L_{25}B_2$ and minimum was observed in treatment combination L_0B_0 . Interaction effect of phosphorus and boron on boron uptake was significant in case of grain during 2016, where maximum was observed in treatment combination $P_{90}B_2$ and minimum in P_0B_0 .

- 17. Lime had a significant effect on soil pH and non-significant effect on electrical conductivity. Maximum pH was recorded in the treatment L₂₅, while minimum was recorded in the treatment L₀. An increase by 8.8% in the soil pH was observed due to liming (L₂₅) over control (L₀). Treatment effect of phosphorus and boron on soil pH and electrical conductivity was non- significant.
- 18. Lime application was significant on percent base saturation and nonsignificant on soil organic carbon. Maximum percent base saturation of post harvest soil was recorded in L₂₅ and minimum in L₀. Effect of phosphorus and boron levels on percent base saturation and organic carbon were non-significant.
- 19. Effect of lime on available N, P and K of post harvest soil was significant. Maximum available N, P and K was recorded in L₂₅, while minimum was recorded in L₀. Effect of phosphorus on available N and P was significant, while its effect on available K was non-significant. There was no significant difference among the treatments with respect to boron application on available N, P and K.
- 20. Lime had a significant effect on exchangeable Ca and available B of post harvest soil, where maximum values for both parameters were recorded in L₂₅ and minimum in L₀. Effect of phosphorus on available B and exchangeable Ca was non-significant. There was no significant effect of boron on exchangeable Ca, whereas significant effect of boron on available boron was observed. The application of boron @ 2 kg ha⁻¹ (B₂) recorded the highest value while control treatment (B₀) recorded the lowest value.

- **21.** Effect of liming on exchangeable hydrogen, exchangeable aluminium and total potential acidity in post harvest soil was significant, where maximum value was recorded in control treatment (L₀) and minimum value was recorded @ 25% lime of LR (L₂₅). A percent decrease in exchangeable hydrogen, aluminium and total potential acidity by 37.9%, 8.3% and 11.3%, respectively was observed with application of L₂₅ level. There was no significant effect of phosphorus and boron on exchangeable hydrogen, aluminium and total potential acidity in post harvest soil.
- 22. Agronomic efficiency of P and B decreased with increase in the dose of phosphorus and boron, respectively. Agronomic efficiency of P was maximum in P₃₀ and minimum in P₉₀, whereas AE of boron was maximum in B₁ and minimum in B₂.
- **23.** The nutrient use efficiency of P and B was observed to decrease with increase in level of phosphorus and boron, respectively. Higher phosphorus use efficiency among treatments was obtained at 30 kg P_2O_5 ha⁻¹ (P_{30}), thereafter it significantly reduced, where lowest was obtained in P_{90} . Boron use efficiency by maize was higher at 1 kg B ha⁻¹ (B_1) as compared to 2 kg B ha⁻¹ (B_2).

CONCLUSION

The following conclusions can be drawn from the following summary:

- 1. Low pH soil affected adversely the plant growth, yield attributes, grain and stover yield, nutrient uptake of maize and soil properties.
- 2. Plant growth, yield attributes, yield, nutrient content and their uptake by maize significantly enhanced on application of lime. Liming at 60% lime of LR proved to be optimum in order to get the desired yield. Application of lime enhanced the pH, base saturation and nutrient content of post harvest soil, while it reduced the exchangeable hydrogen, exchangeable aluminium and total potential acidity of the

soil. Among the maize varieties, RCM-76 proved superior over RCM-75 and RCM-1-1 under both soil pH levels. Superiority on the performance of maize varieties in both soil pH 4.6 and 5.2 followed the order: RCM-76 > RCM-75 > RCM-1-1.

- 3. Plant growth, yield attributes, grain and stover yield, nutrient content and nutrient uptake by maize improved with application of phosphorus and boron. Available P and B of post harvest soil also improved with application of the respective nutrients. Application of 60 kg P₂O₅ ha⁻¹ and 2 kg B ha⁻¹ proved optimum in order to get good performance of maize. However, higher nutrient use efficiency and agronomic efficiency was observed on application of 30 kg P₂O₅ ha⁻¹ and 1 kg B ha⁻¹.
- 4. As observed from the outcome of the experiment, cultivation of maize variety RCM-76 along with liming at 60% lime of LR may be recommended for cultivation of maize in very strongly acidic sandy clay loam soils. In strongly acid soils application of 60 kg P₂O₅ ha⁻¹ and 2 kg B ha⁻¹ combined with liming at 25% lime of LR may be recommended for cultivation of maize in sandy clay loam soils of Nagaland.

REFERENCES

- Abreu, C. H. Muraoka, T. and Lavorante, A. F. 2003. Relationship between acidity and chemical properties of Brazilian soils. *Scientia Agricola*. **60**(2): 337-343.
- Adem, G. N., Zametin, A., Asliham, E., Oguzhan, U., Sinan, A. and Metin, T. 2011. Yield and chemical composition of corn (*Zea mays* L.) as affected by boron management. *International Journal of Plant, Animal and Environmental Science*. **1**(1): 42-51.
- Adhikary, B. H. and Karki, K. B. 2007. Use of fertilizers and lime for enhancing productivity of maize genotypes in Western Hill of Nepal. *Nepal Agriculture Research Journal.* **8**: 42-49.
- Adie, M. M. and Krisnawati, A. 2016. Identification of soybean genotypes adaptive and productive to acid soil agro-ecosystem. *Biodiversitas*. **17**(2): 565-570.
- Adikuru, N. C., Ogoke, I. J., Anyanwu, C. P. and Uzoho, B. U. 2019. Liming effects on reproductive growth and yield components of maize grown on an acid rainforest soil. *Journal of Agriculture and Rural Development in the Tropics and Subtropics*. **120**(2): 141-147.
- Adiloglu, A. and Adiloglu, S. 2006. The effect of boron (B) application on the growth and nutrient contents of maize in zinc (Zn) deficient soils. *Research Journal of Agriculture and Biological Sciences* **2**(1): 1-4.
- Adnan, A., Mavinic, D. S. and Koch, F. A. 2003. Pilot-scale study of phosphorus recovery through struvite crystallization examining to process feasibility. *Journal of Environmental Engineering and Science*. **2**(5): 315-324.
- Agricultural Statistics at a Glance, 2019. Government of India, Ministry of Agriculture & Farmers Welfare, Department of Ministry of Agriculture, Cooperation & Farmers Welfare, Directorate of Economics and Statistics.
- Ahmed, N., Abid, M. and Ahmad, F. 2008. Boron toxicity in irrigated cotton (*Gossypium hirsutum* L.). *Pakistan journal of Botany*. **40**: 2443-2452.
- Akhter, S. and Mahmud, N. U. 2009. Response of hybrid maize to boron and lime. Annual Research Report, 2008-09. Soil Science Division, Bangladesh Agricultural Research Institute, Joydebpur, Gazipur, pp. 244-253.
- Alam, S. M., Naqvi, S. S. M. and Ansari, R. 1999. Impact of soil pH on nutrient uptake by crop plants. In: Handbook of plant and crop stress, 2nd edition (ed. Mohammad Pessarakli), Marcel Dekku, INC, New York. pp-51-60.
- Ali, J., Bakht, J., Shafi, M., Khan, S. and Shah, W. A. 2002. Uptake of nitrogen as affected by various combinations of nitrogen and phosphorus. *Asian Journal of Plant Sciences.* 1: 367-369.

- Ali, S., Khan, R., Mairaj, G., Arif, M., Fida, M. and Bibi, S. 2008. Assessment of different crop nutrient management practices for yield improvement. *Australian Journal of Crop Science*. 2(3): 150-157.
- Alias, A., Usman, M., Ullah, E. and Warraich, E.A. 2003. Effect of different phosphorus levels on the growth and yield of two cultivars of maize. *International Journal of Agriculture and Biology*. 4: 632-634.
- Amanullah and Khan, A. 2015. Phosphorus and compost management influence maize (*Zea mays*) productivity under semiarid condition with and without phosphate solubilizing bacteria. *Frontiers in Plant Science*. Volume 6, Article 1083.
- Amanullah, Asif, M., Malhi, S. S. and. Khattak, R. A. 2009. Effects of P-fertilizer source and plant density on growth and yield of maize in Northwestern Pakistan. Journal of Plant Nutrition. 32: 2080-2093.
- Amanullah, Zakirullah, M., Tariq, M., Nawab, K., Khan, A. Z., Farhatullah, Shah, Z., Jan, A., Khalil, S.K., Jan, M. T., Sajid, M., Hussain, Z. and Rahman, H. U. 2010. Levels and time of phosphorus application influence growth, dry matter partitioning and harvest index in maize. *Pakistan Journal of Botany*. 42(6): 4051-4061.
- Amer, W., Sharma, Y. K. and Sharma, S. K. 2014. Response of maize to integrated nutrient management in acidic soils of Nagaland. *Annals of Plant and Soil Research*. 16 (4): 349-352.
- Andersson, S. and Nilsson, S. I. 2001. Infuence of pH and temperature on microbial activity, substrate availability of soil-solution bacteria and leaching of dissolved organic carbon in a mor humus. *Soil Biology and Biochemistry*. **33**: 1181–1191.
- Andric, L., Rastija, M., Teklic, T. and Kovacevic, V. 2012. Response of maize and soybean to liming. *Turkish Journal of Agriculture and Forestry*. **36**: 415-420.
- Aref, F. 2011. The effect of boron and zinc application on concentration and uptake of nitrogen, phosphorous and potassium in corn grain. *Indian Journal of Science and Technology.* 4(7): 785-791.
- Arora, S., and Chahal, D. S. 2005. Available boron content in some benchmark soils of Punjab under different moisture regimes in relation to soil characteristics. *Agropedology*. 15: 90-94.
- Arunkumar, B. R. and Srinivasa, N. 2018. Effect of gypsum and borax application on yield, nutrient content and uptake in maize under different nutrient management practices. *International Journal of Pure & Applied Bioscience*. 6(4): 181-189.

- Arya, K. C. and Singh, S. N. 2000. Effects of different levels of phosphorus and zinc on yield and nutrients uptake of maize with and without irrigation. *Indian Journal of Agronomy*. 45(2): 717-721.
- Assuero, S. G., Mollier, A. and Pellerin, S. 2004. The decrease in growth of phosphorusdeficient maize leaves is related to a lower cell production. *Plant, Cell & Environment.* 27: 887-895.
- Awani, Z. P. 2012. Effects of wood ash and poultry manure on leaf nutrient content, growth and yield of spinach. *Plant Nutrition*. **18**(16): 141-147.
- Awomi, T. A., Singh, A. K., Kumar, M. and Bordoloi, L. J. 2012. Effect of phosphorus, molybdenum and cobalt nutrition on yield and quality of mungbean (*Vigna radiata* L.) in acidic soil of Northeast India. *Indian Journal of Hill Farming*. 25(2): 22-26.
- Ayden, A. and Sevin, A. 2006. The effect of boron application on the growth and nutrient contents of maize in zinc deficient soil. *Bulgarian Journal of Agricultural Science*. 12: 387-392.
- Ayub, M., Nadeem, M. A., Sharar, M. S. and Mahmood, N. 2002. Response of maize (Zea mays L.) fodder to different levels of nitrogen and phosphorus. Asian Journal of Plant Sciences. 1: 352-354.
- Badole, S., Datta, A., Basak, N., Seth, A., Padhan, D. and Mandal, B. 2015. Liming influences forms of acidity in soils belonging to different orders under Subtropical India. *Communications in Soil Science and Plant Analysis*. **46**(16): 2079-2094.
- Bak, K., Gaj, R and Budka, A. 2016. Accumulation of nitrogen, phosphorus and potassium in mature maize under variable rates of mineral fertilization. *Fragmenta Agronomica*. 33(1): 7-19.
- Baligar, V. C., Pitta, G. V. E., Gama, E. E. G., Schaffert, R. E., Bahia Filho, A. F. C. and Clark, R. B. 1997. Soil acidity effects on nutrient use efficiency in exotic maize genotypes. *Plant and Soil.* 192: 9-13.
- Barman, M., Shukla, L. M., Datta, S. P. and Rattan, R. J. 2014. Effect of applied lime and boron on the availability of nutrients in an acid soil. *Journal of Plant Nutrition*. 37: 357-373.
- Baruah, T. C. and Barthakur, H. P. 1997. A textbook of soil analysis. Vikas Publishing House Pvt. Ltd., 576 Masjid Road, Jangpura, New Delhi.
- Basak, B. 2010. Nutrient dynamics and chemical properties of acid soil under different liming condition. M.Sc. (Ag) Thesis, Department of Agricultural Chemistry, Hajee Mohammad Danesh Science and Technology University, Dinajpur.

- Behera, R. D., Das, S. and Pattanayak, S. K. 2017. Various impact of different sources of liming materials on growth, yield and productivity of the maize crop grown in acid soil of Odisha. *Journal of Pharmacognosy and Phytochemistry*. 6(5): 1831-1835.
- Behera, S. K. and Shukla, A. K. 2015. Spatial distribution of surface soil acidity, electrical conductivity, soil organic carbon content and exchangeable potassium, calcium and magnesium in some cropped acid soils of India. Land Degradation and Development. 26(1): 71-79.
- Bekele, A., Kibret, K., Bedadi, B., Balemi, T. and Yli-Halla, M. 2018. Effects of lime, vermicompost and chemical P fertilizer on yield of maize in Ebantu district, Western Highlands of Ethiopia. *African Journal of Agricultural Research*. **13**(10): 477-489.
- Bell, R. W. and Dell, B. 2008. Micronutrients for sustainable food, feed, fiber and bioenergy production. First edition, International Fertilizer Industry Association. Paris, France.
- Benjala, M. J., Maida, J. H. A., Lowole, M.W. and Kabambe, V. H. 2015. Liming and fertiliser P interaction effects on some indices of fertility of selected Malawi acidic soils. *Journal of Soil Science and Environmental Management*. 6 (9): 249-259.
- Beukes, D. J., Mapumulo, T. C., Fyfield, T. P. and Jezile, G. G. 2012. Effects of liming and inorganic fertiliser application on soil properties and maize growth and yield in rural agriculture in the Mbizana area, Eastern Cape province, South Africa. South African Journal of Plant and Soil. 29(3&4): 127-133.
- Bhat, J. A., Mandal, B. and Hazra, G. C. 2007. Basic slag as a liming material to ameliorate soil acidity in Alfisols of sub-tropical India. *American-Eurasian Journal of Agricultural & Environmental Sciences.* 2(4): 321-327.
- Bhindhu, P. S., Sureshkumar P., Mini Abraham and Kurien, E. K. 2018. Effect of liming on soil properties, nutrient content and yield of wetland rice in acid tropical soils of Kerala. *International Journal of Bio-resource and Stress Management*. 9(4): 541-546.
- Black, C. A. 1965. Methods of Soil Analysis. American Society of Agronomy, Inc, Publisher, Madison, Wisconsin, USA. pp 171-175.
- Borase, C. L., Lomte, D. M., Thorat, S. D. and Dhonde, A. S. 2018. Response of *kharif* maize (*Zea mays* L.) to micronutrients. *Journal of Pharmacognosy and Phytochemistry*. **7**(3): 482-484.
- Brajendra, Vishwakarma, A. K. and Pathak, K. A. 2006. Effect of varying doses of lime on yield and attributes of maize in Mizoram. *Indian Journal of Hill Farming*. **19**(1 & 2): 142-144.
- Bray, R. H. and Kurtz, L. T. 1945. Determination of total organic and available forms of phosphorus in soils. *Soil Science*. **59**: 39-45.

- Brown, T. T., Koenig, R. T., Huggins, D. R., Harsh, J. B. and Rossi, R. E. 2008. Lime effects on soil acidity, crop yield, and aluminum chemistry in direct-seeded cropping systems. *Soil Science Society of America Journal*. **72**: 634-640.
- Bruckner, M. Z. 2012. Water and soil characterization pH and electrical conductivity. Microbial life educational resources, Montana State University, Bozeman. https://serc.carleton.edu. Accessed on 12 April 2020.
- Busari, M. A., Salako, F. K. and Adetunji, M. T. 2008. Soil chemical properties and maize yield after application of organic and inorganic amendments to an acidic soil in Southwestern Nigeria. *Spanish Journal of Agricultural Research*. **6**(4): 691-699.
- Busari, M. A., Salako, F. K., Sobulo, R. A., Adetunji, M. T. and Bello, N. J. 2005. Variation in soil pH and maize yield as affected by the application of poultry manure and lime.
 In: Managing Soil Resources for Food and Sustainable Environment. Proceedings of 29th Annual Conference of Soil Science Society of Nigeria held at University of Agriculture, Abeokuta, Nigeria, December 6-10. pp-139-142.
- Byju, G., Nedunchezhiyan, M. and Naskar, S. K. 2007. Sweet potato response to boron application on an Alfisol in the sub humid tropical climate of India. *Communications in Soil Science and Plant Analysis.* **38**: 2347-2356.
- Caires, E. F., Feldhaus, I. C., Barth, G. and Garbuio, F. J. 2002. Lime and gypsum application on the wheat crop. *Scientia Agricola*. **59**(2): 357-364.
- Ceyhan, E., Onder, M., Harmankaya, M. and Gezgin, S. 2007. Response of chickpea cultivars to application of boron in boron-deficient calcareous soils. *Communications in Soil Science and Plant Analysis*. **38**: 1-19.
- Chanchal, A., Singh, S. K., Patra, A. and Jatav, S. S. 2020. Direct and residual effect of boron application on yield and nutrients content under rice–wheat cropping system. *Current Journal of Applied Science and Technology*. **39**(2): 12-22.
- Chandrakala, M., Srinivasamurthy, C. A., Kumar, S. and Naveen, D. V. 2017. Effect of application of graded level of phosphorus to finger millet - maize cropping system in soils of different P fertility. *International Journal of Current Microbiology and Applied Sciences*. 6(11): 265-280.
- Chapman, D. H. and Pratt, P. F. 1961. Methods of analysis of soils, plants and water. University of California, Riverside, Division of Agriculture Science. pp 309.
- Chapman, H. D. 1965. CEC by ammonium saturation. *American Journal of Agronomy*. **9**: 891-901.
- Chatterjee, A., Dosani, A. A. K., Talashikhar, S. C. and Mehta, V. B. 2005. Effect of lime on yield, quality and nutrient uptake by six groundnut varieties and properties of an Alfisol. *Journal of the Indian Society of Soil Science*. **53**(1): 128-132.

- Chen, M. L., Jiang, X. L., Zoov, B. Y. and Zheri, Z. Y. 1994. Mathematical models and best combination of high yield cultivation technique for rapeseed variety Zhenyouyoum. *Acta Agriculturae Zhejiiangenesis* **6**: 22-26.
- Chirnogeanu, I., Badea, E., Petcu, E. and Picu, I. 1997. The effect of mineral fertilization on uptake of some nutrients in maize under irrigated conditions. *Romanian Agricultural Research.* **7 8**: 55-58.
- Chowdhury, A. K., Singh, G., Tyagi, B. S., Bhattacharya, P. M. and Singha Roy, A. K. 2008. Assessment of wheat (*Triticum aestivum*) cultivars to boron deficiency-induced spike sterility and its impact on grain yield under Terai region of West Bengal. *Indian Journal of Agricultural Sciences* 78(10): 834-7.
- Chuan, L., Ping, H., Pampolino, M. F., Johnston, A. M., Jin, J., Xu, X., Zhao, S. C., Qiu, S. J. and Zhou, W. 2013. Establishing a scientific basis for fertilizer recommendations for wheat in China: Yield response and agronomic efficiency. *Field Crops Research*. 140: 1-8.
- Clark, R. B. and Mgema, W. G., 1993. Screening sorghum for tolerance to excess manganese in solution culture. *Plant and Soil*. **155/156**: 493-496.
- Costa, M.C.G. 2012. Soil and crop responses to lime and fertilizer in a fire free land use system for small holdings in the northern Brazilian Amazon. *Soil & Tillage Research*. **121**: 27-37.
- Darwesh, D. A., Maulood, P. M. and Amin, S. A. 2013. effect of phosphorus fertilizers on growth and physiological phosphorus use efficiency of three soybean cultivars. *Journal of Agriculture and Veterinary Science*. **3**(6): 32-36.
- Das, R. and Saha, D. 2014. Effect of liming on the changes of different forms of potassium in an acid soil treated with N and K fertilizers. *Journal of the Indian Chemical Society*. 91: 1-7.
- Dasog, G. S., Patil, P. L. and Gali, S. K. 2010. Effect of groundnut genotypes, lime and phosphorus levels on the transformation of phosphorus in lowland acidic soils of Coastal Agro-eco-system of Karnataka. *Journal of the Indian Society of Soil Science*. 58(2): 182-188.
- Debnath, P and Ghosh, S. K. 2011. Determination of critical limit of available boron for rice in Terai zone soils of West Bengal. *Journal of the Indian Society of Soil Science* **59**(1): 82-86.
- Devkota, S., Panthi, S. and Shrestha, J. 2019. Evaluation of maize varieties in acid soil condition. *Agrica*. 8: 128-133.

- Dewi-Hayati, P. K., Sutoyo, Syarif, A. and Prasetyo, T. 2014. Performance of maize singlecross hybrids evaluated on acidic soils. *International Journal on Advanced Science Engineering Information Technology*. 4: 30-33.
- Dey, D. and Nath, D. 2015. Assessment of effect of liming and integrated nutrient management on groundnut under acidic soil condition of West Tripura. *An Asian Journal of Soil Science*. **10**(1): 149-153.
- Dible, W. T., Truog, E. and Berger, K. C. 1954. Boron determination in soils and plants: Simplified curcumin procedure. *Analytical Chemistry*. **26**: 418-421.
- Dixit, S. P. 2006. Effect of lime and phosphorus on yield and nutrients uptake by maize in mountain acidic soil of Himacahal Pradesh. *Annals of Agricultural Research.* **27**(3): 277-282.
- Dubey, S. K. 2000. Effectiveness of rock phosphate and superphosphate amended with phosphate solubilizing microorganisms in soybean grown on Vertisols. *Journal of the Indian Society of Soil Science*. **48**: 71-75.
- Etabo, E. M., Wafula, W. N., Korir, N. K. and Gweyi-Onyango, J. P. 2018. Effect of phosphorus levels on soil properties and plant tissues of two Nerica varieties. *Asian Soil Research Journal.* **1**(3): 1-9.
- Fageria, N. K. and Baligar, V. C. 2008. Ameliorating soil acidity of tropical Oxisols by liming for sustainable crop production. In: Advances in Agronomy, volume 99 (ed. Sparks, D. L.), Academic Press. pp- 345-389.
- Fageria, N. K. and Zimmermann, F. J. P. 1998. Influence of pH on growth and nutrient uptake by crop species in an Oxisol. *Communications in Soil Science and Plant Analysis.* 29: 2675-2682.
- Fageria, N. K., Baligar, V. C. and Jones, C. A. 2010. Growth and mineral nutrition of field crops. 3rd Edition. CRC Press. New York. London. pp- 586.
- Ferdous, S. A., Miah, M. N. H., Hoque, M., Hossain, M. S. and Hasan, A. K. 2018. Enhancing rice yield in acidic soil through liming and fertilizer management. *Journal* of Bangladesh Agricultural University. 16(3): 357-365.
- Foy, C. D. 1984. Physiological effects of hydrogen, aluminum and manganese toxicities in acid soils. In: Soil acidity and liming, 2nd Ed (ed. F, Adams). pp 5-97.
- Foy, C. D. 1988. Plant adaptation to acid, aluminium-toxic soils. *Communications in Soil Science and Plant Analysis.* **19**: 959-987.
- Friesen, D. K., Miller, M. H. and Juo, A. S. R. 1980. Liming and lime-phosphorus-zinc interactions in two Nigerian Ultisols: II. Effects on maize root and shoot growth. *Soil Science Society of America Journal.* 44: 1227-1232.

- Gaines, T. P. and Mitchell, G. A. 1979. Boron determination in soils and plants by the azomethine-H method. *Communication in Soil Science and Plant Analysis*. **10**: 1099-1108.
- Gale, J., Koenig, R. and Barnhill, J. 2001. Managing soil pH in Utah. Utah State University Extension. https://digitalcommons.usu.edu/extension_curall/923 Accessed on 12 March 2020.
- Garica, F.V. 1975. Depth of liming on very acid soils. M.Sc. Thesis No. 842, Asian Institute of Technology, Bangkok, Thailand.
- Gazala, N., Upinder, S. and Pardeep, K. 2016. Boron its importance in crop production, status in Indian soils and crop responses to its application. *International Journal of Advanced Research*. **4**(5): 654-660.
- Ghosh, B. N. and Mukhopadhyay, A. K. 2001. Effect of liming on potassium release in a Oxic Paleustalf. *Annals of Agricultural Research*. 22(3): 377-381.
- Giller, K. E., Cadisch, G. and Palm, C. 2002. The North-South Divide! Organic wastes or resources for nutrient management? *Agronomie*. **22**: 645-653.
- Goldbach, H. E. and Wimmer, M. A. 2007. Boron in plants and animals: Is there a role beyond cell structure? *Journal of Plant Nutrition and Soil Science* **170**: 39-48.
- Goldberg, S. 1997. Reaction of boron with soils. *Plant and Soil.* 193: 35-48.
- Gomez, K. A. and Gomez, A. A. 1984. Statistical procedures for agricultural research. 2nd Edition. Wiley Interscience Publication. pp 139-153.
- Goswami, S. P., Sacchidanand, B., Dubey, A. N. and Upadhyay, A. K. 2019. Effect of phosphorus levels on electrochemical properties, growth, yield and quality of soybean [*Glycine max* (L.)]. *Annals of Agricultural Research.* **40**(2): 1-7.
- Grazia, J. D., Tittonell, P. A., Germinara, D. and Chiesa, A. 2003. Phosphorus and nitrogen fertilization in sweet corn (*Zea mays L*) var. saccharata Bailey. *Spanish Journal of Agricultural Research*. **1**(2): 103-107.
- Günes, A., Ataoglu, N., Esringü, A., Uzun, O., Ata, S. and Turan, M. 2011. Yield and chemical composition of corn (*Zea mays L.*) as affected by boron management. *International Journal of Plant, Animal and Environmental Sciences*. **1**(1): 42-53.
- Gupta, R. K., Rai, R. N., Singh, R. D. and Prasad, R. N. 1989. Effect of liming acid soil on yield of some crops and soil characteristics. *Journal of the Indian Society of Soil Science*. 37: 126-130.

- Hamdi, E. L. and Woodard, H. J. 2004. Response of early corn growth to fertilizer phosphorus rates and placement methods. *Journal of Plant Nutrition*. 27: 1103-1120.
- Han, T., Cai, A., Liu, K., Huang, J., Wang, B., Li, D., Qaswar, M., Feng, G. and Zhang, H. 2019. The links between potassium availability and soil exchangeable calcium, magnesium, and aluminum are mediated by lime in acidic soil. *Journal of Soils and Sediments*. 19: 1382-1392.
- Haynes, R. J. 1982. Effects of liming on phosphate availability in acid soils. *Plant and Soil.* 68: 289-308.
- Haynes, R. J. and Naidu, R. 1998. Influence of lime, fertilizer and manure applications on soil organic matter content and soil physical conditions: a review. *Nutrient Cycling in Agroecosystems*. **51**: 123-137.
- Horst, W. J., Puschel, A. K. and Schmohl, N. 1997. Induction of callose formation is a sensitive marker for genotypic aluminium sensitivity in maize. *Plant and Soil.* 192: 23-30.
- Hue, N. V. 2004. Responses of coffee seedlings to calcium and zinc amendments to two Hawaiian acid soils. *Journal of Plant Nutrition*. **27**(2): 261-274.
- Hussain, N., Khan, A. Z., Akbar, H. and Akhtar, S. 2006. Growth factors and yield of maize as influenced by phosphorus and potash fertilization. *Sarhad Journal of Agriculture*. 22(4): 579-583.
- Hussain, N., Khan, A. Z., Akbar, H., Bangash, N. G., Khan, Z. H. and Idree, M. 2007. Response of maize varieties to phosphorus and potassium levels. *Sarhad Journal of Agriculture*. 23(4): 881-887.
- Hussein, A. H. A. 2009. Phosphorus use efficiency by two varieties of corn at different phosphorus fertilizer rates. *Research Journal of Applied Science*. **4**: 85-93.
- Inal, A. and Tarakcioglu, C. 2001. Effects of nitrogen form on growth, nitrate accumulation, membrane permeability and nitrogen use efficiency of hydroponically grown bunch onion under boron deficiency and toxicity. *Journal* of *Plant Nutrition*. 24: 1521-1534.
- Iqbal, T., Sale, P. and Tang, C. 2010. "Phosphorus ameliorates aluminium toxicity of Al sensitive wheat seedlings," In: Proceedings of the 19th World Congress of Soil Science, Soil Solutions for a Changing World, 92-95.
- Irfan, M., Abbas, M., Shah, J. A., Depar, N., Memon, M. Y. and Sial, N. A. 2019. Interactive effect of phosphorus and boron on plant growth, nutrient accumulation and grain yield of wheat grown on calcareous soil. *Eurasian Journal of Soil Science*. 8(1): 17-26.

- Islam, M. A., Islam, M. R. and Sarker, A. B. S. 2008. Effect of phosphorus on nutrient uptake of Japonica and Indica rice. *Journal of Agriculture and Rural Development*. 6(1&2): 7-12.
- Jackson, M. L. 1973. Soil Chemical Analysis. Prentice Hall of India Private Limited, New Delhi.
- Jaetzold, R., Schmidt, B. and Shisanya, C. A. 2006. Farm management handbook of Kenya. Natural Conditions and Farm Information, 2nd edition. Ministry of Agriculture, Nairobi, Kenya (Eastern Province).
- Jibrin, J. M., Chude, V. O., Horst, W. J. and Amapu, I. 2002. Effect of cover crops, lime and rock phosphate on maize (*Zea mays* L.) in an acidic soil of Northern Guinea Savanna of Nigeria. *Journal of Agriculture and Rural Development in the Tropics and Subtropic.* 103(2): 169-176.
- Kai, L. Z., Ngoh, G., Peng, H., Min, G. M., Ken, T. Y., Green, L. and Student, D. 2012. Investigating the effect of soil pH on the germination of *Avicennia alba* seedlings. Little Green Dot Student Research Grant Project Report submitted to nature society (Singapore). https://www.nss.org.sg. Accessed on 12 April 2020.
- Kamaruzzaman, M., Islam, M. N., Siddique, M. N., Sarker, B. C., Islam, M. J. and Rasel, S. M. M. 2014. Liming effect on changes of soil properties of wheat field: A case of Barind area in Bangladesh. *Online Journal of BioSciences and Informatics*. 1(1). www.JournalOnline.in. Accessed on 1 April 2020.
- Kanwar, K. and Paliyal, S. S. 2002. Influence of phosphorus management and organic manuring on uptake and yield of chickpea (*Cicer arietinum*). Annals of Agricultural Research New Series. 23(4): 642-645.
- Kaur, C., Selvakumar, G. and Ganeshamurthy, A. 2019. Acid Tolerant Microbial Inoculants: A Requisite for Successful Crop Production in Acidic Soils. In: Phyto and Rhizo Remediation. Berlin: Springer. pp 235-247.
- Keerthisinghe, G., Zapata, F., Chalk, P. and Hocking, P. 2001. Integrated approaches for improved P nutrition of plants in tropical acid Soils. In: Plant Nutrition Food Security and Sustainability of Agro-Ecosystems (eds. W. J. Horst, M. K. Schenk, A. Bürkert, N. Claaseen, H. Flessa, W. B. Frommer, H.,Goldbach, H. W. Olfs, V. Römheld, B. Sattelmacher, U. Schmidhalter, S. Schubert, N. V. Wirén and L. Wittenmayer). Dordrecht: Springer. pp 974–975.
- Khadka, D., Lamichhane, S. and Thapa, B. 2016. Assessment of relationship between soil pH and macronutrients, Western Nepal. *Journal of Chemical, Biological and Physical Sciences.* **6**(2): 303-311.

- Khan, A., Munsif, F., Akhtar, K., Afridi, M. Z., Ahmad, Z., Fahad, S., Ullah, R., Khan, F. A. and Din, M. 2014. Response of fodder maize to various levels of nitrogen and phosphorus. *American Journal of Plant Sciences*. **5**: 2323-2329.
- Khan, M. A., Abid, M., Hussain, N. and Massood, M. U. 2005. Effect of phosphorous levels on growth and yield of maize (*Zea mays* L.) cultivars under saline conditions. *International Journal of Agriculture and Biology*. **3**: 511-514.
- Khan, M. A., Aslam, M., Tariq-Sultan and Mahmood, I. A. 2002. Response of phosphorus application on growth and yield of inoculated and un-inoculated mungbean (*Vigna radiata*). *International Journal of Agriculture and Biology*. **4**(4): 523-524.
- Khan, W., Singh, V., Sagar, A. and Singh, S. N. 2017. Response of phosphorus application on growth and yield attributes of sweet corn (*Zea mays* L. saccharata) varieties. *Journal of Pharmacognosy and Phytochemistry*. **6**(5): 2144-2146.
- Khurana, M. P. S. and Arora, S. 2012. Comparative efficiency of borax and granubor as boron fertilizers for lentil and soybean grown on alluvial alkaline soils. *Journal of Plant Nutrition.* **35**: 2145-2155.
- Kifuko, M. N., Othieno, C. O., Okalebo, J. R., Kimenye, L. N., Ndungu, K. W. and Kipkoech, A. K. 2007. Effect of combining organic residues with Mijingu phosphate rock on sorption and availability of phosphorus and maize production in acid soils of western Kenya. *Experimental Agriculture*. **43**: 51-66.
- Kisinyo, P. O. 2016. Effect of lime and phosphorus fertilizer on soil chemistry and maize seedlings performance on Kenyan acid soils. *Sky Journal of Agricultural Research*. 5(6): 97-104.
- Kisinyo, P. O., Othieno, C. O., Gudu, S. O., Okalebo, J. R., Opala, P. A., Maghanga, J. K., Ng'etich,W. K., Agalo, J. J., Opile, R. W. and Kisinyo, J. A. 2013. Phosphorus sorption and lime requirements of maize growing acids soil of Kenya. *Sustainable Agriculture Research.* 2: 116-123.
- Kisinyo, P. O., Othieno, C. O., Gudu, S. O., Okalebo, J. R., Opala, P. A., Ng'etich, W. K., Nyambati, R .O., Ouma, E. O., Agalo, J. J., Kebeney, S. J., Too, E .J., Kisinyo, J. A. and Opile, W. R. 2014. Immediate and residual effects of lime and phosphorus fertilizer on soil acidity and maize production in Western Kenya. *Experimental Agriculture*. **50**(1): 128-143.
- Kovacevic, V. and Rastija, M. 2010. Impacts of liming by dolomite on the maize and barley grain yields. *Poljoprivreda*. **16**(2): 3-8.
- Kumar, R., Chatterjee, D., Kumawat, N., Pandey, A., Roy, A. and Kumar, M. 2014. Productivity, quality and soil health as influenced by lime in ricebean cultivars in foothills of Northeastern India. *The Crop Journal.* **2**: 338-344.

- Kumar, M., Khan, M. H., Singh, P., Ngachan, S. V., Rajkhowa, D. J., Kumar, A. and Devi, M. H. 2012. Variable lime requirement based on differences in organic matter content of Iso-acidic soils. *Indian Journal of Hill Farming*. 25(1):26-30.
- Kumar, M., Kumar, R., Singh, B. P., and Jha, A. K. 2010. Potassium release pattern in a long term cropped and fertilized soils. *Environment and Ecology*. **28**: 1190-1192.
- Kumar, M., Singh, M. and Kumar, S. 2006. Effect of nitrogen and phosphorus levels on yield, response curve, partial factor productivity, nutrient use efficiency and economics in maize (*Zea mays* L) under rainfed condition of Nagaland. *Indian Journal of Hill Farming.* **19**(1&2): 118-122.
- Kumar, P., Pandey, S. K. and Kumar, P. 2015. Effect of different phosphorus levels on nutrient content, uptake and economics of urd bean under custard apple based agrihorti system. *Journal of AgriSearch*. **2**(2): 88-93.
- Kumar, R. 2015. Influence of mulching, liming and farm yard manures on production potential, economics and quality of maize (*Zea mays* L.) under rainfed condition of Eastern Himalaya. *Bangladesh Journal of Botany*. **44**(3): 391-398.
- Kumar, R., Ghanshyam, Shambhabhi, S., Kumar, S. and Kumari, S. 2020. Effect of different sources of boron and it's doses on physico-chemical properties of soil and nutrients in soil. *International Journal of Chemical Studies*. **8**(1): 15-20.
- Kumar, S., Wani, J. A., Lone, B. A., Singh, P., Dar, Z. A., Qayoom, S. and Fayaz, A. 2017. Effect of different levels of phosphorus and sulphur on seed & stover yield of soybean (*Glycine max* L. Merill) under 'Eutrochrepts'. *Asian Research Journal of Agriculture*. 5(1): 1-7.
- Kundu, D., Khanam, R., Saha, S., Thingujam, U. and Hazra, G. C. 2017. Boron availability in relation to some important soil chemical properties in acid soils of Cooch Behar district, West Bengal. *Journal of Applied and Natural Science*. **9**(4): 2400-2403.
- Lauchli, A. and Grattan, S. R. 2012. Soil pH extremes. **In**: Plant Stress Physiology (ed. S. Shabala), Wallingford: Centre for Agriculture and Bioscience International. pp 194-209.
- Lege, B. N. 2012. Soil chemical properties as affected by phosphorus and nitrogen-based manure and compost application. *Soil Science Research*. **15**: 1-6.
- Ligeyo, D. O. and Gudu, S. O. 2005. Further Laboratory Screening of more Kenyan Maize inbred lines for tolerance to aluminium (Experiment 2). Third year progress report, March 2004 to February 2005. McKnight Foundation, U.S.A. Project.
- Ligeyo, D.O. 2007. Genetic analysis of maize (*Zea mays* L.) tolerance to aluminium toxicity and low phosphorus stress and development of synthetics for use in acid soils of western Kenya. PhD thesis, Moi University, Kenya.

- Lin, M. H., Gresshoff, P. M. and Ferguson, B. J. 2012. Systemic regulation of soybean nodulation by acidic growth conditions. *Plant Physiology*. **160**: 2028-2039.
- Lokya, T., Mishra, A., Shivhare, S., Ravinder, J., Mali, S. and Pilli, K. 2017. Influence of Integrated nutrient management practices on concentration, uptake and recovery of the macro primary nutrients in maize crop in acid soil. *Bulletin of Environment Pharmacology and Life Sciences*. Vol. 1 Special Issue. pp 319-322.
- Lynrah, A. and Nongmaithem, D. 2017. Effect of lime and integrated nutrient management on soybean under rainfed condition of Nagaland. *International Journal of Bioresource and Stress Management*. **8**(5): 679-683.
- Maize Outlook, January 2019. https://pjtsau.edu.in Accessed on 12 April 2020.
- Malama, C., Goma, H.C. and Manda, T. H. E. 2000. Acid soils and maize experiences from Zambia. Paper presented at Mini workshop. Developing a regional strategy of evaluating elite maize germplasm for tolerance to low soil pH. 17-20 July 2000. Potchetstroom. SA.
- Mallarino, A. P. 2011. Corn and soybean response to soil pH level and liming. **In**: Proceedings of the 23rd Annual Integrated Crop Management Conference, 93-102. DOI: 10.31274/icm-180809-74. Accessed on 1 March 2020.
- Mantovani, J.P.M., Calonego, J.C. and Foloni, J.S.S. 2013. Boron leaf application at different growth stages of peanut. *Revista Ceres*. **60**(2): 270-278.
- Mariano, E. D. and Keltjens, W. G. 2005. Long-term effects of aluminum exposure on nutrient uptake by maize genotypes differing in aluminum resistance. *Journal of Plant Nutrition*. **28**(2): 323-333.
- Marquez, W. and Baucas, M. 1990. Effect of liming on the yield and nutrient uptake of potato, carrot, cabbage, and garden pea. Rome, Italy: FAO. https://agris.fao.org. Accessed on 6 March 2020.
- Marschner, H. 1995. **In**: Mineral nutrition of higher plants, 2nd Ed. London, Academic Press. pp 889.
- Martens, D. C. and Westermann, D. T. 1991. Fertilizer applications for correcting micronutrients deficiencies. **In**: Micronutrients in agriculture (ed: J. J. Mortvedt), 2nd edition, Soil Science Society of America, Madison. pp 549-592.
- Masood, T., Gul, R., Munsif, F., Jalal, F., Hussain, Z., Noreen, N., Khan, H., Nasiruddin and Khan, H. 2011. Effect of different phosphorus levels on the yield and yield components of maize. *Sarhad Journal of Agriculture*. **27** (2): 167-170.

- Matsumoto, H. 2000. Cell biology of aluminum toxicity and tolerance in higher plants. *International Review of Cytology*. **200**: 1-46.
- Meena, R. S. and Varma, D. 2016. Mungbean yield and nutrient uptake performance in response of NPK and lime levels under acid soil in Vindhyan region, India. *Journal of Applied and Natural Science*. **8**(2): 860 863.
- Mehta, Y. K., Shaktawat, M. S. and Singh, S. M. 2005. Influence of sulphur, phosphorus and farmyard manure on yield attributes and yield of maize (*Zea mays* L.) in southern Rajasthan conditions. *Indian Journal of Agronomy*. **50**: 203-205.
- Mendes, A. P., Farina, M. P. W. and Channon, P. 1985. A field evaluation of the differential tolerance to soil acidity of forty-eight South African maize cultivars. *South African Journal of Plant and Soil.* 2(4): 215-220.
- Mikko, S. 1972. Review on individual trace elements. In: Trace Elements in Soils and Agriculture. Rome: Food and Agriculture Organization of the United Nations, Rome. pp. 15–20.
- Mohanty, S. K. and Swain, M. R. 2018. Bioethanol production from corn and wheat: food, fuel, and future. **In:** Bioethanol production from food crops. Elsevier. pp 45-49.
- Montgomery, E. G. 1911. Correlation studies in corn. Nebraska Agricultural Experimental Station. Annual Report. 24: 108-159.
- Moreira, A. and Fageria, N. K. 2010. Liming influence on soil chemical properties, nutritional status and yield of alfalfa grown in acid soil. *Revista Brasileira de Ciencia do Solo*. **34**:1231-1239.
- Moroni, S., Zander, A., Condon, J. and Li, G. 2018. Quantifying the tolerance of cereal cultivars to soil acidity. Grains Research and Development Corporation, State of New South Wales through the Department of Industry, Skills and Regional Development. Issue 12: May 2018.
- Mugwira, L.M. 1980. Growth and Ca, Mg, K, and P uptake by triticale wheat, and rye at four Al levels. *Journal of Plant nutrition*. **5**: 591-606.
- Muhammad. S. H., Subbarayappa, C. T. and Sarala, K. J. 2015 Effect of graded levels of boron on growth and yield attributes of maize in calcareous soils. *Trends in Bioscience*. 8(13): 3291- 3296.
- Muhlbachova, G., Cermak, P., Vavera, R., Kas, M., Pechova, M., Markova, K., Kusa, H., Ruzek, P., Hlusek, J. and Losak, T. 2017. Boron availability and uptake under increasing phosphorus rates in a pot experiment. *Plant, Soil and Environment.* 63(11): 483-490.

- Muindi, E. M., Mrema, J., Semu, E., Mtakwa, P. and Gachene, C. 2015. Effects of Lime-Aluminium-Phosphate Interactions on Maize Growth and Yields in Acid Soils of the Kenya Highlands. *American Journal of Agriculture and Forestry*. **3**(6): 244-252.
- Nedunchezhiyan, M., Byju, G., Naskar, S. K. and Mukherjee, A. 2010. Effect of mulching and graded doses of fertilizer on yield and nutrient uptake of greater yam + maize intercropping system. *Indian Journal of Horticulture*. **67**: 283-287.
- Nekesa, A. O. 2007. Effects of Minjingu phosphate rock and agricultural lime in relation to maize, groundnut and soybean yield on acid soils of western Kenya. MSc thesis, Moi University, Kenya.
- Nsanzabaganwa, E., Das, T. K. and Rana, D. S. 2014. Nitrogen and phosphorus effects on the growth, phenology, heat and nutrients accumulation and yield of winter maize (*Zea mays*) in western Indo-Gangetic Plains. *Indian Journal of Agricultural Sciences*. 84(5): 661-664.
- Nwachukwu. 2002. A survey of available boron in some soils of rain forest and Guinea Savanna Zones of Nigeria. *Journal of the Sustainable Agriculture and Environment*. 4: 97-101.
- Okalebo, J. R., Gathua, K. W. and Woomer, P. L. 2002. Laboratory methods of soil analysis: A working manual, 2nd Ed. TSBR-CIAT and SACRED Africa, Nairobi, Kenya. pp-88.
- Opala, P. A. 2017. Influence of lime and phosphorus application rates on growth of maize in an acid soil. 2017. *Hindawi Advances in Agriculture*. Article ID 7083206. https://doi.org/10.1155/2017/7083206. Accessed on 12 April 2020.
- Opala, P. A., Odendo, M. and Muyekho, F. N. 2018. Effects of lime and fertilizer on soil properties and maize yields in acid soils of Western Kenya. *African Journal of Agricultural Research.* **13**(13): 657-663.
- Ossom, E. M. and Rhykerd, R. L. 2008. Effect of lime on soil and tuber chemical properties and yield of sweet potato in Swaziland. *American-Eurasian Journal of Agronomy*. 1: 1-5.
- Osundwa, M. A., Okalebo, J. R., Ngetich, W.K., Ochuodho, J. O., Othieno, C. O., Langat, B. and Omenyo, V. S. 2013. Influence of agricultural lime on soil properties and wheat (*Triticum aestivum* L.) yield on acidic soils of Uasin Gishu County, Kenya. *American Journal of Experimental Agriculture*. 3(4): 806-823.
- Otieno, H. M. O., Chemining'wa, G. N. and Zingore, S. 2018. Effect of farmyard manure, lime and inorganic fertilizer applications on soil pH, nutrients uptake, growth and nodulation of soybean in acid soils of Western Kenya. *Journal of Agricultural Science*. **10**(4): 199-206.

- Oya, K. and Khondaker, M. 1996. Effect of liming on the yield and nutrient uptake of common millet grown on a yellow acid soil of Okinawa. Science Bulletin, College of Agriculture, University of Ryukyus. 43: 81-88.
- Pal, B., Hirpara, D. S., Vora, V. D., Vekaria, P. D., Sutaria, G. S., Akbari, K. N. and Verma, H. P. 2017. Effect of nitrogen and phosphorus on yield and yield attributes of maize in South Saurashtra, India. *International Journal of Current Microbiology and Applied Sciences.* 6(3): 1945-1949.
- Pan, X., Al Baquy, M. A., Guan, P., Yan, J., Wang, R., Xu, R. and Xie, L. 2020. Effect of soil acidification on the growth and nitrogen use efficiency of maize in Ultisols. *Journal of Soils and Sediments*. 20: 1435-1445.
- Patil, Y. J., Patil, H. M., Bodake, P. S., Lende, N. S. and Patil, V. S. 2017. Effect of soil application of boron on growth, yield and soil properties of lowland paddy. *International Journal of Chemical Studies*. 5(5): 972-975.
- Patiram. 1991. Liming of acid soils and crop production in Sikkim. *Journal of Hill Research*. **4**: 6-12.
- Peoples, M. B., Herridge, D. F. and Ladha, J. K. 1995. Enhancing legume N fixation through plant and soil management. *Plant and Soil*. **174**: 83-101.
- Piper, C. S. 1966. Soil and Plant Analysis: a laboratory manual of methods for the examination of soils and the determination of the inorganic constituents of plants. Maver Publisher, Bombay, India. pp 368.
- Prajapati, J. P., Singh, R.P., Kumar, S., Kushwaha, J. K. and Yadav, P. K. 2013. Yield and nutrient uptake of mungbean [*Vigna radiata* (L.) Wilczek]. Agriculture for Sustainable Development. 1(1): 49-51.
- Prasad, R. 1992. Effect of liming on yield of soybean and nutrient availability in acid soil. *Journal of the Indian Society of Soil Science* **40**: 377-379.
- Prasad, R. 1998. A practical manual for soil fertility. Division of Agronomy, IARI, New Delhi.
- Quaggio, J. A., Ramos, V. J. and Furlani, P. 1991. Liming and molybdenum effects on nitrogen uptake and grain yield of corn. In: Plant-soil interactions at low pH (eds. R. J. Wright, V. C. Baligar, R. P. Murrnan), Dordrecht : Kluwer Academic. pp 327-332.
- Rahim, M., Ali, H. and Mahmood, T. 2004. Impact of nitrogen and boron application on growth and yield of maize (*Zea mays L.*) crop. *Journal of Research in Science*. 15: 153-157.
- Rahman, M. A., Chikushi, J., Duxbury, J. M., Meisner, C. A., Lauen, J. G. and Yasunaga, E. 2005. Chemical control of soil environment by lime and nutrients to improve the

productivity of acidic alluvial soils under rice wheat cropping system in Bangladesh. *Environmental Control in Biology*. **43**(4): 259-266.

- Rahman, M. S., Islam, M. T., Ishtiaque, S., Sarker, M. J. U. and Khan, A. S. M. M. R. 2018. Response of hybrid maize to boron application. *Bangladesh Journal of Agricultural Research*. 43(2): 281-288.
- Ranjit, R., Dasog, G. S. and Patil, P. L. 2007. Effect of lime and phosphorus levels on nutrient uptake by groundnut genotypes in acid soils of coastal ecosystem of Karnataka. *Karnataka Journal of Agricultural Sciences*. 20(3): 631-633.
- Rashid, M. and Iqbal, M. 2012. Effect of phosphorus fertilizer on the yield and quality of maize (Zea mays L) fodder on clay loam soil. The Journal of Animal & Plant Sciences. 22(1): 199-203.
- Rehim, A., Saleem, J., Bashir, M. A., Imran, M., Naveed, S., Sial, M. U. and Ahmed, F. 2018. Potassium and boron fertilization approaches to increase yield and nutritional attributes in maize crop. *Science, Technology and Development.* **37**(2): 69-77.
- Rekhi, R. S., Benbi, D. K. and Singh, B. 2000. Effect of fertilizers and organic manures on crop yields and soil properties in rice-wheat cropping system. *Rice Wheat Consortium Paper Series.* **6**:1-6.
- Rosolem, C. A. and Caires, E. F. 1998. Yield and nitrogen uptake of peanuts as affected by lime, cobalt and molybdenum. *Journal of Plant Nutrition.* **21**: 827-835.
- Sadiq, G., Khan, A. A., Inamullah, Rab, A., Fayyaz, H., Naz, G., Nawaz, H., Ali, I., Raza, H., Amin, J., Ali, S., Khan, H. A., Khan, A. A. and Khattak, W. A. 2017. Impact of phosphorus and potassium levels on yield and yield components of maize. *Pure and Applied Biology*. 6(3): 1071-1078.
- Saeed, M. T., Wahid, M. A., Saleem, M. F. and Aziz, T. 2017. Enhancing phosphorus use efficiency by supplementing through soil applications and seed phosphorus reserves in maize (*Zea mays*). *International Journal of Agriculture and Biology*. **19**(6): 1394-1400.
- Saha, A. R. and Haldar, M. 1998. Effect of phosphorus, lime and boron application on the changes in available B and P content of Aeric Haplaquept. *Journal of the Indian Society of Soil Science*. **46**(1): 22-26.
- Sahin, S. 2014. Effect of boron fertilizer applications on the growth and B, N uptake of maize (*Zea mays* L.) under the different soils. *Journal of Food, Agriculture & Environment*. 12(2): 1323-1327.
- Saleem, M., Gulaba. K., Gandahi, W., Bhattis, S. M. and Velo, S. 2016. Efficacy of colemmenite ore as boron fertilizer for maize (*Zea mays L.*) growth and yield. *Science International.* 28(3): 3071-3074.

- Saleem, M., Khanif, Y. M., Fauziah, I., Samsuri, A. W. and Hafeez, B. 2011. Importance of boron for agriculture productivity. *International Research Journal of Agricultural Science and Soil Science*. 1(8): 293-300.
- Salisbury, F. B. and Ross C. W. 1992. Plant Physiology, 4th Ed. Wadsworth Publishing Company, USA.
- Samant, T. K. 2015. Effect of lime and sowing date on soil properties with yield, quality and nutrient uptake by sunflower (*Helianthus annus* L.). *Journal of Global Sciences*. 4: 1830-1835.
- Sarkar, D., Ghosh, S., Batabyal, K., Mandal, B. and Chattopadhyay, A. P. 2015. Liming effects on extractable boron in six acidic soils. *Communications in Soil Science and Plant Analysis*. **46**: 1320-1325.
- Sarkar, G. K. 2009. Effect of liming on the forms and quantity/intensity (Q/I) parameters of potassium in some selected acidic soils of West Bengal. *Journal of Interacademicia*. 13: 418-29.
- Sarma, N. N., Paul, S. R. and Sarma, D. 2000. Response of maize to nitrogen and phosphorus under rainfed conditions of the hills zone of Assam. *Indian Journal of Agronomy*. 45(1): 128-131.
- Scott, B. J., Fisher, J. A. and Cullis, B. R. 2001. Aluminium tolerance and lime increase wheat yield on the acidic soils of central and southern New South Wales. *Australian Journal of Experimental Agriculture*. **41**: 523–532.
- Sentimenla, Singh, A. K., Singh, M. and Gupta, R. C. 2013. Effect of phosphorus and boron on production potential, profitability and efficiency indices of soybean. *Soybean Research.* 11(2):79-85.
- Sepat, S. and Rai, R. K. 2013. Effect of phosphorus levels and sources on productivity, nutrient uptake and soil fertility of maize-wheat cropping system. *Indian Journal of Agronomy*. **58**(3): 292-297.
- Sethi, D. 2015. Effect of LD slag as soil ameliorant and its impact on native PSB population in acid Alfisols of Bhubaneswar. M.Sc. (Ag) Thesis, Department of Soil Science and Agricultural Chemistry, Odisha University of Agriculture and Technology, Bhubaneswar.
- Shankarlingappa, B. C., Shivraj, B. and Vishwanath, K. P. 2000. Interaction effect of phosphorus and sulphur on nutrient uptake of nitrogen, phosphorus, potassium and sulphur by cowpea. *Karnataka Journal of Agricultural Sciences*. **13**(2): 295-298.
- Sharma, A., Rawat, U. S. and Yadav, B. K. 2011. Influence of phosphorus levels and phosphorus solubilizing fungi on yield and nutrient uptake by wheat under sub-humid

region of Rajasthan, India. *International Scholarly Research Network Agronomy*. https://doi.org/10.5402/2012/234656. Accessed on 12 April 2020.

- Sharma, P. P., Pandey, G., Jawahar, S., Kalaiyarasan, C. and Suseendran, K. 2018. Effect of different levels of nitrogen and phosphorus on yield and economics of hybrid maize (*Zea mays* L.). *International Journal of Research and Analytical Reviews*. 5(4): 527-529.
- Sharma, R., Dahiya, S. S., Singh, M., Malik, R. K. and Singh, D. 2008. Effect of sulphur and phosphorus interactions on growth and nutrient content in green gram. *Haryana Agricultural University Journal of Research*. **38**(1/2): 41-7.
- Sharma, U. C. and Singh, R. P. 2002. Acid soils of India: their distribution, management and future strategies for higher productivity. *Fertilizer News*. **47**: 45-52.
- Shelp, B. J. 1993. Physiology and biochemistry of boron in plants. **In**: Boron and Its Role in Production (ed. U. C. Gupta), CRC Press, Boca Raton, Florida. pp 53-58.
- Shen, Y., Lin, H., Song, X., Liu, P., Bo, L., Chen, J. and Yang, L. 2019. Short-term effects of byproduct amendments and lime on physicochemical and microbiological properties of acidic soil from Jiaodong Peninsula of China. *Ciência Rural.* **49**(6). Scielo. http://dx.doi.org/10.1590/0103-8478cr20180098. Accessed on 9 May 2020.
- Shoemaker, H. E., Mclean, E. O. and Pratt, P. F. 1961. Buffer methods for determining lime requirement of soils with appreciable amounts of extractable aluminium. *Soil Science Society of America Journal.* 25(4): 274-277.
- Shorrocks, V. M. 1997. The occurrence and correction of boron deficiency. *Plant and Soil*. **193**: 121-148.
- Shorrocks V. M. and Blaza A. J. 1973. The boron nutrition of maize. Field Crops. 25: 25-27.
- Shrestha, S., Becker, M., Lamers, J. P. A. and Wimme, M. A. 2020. Boron and zinc fertilizer applications are essential in emerging vegetable-based crop rotations in Nepal. *Journal of Plant Nutrition and Soil Science*. 000: 1-16. https://doi.org/10.1002/jpln.202000151. Accessed on 12 April 2020.
- Sierra, J., Noel, C., Dufour, L., Ozier-Lafontaine, H., Welcker, C. and Desfontaines, L. 2003. Mineral nutrition and growth of tropical maize as affected by soil acidity. *Plant and Soil.* **252**: 215-226.
- Singh, S. K., Kumari, N., Singh, C. S., Karmakar, S. and Puran, A. N. 2016. Performance of hybrid pigeonpea (*Cajanus Cajan* L.) as influenced by nutrient and lime levels under rainfed condition of Jharkhand. *The Bioscan.* 11(1): 327-330.
- Singh, A. K., Sarkar, A. K., Kumar, A. and Singh, B. P. 2009. Effect of long term use of mineral fertilizers, lime and farmyard manure on the crop yield, available plant

nutrient and heavy metal status in an acidic loam soil. *Journal of the Indian Society of Soil Science*. **57**(3): 362-365.

- Singh, A. K. and Singh, R. S. 2012. Effect of phosphorus and bioinoculants on yield, nutrient uptake and economics of long duration pigeonpea (*Cajanus cajan*). *Indian Journal of Agronomy*. 57(3): 265-269.
- Singh, D. K., Singh, A. K., Singh, M., Bordoloi, L. J. and Srivastava, O. P. 2012. Production potential and nutrient uptake efficiency of pea (*Pisum sativum L*) as influenced by different fertility levels and micronutrients. *Journal of the Indian Society of Soil Science*. 60(2): 150-155.
- Singh, M. V. 2004. Micronutrient deficiencies in Indian soils and field usable practices for their correction. IFA International Conference on Micronutrients, Feb. 23-24, 2004, New Delhi.
- Singh, R. N., Kumar, B., Prasad, N. K. and Singh, S. 2002. Effect of lime and boron application to maize-gram sequence in soils of Jharkhand. *Indian Journal of Agricultural Sciences*. **72**(6): 346-347.
- Snehlata. 2015. Performance of quality protein maize (*Zea mays* L.) varieties at varying fertility levels. M.Sc. (Ag) Thesis, Department of Agronomy, Maharana Pratap University of Agriculture and Technology, Udaipur.
- Soomro, Z. H., Baloch, P. A. and Gandhai, A. W. 2011. Comparative effects of foliar and soil applied boron on growth and fodder yield of maize. *Pakistan Journal of Agriculture, Agricultural Engineering and Veterinary Sciences*. 27(1): 18-26.
- Soylu, S., Topal, A., Sade, B., Akgun, N., Gezgin, S. and Babaoglu, M. 2004. Yield and yield attributes of durum wheat genotypes as affected by boron application in boron deficient calcareous soils: an evaluation of major Turkish genotypes for boron efficiency. *Journal of Plant Nutrition.* 27(6): 1077-1106.
- Statistical Handbook of Nagaland, 2017. Directorate of Economics and Statistics, Government of Nagaland.
- Stol, R. J., Van Helden, A. K. and Bruyn, P. L. 1976. Hydrolysis precipitation studies of aluminium (III) solutions. 2. A kinetic study and model. *Journal of Colloid Interface Science*. 57: 115-131.
- Su, C., Evans, L. J., Bates, T. E. and Spiers, G. A. 1994. Extractable soil boron and alfalfa uptake - calcium-carbonate effects on acid soil. *Soil Science Society* of *America Journal.* 58: 1145-1150.
- Subbiah, B. V. and Asija, G. L. 1956. A rapid procedure for the determination of available nitrogen in soil. *Current Science*. **25**: 259–60.

- Sultana, B. S., Miah, M. M. H., Islam, M. R., Rahman, M. M., Sarker, B. C. and Zoha, M. S. 2009. Effect of liming on soil properties, yield and nutrient uptake by wheat. *Current World Environment*. 4(1): 39-47.
- Swift, M. J., Dvorak, K. A., Mulongoy, K., Musoka, M., Sanginga, N. and Tian, G. 1994. The role of soil organism in the sustainability of tropical cropping systems In: Soil science and sustainable land management in tropics. (eds. S. K. Syers and D. I. Rimmer), CAB International, Cambridge University Press, U.K. pp 155-170.
- Tahir, M., Ali, A., Khalid, F., Naeem, M., Fiaz, N. and Waseem, M. 2012. Effect of foliar applied boron application on growth, yield and quality of maize (*Zea mays L.*). *Pakistan Journal of Scientific and Industrial Research Series B: Biological Sciences*. 55(3): 117-121.
- Tandzi, N. L., Ngonkeu, E. L. M., Youmbi, E., Nartey, E., Yeboah, M., Gracen, V., Ngeve, J. and Mafouasson, H. A. 2015. Agronomic performance of maize hybrids under acid and control soil conditions. *International Journal of Agronomy and Agricultural Research.* 6(4): 275-291.
- Teshome, N. and Garadew, W. 2019. Effect of potassium fertilizer and lime application on yield and quality of soybean (*Glycine max* L. (merrill) in acidic soil of Gobu Sayo district, Western Oromia, Ethiopia. *Food Science and Quality Management*. DOI: 10.7176/FSQM Vol.90, 2019. Accessed on 14 February 2020.
- The, C., Calba, H., Zonkeng, C., Ngonkeu, E. L. M. and Adetimirin V. O. 2006. Response of maize grain yield to changes in acid soil characteristics after soil amendment. *Plant* and Soil. 284: 45-57.
- Timlin, D. J., Naidu, T. C. M., Fleisher, D. H. and Reddy, V. R. 2017. Quantitative effects of phosphorus on maize canopy photosynthesis and biomass. *Crop Science*. 57: 3156– 3169.
- Toppo, N. and Kumar, S. B. 2018. Effect of liming materials on available phosphorus and organic carbon in acid soil of Ranchi, Jharkhand. *International Journal of Current Microbiology and Applied Sciences*. Special Issue-7: 3442-3447.
- Trautmann, R. R., Lana, M. C., Guimaraes, V. F., Goncalves, A.C. and Steiner, F. 2014. Soil water potential and boron fertilization in growth and uptake of the nutrient for the soybean crop. *The Revista Brasileira de Ciencia do Solo.* **38**: 240-251.
- Umeri, C., Moseri, H. and Onyemekonwu, R. C. 2016. Effects of nitrogen and phosphorus on the growth performance of maize (*Zea mays*) in selected soils of Delta State, Nigeria. *Advances in Crop Science and Technology*. **4**(1): 207.
- Uygur, V. and Sen, M. 2018. The effect of phosphorus application on nutrient uptake and translocation in wheat cultivars. *International Journal of Agriculture, Forestry and Life Science*. **2**(2): 171-179.

- Venkatesh, M. S., Majumdar, B., Kumar, K. and Patiram. 2002. Effect of phosphorus, FYM and lime on yield, P uptake by maize and forms of soil acidity in Typic Hapludalf of Meghalaya. *Journal of the Indian Society of Soil Science*. **50** (3): 254-258.
- Verde, B. S. D., Oginga, B. and Njeri, M. J. 2013. Effects of manure, lime and mineral P fertilizer on soybean yields and soil fertility in humic Nitisol in the Central Highlands of Kenya. *International Journal of Agricultural Science Research.* 2: 283-291.
- Voor, A. S. 2012. Growth rate and response of bacteria communities to pH in limed and ash treated forest Alfisol. *Soil Microbiology*. **33**: 881-888.
- Wafula, W. N., Korir, N. K., Ojulong, H. F., Siambi, M. and Gweyi-Onyango, J. P. 2018. Protein, calcium, zinc, and iron contents of finger millet grain response to varietal differences and phosphorus application in Kenya. *Agronomy*. 8(24). https://doi.org/10.3390/agronomy8020024. Accessed on 14 February 2020.
- Walkey, A. and Black, I. A. 1934. An examination of the degtjareff method for determining soil organic carbon matter and a purposed modification of chromic acid titration method. *Soil Science*. **37**: 29-38.
- Weisz, R., Whit, J., Knox, B. and Reed, L. 2003. Long term-variable rate lime and P application for piedmont no-till field crops. *Precision Agriculture*. **4**: 311-330.
- Woodruff, J. R., Moore, F. W. and Musen, H. L. 1987. Potassium, boron, nitrogen and lime effects on corn yield and ear leaf nutrients concentration. *Agronomy Journal*. **79**: 520–524.
- Yadesa, W., Tadesse, A., Kibret, K. and Dechassa, N. 2019. Effect of liming and applied phosphorus on growth and P uptake of maize (*Zea mays* subsp.) plant grown in acid soils of West Wollega, Ethiopia. *Journal of Plant Nutrition*. 42(5): 477-490.
- Zesith, E. U. 2011. Fertility status of an Alfisol and maize yield as affected by lime rates and poultry manure application. *Soil and Land* Resources. **32**: 333-339.
- Zhihao, Y., Zhihua, H., Shichao, W., Shengchang, H., Hongliang, W., Jinyu, W., Tingting, X., Xichu, Y., Daming, L. and Changai, L. U. 2019. Effects of lime content on soil acidity, soil nutrients and crop growth in rice-rape rotation system. *Scientia Agricultura Sinica*. 52(23): 4285-4295.

APPENDIX

SOV		2016		0	2017			Pooled			Etab
501	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F tab
pН	1	108.29	108.29	12.41*	492.46	492.46	81.60*	600.75	300.37	40.70*	4.04
Lime (L)	3	1373.42	457.81	52.47*	853.01	284.34	47.12*	2226.44	371.07	50.28*	2.80
Variety(V)	2	316.80	158.40	18.15*	324.80	162.40	26.91*	641.60	160.40	21.73*	3.19
pH x L	3	84.27	28.09	3.22*	84.88	28.29	4.69*	169.14	28.19	3.82*	2.80
L x V	6	6.75	1.12	0.13	15.62	2.60	0.43	22.37	1.86	0.25	2.29
pH x V	2	2.76	1.38	0.16	1.55	0.78	0.13	4.31	1.08	0.15	3.19
pHxLxV	6	11.30	1.88	0.22	8.34	1.39	0.23	19.64	1.64	0.22	2.29
Error	48	418.81	8.73		289.67	6.03		708.48	7.38		
TSS	71	2322.40	32.71		2070.33	29.16		4393.18	30.72		
Appendix II: Effect of pH, lime and varieties on plant height of maize at 60 DAS											
SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	4180.03	4180.03	205.69*	4278.12	4278.12	178.10*	8458.15	4229.08	190.74*	4.04
Lime (L)	3	13938.48	4646.16	228.63*	14053.14	4684.38	195.01*	27991.62	4665.27	210.42*	2.80
Variety(V)	2	975.35	487.68	24.00*	1016.05	508.03	21.15*	1991.40	497.85	22.45*	3.19
pH x L	3	44.97	14.99	0.74	156.58	52.19	2.17	201.55	33.59	1.52	2.80
L x V	6	64.07	10.68	0.53	129.64	21.61	0.90	193.72	16.14	0.73	2.29
pH x V	2	3.94	1.97	0.10	145.76	72.88	3.03	149.69	37.42	1.69	3.19
pHxLxV	6	20.46	3.41	0.17	33.87	5.64	0.23	54.33	4.53	0.20	2.29
Error	48	975.45	20.32		1153.02	24.02		2128.47	22.17		
TSS	71	20202.75	284.55		20966.18	295.30		42626.01	298.08		
Appendix III:	Effect of pl	H, lime and var	rieties on pla	nt height of 1	naize at harv	rest					
SOV	DE	2016			2017		_	Pooled	-		Etab
301	DI	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	1 tab
pН	1	295.65	295.65	12.71*	509.87	509.87	20.97*	805.52	402.76	16.93*	4.04
Lime (L)	3	19551.42	6517.14	280.23*	19910.52	6636.84	272.94*	39461.93	6576.99	276.50*	2.80
Variety(V)	2	1452.00	726.00	31.22*	1117.16	558.58	22.97*	2569.16	642.29	27.00*	3.19
pH x L	3	139.12	46.37	1.99	166.21	55.40	2.28	305.33	50.89	2.14	2.80
L x V	6	32.79	5.47	0.24	54.19	9.03	0.37	86.99	7.25	0.30	2.29
pH x V	2	77.11	38.56	1.66	39.49	19.74	0.81	116.60	29.15	1.23	3.19
pHxLxV	6	42.97	7.16	0.31	108.70	18.12	0.75	151.68	12.64	0.53	2.29
Error	48	1116.31	23.26		1167.19	24.32		2283.51	23.79		
TSS	71	22707.38	319.82		23073.33	324.98		48058.39	336.07		

Appendix I: Effect of pH, lime and varieties on plant height of maize at 30 DAS

* Significant at 5% level

SOV	DE	2016			2017			Pooled	Etab		
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
pН	1	0.13	0.13	0.60	0.35	0.35	1.19	0.47	0.24	0.94	4.04
Lime (L)	3	13.49	4.50	21.58*	5.26	1.75	6.02*	18.75	3.13	12.50*	2.80
Variety(V)	2	4.08	2.04	9.80*	3.03	1.51	5.19*	7.11	1.78	7.11*	3.19
pH x L	3	0.38	0.13	0.60	0.82	0.27	0.94	1.19	0.20	0.80	2.80
L x V	6	0.81	0.13	0.64	0.19	0.03	0.11	1.00	0.08	0.33	2.29
pH x V	2	0.75	0.38	1.80	0.36	0.18	0.62	1.11	0.28	1.11	3.19
pHxLxV	6	1.25	0.21	1.00	2.64	0.44	1.51	3.89	0.32	1.30	2.29
Error	48	10.00	0.21		14.00	0.29		24.00	0.25		
TSS	71	30.88	0.43		26.65	0.38		59.31	0.84		

Appendix IV: Effect of pH, lime and varieties on number of leaves of maize at 30 DAS

Appendix V: Effect of pH, lime and varieties on number of leaves of maize at 60 DAS

SOV	DE	2016			2017			Pooled			E tob
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	0.50	0.50	2.40	0.89	0.89	3.05	1.39	0.69	2.78	4.04
Lime (L)	3	10.50	3.50	16.80*	6.11	2.04	6.98*	16.61	2.77	11.07*	2.80
Variety(V)	2	7.53	3.76	18.07*	6.03	3.01	10.33*	13.56	3.39	13.56*	3.19
pH x L	3	0.94	0.31	1.51	0.11	0.04	0.13	1.06	0.18	0.70	2.80
L x V	6	0.25	0.04	0.20	0.31	0.05	0.17	0.56	0.05	0.19	2.29
pH x V	2	0.08	0.04	0.20	0.53	0.26	0.90	0.61	0.15	0.61	3.19
pHxLxV	6	0.14	0.02	0.11	0.47	0.08	0.27	0.61	0.05	0.20	2.29
Error	48	10.00	0.21		14.00	0.29		24.00	0.25		
TSS	71	29.94	0.42		28.44	0.40		60.64	0.85		

Appendix VI: Effect of pH, lime and varieties on number of leaves of maize at harvest

SOV	DE	2016			2017			Pooled			Etab
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
pН	1	0.89	0.89	3.05	0.68	0.68	3.77	1.57	0.78	3.32	4.04
Lime (L)	3	5.00	1.67	5.71*	4.04	1.35	7.46*	9.04	1.51	6.38*	2.80
Variety(V)	2	4.11	2.06	7.05*	1.36	0.68	3.77*	5.47	1.37	5.79*	3.19
pH x L	3	1.67	0.56	1.90	0.04	0.01	0.08	1.71	0.28	1.21	2.80
L x V	6	2.67	0.44	1.52	2.08	0.35	1.92	4.75	0.40	1.68	2.29
pH x V	2	0.44	0.22	0.76	0.86	0.43	2.38	1.31	0.33	1.38	3.19
pHxLxV	6	0.33	0.06	0.19	1.92	0.32	1.77	2.25	0.19	0.79	2.29
Error	48	14.00	0.29		8.67	0.18		22.67	0.24		
TSS	71	29.11	0.41		19.65	0.28		52.44	0.74		

* Significant at 5% level

SOV	DE	2016			2017			Pooled	Etab		
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pH	1	0.00	0.00	0.44	0.04	0.04	2.29	0.04	0.02	1.70	4.04
Lime (L)	3	0.05	0.02	2.14	0.06	0.02	1.14	0.11	0.02	1.47	2.80
Variety(V)	2	0.02	0.01	1.14	0.05	0.03	1.65	0.07	0.02	1.48	3.19
pH x L	3	0.01	0.00	0.40	0.00	0.00	0.01	0.01	0.00	0.14	2.80
L x V	6	0.01	0.00	0.21	0.00	0.00	0.01	0.01	0.00	0.08	2.29
pH x V	2	0.00	0.00	0.12	0.01	0.01	0.32	0.01	0.00	0.25	3.19
pHxLxV	6	0.02	0.00	0.45	0.01	0.00	0.09	0.03	0.00	0.20	2.29
Error	48	0.38	0.01		0.79	0.02		1.17	0.01		
TSS	71	0.49	0.0069		0.96	0.014		1.53	0.022		

Appendix VII: Effect of pH, lime and varieties on leaf area index of maize at 30 DAS

Appendix VIII: Effect of pH, lime and varieties on leaf area index of maize at 60 DAS

SOV	DE	2016			2017			Pooled	Etab		
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F tao
pН	1	0.01	0.01	1.83	0.00	0.00	0.04	0.01	0.00	0.87	4.04
Lime (L)	3	0.04	0.01	2.83*	0.05	0.02	3.11*	0.08	0.01	2.98*	2.80
Variety(V)	2	0.01	0.01	1.39	0.02	0.01	1.88	0.03	0.01	1.65	3.19
pH x L	3	0.01	0.00	0.47	0.00	0.00	0.31	0.01	0.00	0.38	2.80
L x V	6	0.02	0.00	0.69	0.01	0.00	0.29	0.03	0.00	0.48	2.29
pH x V	2	0.00	0.00	0.25	0.01	0.01	1.04	0.01	0.00	0.67	3.19
pHxLxV	6	0.02	0.00	0.65	0.02	0.00	0.54	0.03	0.00	0.59	2.29
Error	48	0.21	0.00		0.24	0.01		0.45	0.00		
TSS	71	0.31	0.0044		0.35	0.0049		0.78	0.011		

Appendix IX: Effect of	pH. li	me and	varieties on	leaf are	a index	of maize	at harvest

SOV	DE	2016			2017			Pooled	Etab		
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	0.00	0.00	0.70	0.00	0.00	0.01	0.00	0.00	0.46	4.04
Lime (L)	3	0.04	0.01	3.44*	0.03	0.01	3.92*	0.07	0.01	3.61*	2.80
Variety(V)	2	0.00	0.00	0.07	0.01	0.00	1.59	0.01	0.00	0.61	3.19
pH x L	3	0.00	0.00	0.26	0.00	0.00	0.31	0.01	0.00	0.28	2.80
L x V	6	0.00	0.00	0.09	0.00	0.00	0.19	0.00	0.00	0.13	2.29
pH x V	2	0.01	0.00	1.10	0.00	0.00	0.15	0.01	0.00	0.76	3.19
pHxLxV	6	0.00	0.00	0.07	0.00	0.00	0.13	0.00	0.00	0.09	2.29
Error	48	0.20	0.00		0.11	0.00		0.31	0.00		
TSS	71	0.27	0.0038		0.15	0.0021		0.45	0.0063		

* Significant at 5% level
| SOV | DE | 2016 | 2016 | | | | | Pooled | | | Etab |
|------------|----|-------|------|--------|-------|------|--------|--------|------|--------|-------|
| 301 | DF | SS | MSS | F cal | SS | MSS | F cal | SS | MSS | F cal | r lau |
| pH | 1 | 3.65 | 3.65 | 34.89* | 3.08 | 3.08 | 35.47* | 6.73 | 3.36 | 35.15* | 4.04 |
| Lime (L) | 3 | 8.64 | 2.88 | 27.50* | 5.51 | 1.84 | 21.18* | 14.15 | 2.36 | 24.64* | 2.80 |
| Variety(V) | 2 | 1.80 | 0.90 | 8.60* | 0.70 | 0.35 | 4.03* | 2.50 | 0.62 | 6.53* | 3.19 |
| pH x L | 3 | 0.82 | 0.27 | 2.61 | 0.33 | 0.11 | 1.26 | 1.15 | 0.19 | 2.00 | 2.80 |
| L x V | 6 | 0.42 | 0.07 | 0.67 | 0.25 | 0.04 | 0.49 | 0.68 | 0.06 | 0.59 | 2.29 |
| pH x V | 2 | 0.26 | 0.13 | 1.22 | 0.02 | 0.01 | 0.10 | 0.27 | 0.07 | 0.71 | 3.19 |
| pHxLxV | 6 | 0.23 | 0.04 | 0.36 | 0.29 | 0.05 | 0.56 | 0.52 | 0.04 | 0.45 | 2.29 |
| Error | 48 | 5.03 | 0.10 | | 4.16 | 0.09 | | 9.19 | 0.10 | | |
| TSS | 71 | 20.85 | 0.29 | | 14.33 | 0.20 | | 39.14 | 0.27 | | |

Appendix X: Effect of pH, lime and varieties on cob length of maize

Appendix XI: Effect of pH, lime and varieties on cob girth of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
pН	1	2.38	2.38	11.67*	1.93	1.93	17.30*	4.31	2.16	13.66*	4.04
Lime (L)	3	4.22	1.41	6.89*	3.98	1.33	11.90*	8.20	1.37	8.66*	2.80
Variety(V)	2	1.36	0.68	3.32*	1.24	0.62	5.57*	2.60	0.65	4.11*	3.19
pH x L	3	0.59	0.20	0.96	0.54	0.18	1.61	1.13	0.19	1.19	2.80
L x V	6	0.18	0.03	0.15	0.62	0.10	0.93	0.80	0.07	0.42	2.29
pH x V	2	0.03	0.02	0.08	0.39	0.19	1.74	0.42	0.11	0.67	3.19
pHxLxV	6	0.43	0.07	0.35	0.77	0.13	1.15	1.20	0.10	0.64	2.29
Error	48	9.80	0.20		5.35	0.11		15.15	0.16		
TSS	71	19.00	0.27		14.81	0.21		33.92	0.24		

Appendix XII: Effect of pH, lime and varieties on cob weight of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
pН	1	1005.31	1005.31	75.45*	1146.49	1146.49*	62.16	2151.80	1075.90*	67.74	4.04
Lime (L)	3	5079.58	1693.19	127.07*	5248.03	1749.34*	94.85	10327.61	1721.27*	108.37	2.80
Variety(V)	2	529.34	264.67	19.86*	477.80	238.90*	12.95	1007.14	251.79*	15.85	3.19
pH x L	3	7.10	2.37	0.18	1.42	0.47	0.03	8.51	1.42	0.09	2.80
L x V	6	12.82	2.14	0.16	8.63	1.44	0.08	21.46	1.79	0.11	2.29
pH x V	2	1.56	0.78	0.06	0.25	0.12	0.01	1.81	0.45	0.03	3.19
pHxLxV	6	6.55	1.09	0.08	12.13	2.02	0.11	18.68	1.56	0.10	2.29
Error	48	639.57	13.32		885.28	18.44		1524.85	15.88		
TSS	71	7281.84	102.56		7780.02	109.58		15134.81	105.84		

* Significant at 5% level

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
pH	1	102.72	102.72	308.17*	105.13	105.13	189.23*	207.85	103.92	233.83*	4.04
Lime (L)	3	29.00	9.67	29.00*	32.38	10.79	19.43*	61.38	10.23	23.02*	2.80
Variety(V)	2	13.36	6.68	20.04*	20.08	10.04	18.08*	33.44	8.36	18.81*	3.19
pH x L	3	1.17	0.39	1.17	2.60	0.87	1.56	3.76	0.63	1.41	2.80
L x V	6	1.42	0.24	0.71	0.58	0.10	0.17	2.00	0.17	0.37	2.29
pH x V	2	4.36	2.18	6.54*	1.75	0.88	1.58	6.11	1.53	3.44*	3.19
pHxLxV	6	1.75	0.29	0.88	2.69	0.45	0.81	4.44	0.37	0.83	2.29
Error	48	16.00	0.33		26.67	0.56		42.67	0.44		
TSS	71	169.78	2.39		191.88	2.70		361.83	2.53		

Appendix XIII: Effect of pH, lime and varieties on number of grains per row of maize

Appendix XIV: Effect of pH, lime and varieties on number of rows per cob of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lao
pН	1	0.35	0.35	1.39	0.06	0.06	0.19	0.40	0.20	0.74	4.04
Lime (L)	3	15.15	5.05	20.20*	22.28	7.43	25.46*	37.43	6.24	23.03*	2.80
Variety(V)	2	4.33	2.17	8.67*	4.69	2.35	8.05*	9.03	2.26	8.33*	3.19
pH x L	3	3.60	1.20	4.80*	4.06	1.35	4.63*	7.65	1.28	4.71*	2.80
L x V	6	0.56	0.09	0.37	0.31	0.05	0.17	0.86	0.07	0.26	2.29
pH x V	2	0.44	0.22	0.89	0.36	0.18	0.62	0.81	0.20	0.74	3.19
pHxLxV	6	0.44	0.07	0.30	0.86	0.14	0.49	1.31	0.11	0.40	2.29
Error	48	12.00	0.25		14.00	0.29		26.00	0.27		
TSS	71	36.88	0.52		46.61	0.66		85.49	0.60		

Appendix XV: Effect of pH, lime and varieties on number of grains per cob of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	29484.01	29484.01	41.88*	34892.01	34892.01	55.12*	64376.03	32188.01	48.15*	4.04
Lime (L)	3	57159.15	19053.05	27.06*	59965.82	19988.61	31.58*	117124.97	19520.83	29.20*	2.80
Variety(V)	2	12826.69	6413.35	9.11*	11998.58	5999.29	9.48*	24825.28	6206.32	9.28*	3.19
pH x L	3	1650.71	550.24	0.78	2585.82	861.94	1.36	4236.53	706.09	1.06	2.80
L x V	6	1025.31	170.88	0.24	3074.64	512.44	0.81	4099.94	341.66	0.51	2.29
pH x V	2	1155.19	577.60	0.82	1377.19	688.60	1.09	2532.39	633.10	0.95	3.19
pHxLxV	6	1269.25	211.54	0.30	2472.47	412.08	0.65	3741.72	311.81	0.47	2.29
Error	48	33791.33	703.99		30383.33	632.99		64174.67	668.49		
TSS	71	138361.65	1948.76		146749.88	2066.90		288379.56	2016.64		

* Significant at 5% level

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
pH	1	6.31	6.31	19.89*	7.32	7.32	8.86*	13.63	6.82	11.92*	4.04
Lime (L)	3	30.28	10.09	31.79*	26.26	8.75	10.59*	56.54	9.42	16.47*	2.80
Variety(V)	2	1.17	0.59	1.85	1.19	0.60	0.72	2.36	0.59	1.03	3.19
pH x L	3	0.58	0.19	0.61	0.81	0.27	0.33	1.39	0.23	0.41	2.80
L x V	6	1.79	0.30	0.94	0.58	0.10	0.12	2.37	0.20	0.35	2.29
pH x V	2	0.73	0.37	1.16	0.10	0.05	0.06	0.84	0.21	0.37	3.19
pHxLxV	6	3.04	0.51	1.60	0.44	0.07	0.09	3.48	0.29	0.51	2.29
Error	48	15.24	0.32		39.68	0.83		54.92	0.57		
TSS	71	59.14	0.83		76.40	1.08		136.23	0.95		

Appendix XVI: Effect of pH, lime and varieties on test weight of maize

Appendix XVII: Effect of pH, lime and varieties on grain yield of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	1059.61	1059.61	198.74*	1581.38	1581.38	304.87*	2640.99	1320.49	251.08*	4.04
Lime (L)	3	3200.22	1066.74	200.08*	3296.79	1098.93	211.86*	6497.01	1082.84	205.89*	2.80
Variety(V)	2	457.47	228.73	42.90*	397.63	198.82	38.33*	855.10	213.77	40.65*	3.19
pH x L	3	30.20	10.07	1.89	32.65	10.88	2.10	62.86	10.48	1.99	2.80
L x V	6	38.37	6.39	1.20	12.16	2.03	0.39	50.53	4.21	0.80	2.29
pH x V	2	3.67	1.83	0.34	9.51	4.75	0.92	13.18	3.29	0.63	3.19
pHxLxV	6	14.08	2.35	0.44	4.53	0.76	0.15	18.61	1.55	0.29	2.29
Error	48	255.92	5.33		248.98	5.19		504.90	5.26		
TSS	71	5059.53	71.26		5583.63	78.64		10652.68	74.49		

Appendix XVIII: Effect of pH, lime and varieties on stover yield of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	2322.21	2322.21	49.59*	2522.87	2522.87	56.30*	4845.08	2422.54	52.88*	4.04
Lime (L)	3	7554.97	2518.32	53.78*	7337.55	2445.85	54.59*	14892.52	2482.09	54.18*	2.80
Variety(V)	2	518.97	259.49	5.54*	727.10	363.55	8.11*	1246.07	311.52	6.80*	3.19
pH x L	3	3.26	1.09	0.02	3.83	1.28	0.03	7.09	1.18	0.03	2.80
L x V	6	9.45	1.58	0.03	6.63	1.11	0.02	16.09	1.34	0.03	2.29
pH x V	2	2.69	1.35	0.03	0.22	0.11	0.00	2.91	0.73	0.02	3.19
pHxLxV	6	11.76	1.96	0.04	1.24	0.21	0.00	13.00	1.08	0.02	2.29
Error	48	2247.59	46.82		2150.75	44.81		4398.33	45.82		
TSS	71	12670.90	178.46		12750.18	179.58		25696.92	179.70		

* Significant at 5% level

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pH	1	0.01	0.01	3.17	0.00	0.00	0.71	0.02	0.01	2.01	4.04
Lime (L)	3	0.73	0.24	56.57*	0.95	0.32	83.33*	1.68	0.28	69.14*	2.80
Variety(V)	2	0.03	0.01	2.99	0.02	0.01	2.29	0.04	0.01	2.66	3.19
pH x L	3	0.01	0.00	1.04	0.02	0.01	1.75	0.03	0.01	1.38	2.80
L x V	6	0.01	0.00	0.24	0.00	0.00	0.03	0.01	0.00	0.14	2.29
pH x V	2	0.00	0.00	0.19	0.00	0.00	0.11	0.00	0.00	0.16	3.19
pHxLxV	6	0.01	0.00	0.24	0.00	0.00	0.21	0.01	0.00	0.23	2.29
Error	48	0.21	0.00		0.18	0.00		0.39	0.00		
TSS	71	1.00	0.0141		1.18	0.0166		2.20	0.0154		

Appendix XIX: Effect of pH, lime and varieties on nitrogen content in grain

Appendix XX: Effect of pH, lime and varieties on nitrogen content in stover

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
pН	1	0.01	0.01	2.30	0.01	0.01	2.30	0.01	0.01	2.30	4.04
Lime (L)	3	0.20	0.07	29.99*	0.18	0.06	26.47*	0.38	0.06	28.23*	2.80
Variety(V)	2	0.01	0.00	1.68	0.01	0.00	1.68	0.02	0.00	1.68	3.19
pH x L	3	0.00	0.00	0.46	0.00	0.00	0.46	0.01	0.00	0.46	2.80
L x V	6	0.00	0.00	0.31	0.00	0.00	0.31	0.01	0.00	0.31	2.29
pH x V	2	0.00	0.00	0.57	0.00	0.00	0.57	0.01	0.00	0.57	3.19
pHxLxV	6	0.00	0.00	0.10	0.00	0.00	0.10	0.00	0.00	0.10	2.29
Error	48	0.11	0.00		0.11	0.00		0.22	0.00		
TSS	71	0.33	0.0046		0.31	0.0044		0.67	0.0047		

Appendix XXI: Effect of pH, lime and varieties on phosphorus content in grain

SOV	DE	2016	2016			2017					Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
pН	1	0.04	0.04	57.00*	0.05	0.05	48.27*	0.09	0.05	51.85*	4.04
Lime (L)	3	0.08	0.03	38.83*	0.16	0.05	52.70*	0.25	0.04	47.02*	2.80
Variety(V)	2	0.01	0.00	6.20*	0.03	0.02	14.68*	0.04	0.01	11.21*	3.19
pH x L	3	0.01	0.00	3.34*	0.01	0.00	2.87*	0.02	0.00	3.06*	2.80
L x V	6	0.00	0.00	0.08	0.00	0.00	0.11	0.00	0.00	0.10	2.29
pH x V	2	0.00	0.00	0.18	0.00	0.00	0.03	0.00	0.00	0.09	3.19
pHxLxV	6	0.00	0.00	0.10	0.00	0.00	0.02	0.00	0.00	0.05	2.29
Error	48	0.03	0.00		0.05	0.00		0.08	0.00		
TSS	71	0.17	0.0024		0.30	0.0042		0.53	0.0037		

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
pH	1	0.02	0.02	18.30*	0.01	0.01	6.62*	0.03	0.01	12.08*	4.04
Lime (L)	3	0.03	0.01	8.51*	0.02	0.01	4.18*	0.04	0.01	6.20*	2.80
Variety(V)	2	0.00	0.00	2.20	0.01	0.00	2.13	0.01	0.00	2.16	3.19
pH x L	3	0.00	0.00	0.38	0.00	0.00	0.42	0.00	0.00	0.40	2.80
L x V	6	0.00	0.00	0.08	0.00	0.00	0.02	0.00	0.00	0.05	2.29
pH x V	2	0.00	0.00	0.05	0.00	0.00	0.01	0.00	0.00	0.03	3.19
pHxLxV	6	0.00	0.00	0.09	0.00	0.00	0.10	0.00	0.00	0.09	2.29
Error	48	0.05	0.00		0.06	0.00		0.11	0.00		
TSS	71	0.11	0.0015		0.09	0.0013		0.20	0.0014		

Appendix XXII: Effect of pH, lime and varieties on phosphorus content in stover

Appendix XXIII: Effect of pH, lime and varieties on potassium content in grain

SOV	DE	2016	2016					Pooled			Etab
50V	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
pН	1	0.09	0.09	23.89*	0.09	0.09	27.04*	0.17	0.09	25.40*	4.04
Lime (L)	3	0.08	0.03	7.70*	0.09	0.03	8.67*	0.17	0.03	8.16*	2.80
Variety(V)	2	0.04	0.02	6.15*	0.04	0.02	5.83*	0.08	0.02	6.00*	3.19
pH x L	3	0.00	0.00	0.42	0.00	0.00	0.38	0.01	0.00	0.40	2.80
L x V	6	0.00	0.00	0.02	0.00	0.00	0.10	0.00	0.00	0.06	2.29
pH x V	2	0.00	0.00	0.15	0.00	0.00	0.19	0.00	0.00	0.17	3.19
pHxLxV	6	0.00	0.00	0.01	0.00	0.00	0.10	0.00	0.00	0.05	2.29
Error	48	0.17	0.00		0.16	0.00		0.33	0.00		
TSS	71	0.39	0.0055		0.38	0.0054		0.78	0.0055		

Appendix XXIV: Effect of pH, lime and varieties on potassium content in stover

SOV	DE	2016	2016			2017					Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	0.02	0.02	11.10*	0.03	0.03	11.94*	0.05	0.03	11.54*	4.04
Lime (L)	3	0.22	0.07	34.53*	0.26	0.09	37.37*	0.47	0.08	36.00*	2.80
Variety(V)	2	0.00	0.00	0.95	0.01	0.00	1.63	0.01	0.00	1.30	3.19
pH x L	3	0.00	0.00	0.73	0.01	0.00	1.00	0.01	0.00	0.87	2.80
L x V	6	0.00	0.00	0.04	0.00	0.00	0.03	0.00	0.00	0.04	2.29
pH x V	2	0.00	0.00	0.09	0.00	0.00	0.01	0.00	0.00	0.05	3.19
pHxLxV	6	0.00	0.00	0.19	0.00	0.00	0.05	0.00	0.00	0.11	2.29
Error	48	0.10	0.00		0.11	0.00		0.21	0.00		
TSS	71	0.36	0.0051		0.41	0.0058		0.76	0.0053		

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
pH	1	0.12	0.12	57.53*	0.15	0.15	56.56*	0.26	0.13	56.99*	4.04
Lime (L)	3	0.81	0.27	131.68*	0.95	0.32	122.25*	1.75	0.29	126.42*	2.80
Variety(V)	2	0.00	0.00	0.22	0.00	0.00	0.47	0.00	0.00	0.36	3.19
pH x L	3	0.00	0.00	0.12	0.00	0.00	0.08	0.00	0.00	0.10	2.80
L x V	6	0.00	0.00	0.06	0.00	0.00	0.04	0.00	0.00	0.05	2.29
pH x V	2	0.00	0.00	0.00	0.00	0.00	0.17	0.00	0.00	0.09	3.19
pHxLxV	6	0.00	0.00	0.07	0.00	0.00	0.01	0.00	0.00	0.04	2.29
Error	48	0.10	0.00		0.12	0.00		0.22	0.00		
TSS	71	1.03	0.0145		1.22	0.0172		2.25	0.0157		

Appendix XXV: Effect of pH, lime and varieties on calcium content in grain

Appendix XXVI: Effect of pH, lime and varieties on calcium content in stover

SOV	DE	2016	2016					Pooled			Etab
50V	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
pН	1	0.12	0.12	64.40*	0.06	0.06	33.27*	0.18	0.09	48.68*	4.04
Lime (L)	3	0.94	0.31	174.50*	0.81	0.27	148.90*	1.75	0.29	161.58*	2.80
Variety(V)	2	0.00	0.00	0.13	0.00	0.00	0.19	0.00	0.00	0.16	3.19
pH x L	3	0.00	0.00	0.84	0.00	0.00	0.52	0.01	0.00	0.68	2.80
L x V	6	0.00	0.00	0.12	0.00	0.00	0.06	0.00	0.00	0.09	2.29
pH x V	2	0.00	0.00	0.02	0.00	0.00	0.12	0.00	0.00	0.07	3.19
pHxLxV	6	0.00	0.00	0.19	0.00	0.00	0.05	0.00	0.00	0.12	2.29
Error	48	0.09	0.00		0.09	0.00		0.17	0.00		
TSS	71	1.15	0.0162		0.97	0.0137		2.12	0.0148		

Appendix XXVII: Effect of pH, lime and varieties on boron content in grain

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
pН	1	0.01	0.01	2.72	0.02	0.02	2.66	0.04	0.02	2.68	4.04
Lime (L)	3	0.58	0.19	38.40*	0.63	0.21	23.63*	1.20	0.20	28.98*	2.80
Variety(V)	2	0.00	0.00	0.46	0.01	0.00	0.55	0.01	0.00	0.51	3.19
pH x L	3	0.00	0.00	0.16	0.00	0.00	0.12	0.01	0.00	0.13	2.80
L x V	6	0.00	0.00	0.07	0.00	0.00	0.05	0.00	0.00	0.05	2.29
pH x V	2	0.00	0.00	0.01	0.00	0.00	0.10	0.00	0.00	0.07	3.19
pHxLxV	6	0.00	0.00	0.09	0.01	0.00	0.10	0.01	0.00	0.10	2.29
Error	48	0.24	0.01		0.42	0.01		0.66	0.01		
TSS	71	0.84	0.0118		1.10	0.0155		1.96	0.0137		

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
pH	1	0.03	0.03	17.03*	0.02	0.02	11.01*	0.05	0.02	13.57*	4.04
Lime (L)	3	0.41	0.14	87.78*	0.42	0.14	67.42*	0.83	0.14	76.08*	2.80
Variety(V)	2	0.00	0.00	0.41	0.00	0.00	0.39	0.00	0.00	0.40	3.19
pH x L	3	0.00	0.00	0.88	0.01	0.00	0.82	0.01	0.00	0.85	2.80
L x V	6	0.00	0.00	0.11	0.00	0.00	0.04	0.00	0.00	0.07	2.29
pH x V	2	0.00	0.00	0.10	0.00	0.00	0.16	0.00	0.00	0.13	3.19
pHxLxV	6	0.00	0.00	0.03	0.00	0.00	0.03	0.00	0.00	0.03	2.29
Error	48	0.07	0.00		0.10	0.00		0.18	0.00		
TSS	71	0.52	0.0073		0.56	0.0079		1.13	0.0079		

Appendix XVIII: Effect of pH, lime and varieties on boron content in stover

Appendix XXIX: Effect of pH, lime and varieties on nitrogen uptake in grain

SOV 1	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
pН	1	203847.51	203847.51	71.35*	262046.97	262046.97	124.95*	465894.48	232947.24	94.04*	4.04
Lime (L)	3	1399535.44	466511.81	163.28*	1667930.65	555976.88	265.10*	3067466.08	511244.35	206.38*	2.80
Variety(V)	2	134282.26	67141.13	23.50*	112391.20	56195.60	26.79*	246673.45	61668.36	24.89*	3.19
pH x L	3	8610.77	2870.26	1.00	2403.00	801.00	0.38	11013.77	1835.63	0.74	2.80
L x V	6	7726.06	1287.68	0.45	4394.81	732.47	0.35	12120.87	1010.07	0.41	2.29
pH x V	2	1023.13	511.56	0.18	153.82	76.91	0.04	1176.94	294.24	0.12	3.19
pHxLxV	6	8279.62	1379.94	0.48	2919.38	486.56	0.23	11199.00	933.25	0.38	2.29
Error	48	137139.66	2857.08		100668.35	2097.26		237808.01	2477.17		
TSS	71	1900444.44	26766.82		2152908.17	30322.65		4072681.49	28480.29		

Appendix XXX: Effect of pH, lime and varieties on nitrogen uptake in stover

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	172280.59	172280.59	32.95*	201368.10	201368.10	26.56*	373648.69	186824.34	29.17*	4.04
Lime (L)	3	1161270.02	387090.01	74.04*	1166250.23	388750.08	51.28*	2327520.26	387920.04	60.57*	2.80
Variety(V)	2	68347.59	34173.80	6.54*	90163.62	45081.81	5.95*	158511.22	39627.80	6.19*	3.19
pH x L	3	7581.49	2527.16	0.48	9117.21	3039.07	0.40	16698.69	2783.12	0.43	2.80
L x V	6	7554.43	1259.07	0.24	7261.85	1210.31	0.16	14816.28	1234.69	0.19	2.29
pH x V	2	5743.91	2871.95	0.55	4889.33	2444.66	0.32	10633.23	2658.31	0.42	3.19
pHxLxV	6	4521.04	753.51	0.14	2643.17	440.53	0.06	7164.21	597.02	0.09	2.29
Error	48	250944.39	5228.01		363863.06	7580.48		614807.44	6404.24		
TSS	71	1678243.45	23637.23		1845556.56	25993.75		3629003.48	25377.65		

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pH	1	59272.39	59272.39	134.20*	91651.22	91651.22	174.10*	150923.61	75461.80	155.90*	4.04
Lime (L)	3	135847.62	45282.54	102.52*	221569.58	73856.53	140.30*	357417.20	59569.53	123.06*	2.80
Variety(V)	2	18942.80	9471.40	21.44*	36518.61	18259.30	34.69*	55461.40	13865.35	28.64*	3.19
pH x L	3	3090.02	1030.01	2.33	4850.56	1616.85	3.07*	7940.58	1323.43	2.73	2.80
L x V	6	1090.83	181.80	0.41	588.48	98.08	0.19	1679.30	139.94	0.29	2.29
pH x V	2	114.81	57.40	0.13	61.72	30.86	0.06	176.52	44.13	0.09	3.19
pHxLxV	6	350.75	58.46	0.13	243.66	40.61	0.08	594.41	49.53	0.10	2.29
Error	48	21200.74	441.68		25268.36	526.42		46469.10	484.05		
TSS	71	239909.95	3379.01		380752.18	5362.71		650751.37	4550.71		

Appendix XXXI: Effect of pH, lime and varieties on phosphorus uptake in grain

Appendix XXXII: Effect of pH, lime and varieties on phosphorus uptake in stover

SOV 3	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
pН	1	62276.85	62276.85	32.01*	35147.40	35147.40	18.22*	97424.25	48712.13	25.15*	4.04
Lime (L)	3	105314.81	35104.94	18.05*	69080.97	23026.99	11.94*	174395.78	29065.96	15.00*	2.80
Variety(V)	2	10486.56	5243.28	2.70	10818.73	5409.37	2.80	21305.29	5326.32	2.75	3.19
pH x L	3	4270.70	1423.57	0.73	3919.97	1306.66	0.68	8190.67	1365.11	0.70	2.80
L x V	6	611.60	101.93	0.05	451.28	75.21	0.04	1062.88	88.57	0.05	2.29
pH x V	2	90.45	45.22	0.02	1.64	0.82	0.00	92.09	23.02	0.01	3.19
pHxLxV	6	743.39	123.90	0.06	1120.88	186.81	0.10	1864.28	155.36	0.08	2.29
Error	48	93375.19	1945.32		92598.64	1929.14		185973.83	1937.23		
TSS	71	277169.55	3903.80		213139.52	3001.97		493529.44	3451.25		

Appendix XXXIII: Effect of pH, lime and varieties on potassium uptake in grain

SOVIIpH1Lime (L)3Variety(V)2variety(V)2	DE	2016	2016					Pooled			E tob
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	373648.69	186824.34	29.17*	170276.97	170276.97	87.44*	300542.11	150271.06	78.49*	4.04
Lime (L)	3	2327520.26	387920.04	60.57*	223336.24	74445.41	38.23*	420854.62	70142.44	36.64*	2.80
Variety(V)	2	158511.22	39627.80	6.19*	41049.28	20524.64	10.54*	87629.88	21907.47	11.44*	3.19
pH x L	3	16698.69	2783.12	0.43	7943.14	2647.71	1.36	12140.27	2023.38	1.06	2.80
L x V	6	14816.28	1234.69	0.19	2778.17	463.03	0.24	5470.68	455.89	0.24	2.29
pH x V	2	10633.23	2658.31	0.42	167.60	83.80	0.04	434.90	108.73	0.06	3.19
pHxLxV	6	7164.21	597.02	0.09	908.32	151.39	0.08	1427.06	118.92	0.06	2.29
Error	48	614807.44	6404.24		93475.93	1947.42		183800.87	1914.59		
TSS	71	3629003.48	25377.65		539935.67	7604.728		1020490.50	7136.30		

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lab
pН	1	579119.36	579119.36	57.46*	635034.45	635034.45	64.25*	1214153.81	607076.91	60.82*	4.04
Lime (L)	3	2364449.41	788149.80	78.20*	2527899.54	842633.18	85.26*	4892349.0	815391.49	81.70*	2.80
Variety(V)	2	110976.68	55488.34	5.51*	175548.29	87774.14	8.88*	286524.96	71631.24	7.18*	3.19
pH x L	3	17148.79	5716.26	0.57	22308.60	7436.20	0.75	39457.39	6576.23	0.66	2.80
L x V	6	3563.48	593.91	0.06	2872.18	478.70	0.05	6435.66	536.30	0.05	2.29
pH x V	2	1385.81	692.90	0.07	68.69	34.35	0.00	1454.50	363.63	0.04	3.19
pHxLxV	6	5377.85	896.31	0.09	1914.97	319.16	0.03	7292.82	607.73	0.06	2.29
Error	48	483754.60	10078.22		474399.39	9883.32		958153.99	9980.77		
TSS	71	3565775.98	50222.2		3840046.10	54085.16		7451779.79	52110.35		

Appendix XXXIV: Effect of pH, lime and varieties on potassium uptake in stover

Appendix XXXV: Effect of pH, lime and varieties on calcium uptake in grain

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	141534.17	141534.17	124.35*	203476.28	203476.28	160.31*	345010.45	172505.22	143.31*	4.04
Lime (L)	3	646145.77	215381.92	189.23*	747387.40	249129.13	196.27*	1393533.17	232255.53	192.94*	2.80
Variety(V)	2	14763.11	7381.55	6.49*	16267.91	8133.95	6.41*	31031.02	7757.75	6.44*	3.19
pH x L	3	1669.21	556.40	0.49	6505.02	2168.34	1.71	8174.23	1362.37	1.13	2.80
L x V	6	2066.59	344.43	0.30	1332.85	222.14	0.18	3399.44	283.29	0.24	2.29
pH x V	2	5.26	2.63	0.00	252.13	126.07	0.10	257.39	64.35	0.05	3.19
pHxLxV	6	589.27	98.21	0.09	112.31	18.72	0.01	701.58	58.46	0.05	2.29
Error	48	54633.14	1138.19		60926.39	1269.30		115559.53	1203.75		
TSS	71	861406.52	12132.49		1036260.28	14595.22		1904261.32	13316.51		

Appendix XXXVI: Effect of pH, lime and varieties on calcium uptake in stover

SOV D	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
pН	1	428687.07	428687.07	105.72*	320089.34	320089.34	81.28*	748776.41	374388.21	93.68*	4.04
Lime (L)	3	2332050.57	777350.19	191.71*	2206432.25	735477.42	186.76*	4538482.8	756413.80	189.27*	2.80
Variety(V)	2	9085.99	4542.99	1.12	19210.15	9605.08	2.44	28296.14	7074.04	1.77	3.19
pH x L	3	8241.87	2747.29	0.68	2803.58	934.53	0.24	11045.45	1840.91	0.46	2.80
L x V	6	3738.67	623.11	0.15	1414.39	235.73	0.06	5153.07	429.42	0.11	2.29
pH x V	2	283.30	141.65	0.03	480.79	240.40	0.06	764.09	191.02	0.05	3.19
pHxLxV	6	3364.84	560.81	0.14	768.88	128.15	0.03	4133.72	344.48	0.09	2.29
Error	48	194631.97	4054.83		189027.59	3938.07		383659.56	3996.45		
TSS	71	2980084.28	41973.02		2740226.98	38594.75		5755133.30	40245.69		

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pH	1	100467.02	100467.02	191.66*	152206.67	152206.67	294.77*	252673.69	126336.8	242.83*	4.04
Lime (L)	3	324794.88	108264.96	206.54*	336988.14	112329.38	217.54*	661783.02	110297.1	212.00*	2.80
Variety(V)	2	42654.17	21327.09	40.69*	37843.06	18921.53	36.64*	80497.23	20124.31	38.68*	3.19
pH x L	3	2319.17	773.06	1.47	3431.28	1143.76	2.22	5750.45	958.41	1.84	2.80
L x V	6	3528.80	588.13	1.12	1020.58	170.10	0.33	4549.38	379.12	0.73	2.29
pH x V	2	333.65	166.83	0.32	673.33	336.66	0.65	1006.98	251.74	0.48	3.19
pHxLxV	6	1419.58	236.60	0.45	504.87	84.14	0.16	1924.45	160.37	0.31	2.29
Error	48	25160.69	524.18		24785.01	516.35		49945.70	520.27		
TSS	71	500677.95	7051.802		557452.94	7851.45		1059743.86	7410.80		

Appendix XXXVII: Effect of pH, lime and varieties on boron uptake in grain

Appendix XXXVIII: Effect of pH, lime and varieties on boron uptake in stover

SOV	DE	2016	2016					Pooled			Etab
50 V	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
pН	1	31314.62	31314.62	63.97*	34037.45	34037.45	57.39*	65352.07	32676.03	60.36*	4.04
Lime (L)	3	116140.35	38713.45	79.09*	118507.80	39502.60	66.60*	234648.14	39108.02	72.25*	2.80
Variety(V)	2	6017.30	3008.65	6.15*	7934.59	3967.29	6.69*	13951.89	3487.97	6.44*	3.19
pH x L	3	53.04	17.68	0.04	115.70	38.57	0.07	168.74	28.12	0.05	2.80
L x V	6	120.37	20.06	0.04	61.81	10.30	0.02	182.18	15.18	0.03	2.29
pH x V	2	28.78	14.39	0.03	16.19	8.10	0.01	44.97	11.24	0.02	3.19
pHxLxV	6	135.54	22.59	0.05	20.51	3.42	0.01	156.05	13.00	0.02	2.29
Error	48	23496.43	489.51		28469.55	593.12		51965.97	541.31		
TSS	71	177306.42	2497.274		189163.59	2664.276		373761.18	2613.71		

Appendix XXXIX: Effect of pH, lime and varieties on pH of post-harvest soil

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
pН	1	1.68	1.68	1259.44*	1.58	1.58	1132.82*	3.26	1.63	1194.65*	4.04
Lime (L)	3	23.00	7.67	5755.71*	21.61	7.20	5159.54*	44.60	7.43	5450.64*	2.80
Variety(V)	2	0.00	0.00	0.28	0.00	0.00	0.24	0.00	0.00	0.26	3.19
pH x L	3	0.88	0.29	219.27*	0.96	0.32	229.77*	1.84	0.31	224.65*	2.80
L x V	6	0.01	0.00	0.67	0.00	0.00	0.34	0.01	0.00	0.50	2.29
pH x V	2	0.00	0.00	0.31	0.00	0.00	0.71	0.00	0.00	0.51	3.19
pHxLxV	6	0.00	0.00	0.41	0.01	0.00	0.68	0.01	0.00	0.55	2.29
Error	48	0.06	0.00		0.07	0.00		0.13	0.00		
TSS	71	25.63	0.36		24.23	0.34		49.85	0.35		

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	0.00	0.004	1.56	0.00	0.00	0.80	0.01	0.00	1.19	4.04
Lime (L)	3	0.01	0.00	1.98	0.01	0.00	1.40	0.02	0.00	1.69	2.80
Variety(V)	2	0.00	0.00	0.60	0.00	0.00	0.11	0.00	0.00	0.36	3.19
pH x L	3	0.01	0.00	0.83	0.00	0.00	0.19	0.01	0.00	0.51	2.80
L x V	6	0.00	0.00	0.15	0.00	0.00	0.15	0.00	0.00	0.15	2.29
pH x V	2	0.00	0.00	0.30	0.00	0.00	0.11	0.00	0.00	0.21	3.19
pHxLxV	6	0.00	0.00	0.20	0.00	0.00	0.05	0.00	0.00	0.13	2.29
Error	48	0.11	0.0023		0.11	0.00		0.22	0.00		
TSS	71	0.14	0.0020		0.12	0.0017		0.29	0.0020		

Appendix XL: Effect of pH, lime and varieties on electrical conductivity of post-harvest soil

Appendix XLI: Effect of pH, lime and varieties on percent base saturation of post-harvest soil

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
pН	1	8.31	8.31	1.45	3.56	3.56	0.84	11.87	5.94	1.19	4.04
Lime (L)	3	1040.20	346.73	60.44*	841.47	280.49	66.28*	1881.67	313.61	62.91*	2.80
Variety(V)	2	1.42	0.71	0.12	0.31	0.15	0.04	1.73	0.43	0.09	3.19
pH x L	3	45.93	15.31	2.67	67.01	22.34	5.28*	112.94	18.82	3.78*	2.80
L x V	6	1.45	0.24	0.04	0.24	0.04	0.01	1.70	0.14	0.03	2.29
pH x V	2	0.11	0.05	0.01	0.05	0.03	0.01	0.16	0.04	0.01	3.19
pHxLxV	6	1.28	0.21	0.04	0.14	0.02	0.01	1.42	0.12	0.02	2.29
Error	48	275.39	5.74		203.15	4.23		478.53	4.98		
TSS	71	1374.09	19.35		1115.93	15.72		2499.91	35.21		

Appendix XLII: Effect of pH, lime and varieties on organic carbon of post-harvest soil

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	0.00	0.00	0.47	0.00	0.00	0.76	0.00	0.00	0.61	4.04
Lime (L)	3	0.01	0.00	0.57	0.00	0.00	0.36	0.01	0.00	0.47	2.80
Variety(V)	2	0.00	0.00	0.19	0.00	0.00	0.32	0.00	0.00	0.25	3.19
pH x L	3	0.00	0.00	0.50	0.00	0.00	0.11	0.01	0.00	0.32	2.80
L x V	6	0.00	0.00	0.17	0.00	0.00	0.04	0.00	0.00	0.11	2.29
pH x V	2	0.00	0.00	0.00	0.00	0.00	0.16	0.00	0.00	0.08	3.19
pHxLxV	6	0.00	0.00	0.06	0.00	0.00	0.10	0.00	0.00	0.08	2.29
Error	48	0.15	0.00		0.14	0.00		0.29	0.00		
TSS	71	0.17	0.0024		0.15	0.0021		0.32	0.0045		

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
pН	1	148.26	148.26	1.74	51.44	51.44	0.58	199.71	99.85	1.15	4.04
Lime (L)	3	17259.93	5753.31	67.33*	17672.00	5890.67	66.84*	34931.93	5821.99	67.08*	2.80
Variety(V)	2	37.93	18.97	0.22	58.67	29.33	0.33	96.60	24.15	0.28	3.19
pH x L	3	122.22	40.74	0.48	298.91	99.64	1.13	421.12	70.19	0.81	2.80
L x V	6	25.13	4.19	0.05	22.02	3.67	0.04	47.15	3.93	0.05	2.29
pH x V	2	0.46	0.23	0.00	12.95	6.47	0.07	13.41	3.35	0.04	3.19
pHxLxV	6	39.24	6.54	0.08	30.51	5.08	0.06	69.74	5.81	0.07	2.29
Error	48	4101.50	85.45		4230.16	88.13		8331.65	86.79		
TSS	71	21734.67	306.12		22376.65	315.16		44142.55	308.69		

Appendix XLIII: Effect of pH, lime and varieties on available nitrogen of post-harvest soil

Appendix XLIV: Effect of pH, lime and varieties on available phosphorus of post-harvest soil

SOV	DE	2016	2016			2017		Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	19.46	19.46	16.82*	22.78	22.78	8.76*	42.24	21.12	11.25*	4.04
Lime (L)	3	133.40	44.47	38.45*	67.98	22.66	8.72*	201.38	33.56	17.87*	2.80
Variety(V)	2	0.27	0.14	0.12	1.11	0.56	0.21	1.38	0.35	0.18	3.19
pH x L	3	0.66	0.22	0.19	4.79	1.60	0.61	5.45	0.91	0.48	2.80
L x V	6	1.29	0.22	0.19	0.73	0.12	0.05	2.02	0.17	0.09	2.29
pH x V	2	0.59	0.30	0.26	0.01	0.00	0.00	0.60	0.15	0.08	3.19
pHxLxV	6	0.87	0.14	0.13	0.28	0.05	0.02	1.15	0.10	0.05	2.29
Error	48	55.51	1.16		124.76	2.60		180.28	1.88		
TSS	71	212.06	2.99		222.44	3.13		456.84	3.19		

Appendix XLV: Effect of pH, lime and varieties on available potassium of post-harvest soil

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
pН	1	18.97	18.97	1.80	5.81	5.81	0.56	24.79	12.39	1.19	4.04
Lime (L)	3	104.24	34.75	3.31*	262.75	87.58	8.47*	366.99	61.17	5.87*	2.80
Variety(V)	2	5.10	2.55	0.24	0.58	0.29	0.03	5.68	1.42	0.14	3.19
pH x L	3	63.18	21.06	2.00	10.76	3.59	0.35	73.94	12.32	1.18	2.80
L x V	6	10.51	1.75	0.17	14.71	2.45	0.24	25.23	2.10	0.20	2.29
pH x V	2	0.51	0.25	0.02	2.52	1.26	0.12	3.02	0.76	0.07	3.19
pHxLxV	6	7.77	1.29	0.12	13.28	2.21	0.21	21.04	1.75	0.17	2.29
Error	48	504.61	10.51		496.45	10.34		1001.05	10.43		
TSS	71	714.89	10.07		806.86	11.36		1525.21	21.48		

SOV I	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
pН	1	1.05	1.05	8.41*	1.03	1.03	8.06*	2.09	1.04	8.23*	4.04
Lime (L)	3	143.04	47.68	380.68*	145.48	48.49	378.58*	288.52	48.09	379.62*	2.80
Variety(V)	2	0.00	0.00	0.01	0.01	0.00	0.02	0.01	0.00	0.01	3.19
pH x L	3	0.14	0.05	0.36	0.22	0.07	0.57	0.35	0.06	0.46	2.80
L x V	6	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	2.29
pH x V	2	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	3.19
pHxLxV	6	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	2.29
Error	48	6.01	0.13		6.15	0.13		12.16	0.13		
TSS	71	150.25	2.12		152.89	2.15		303.14	2.12		

Appendix XLVI: Effect of pH, lime and varieties on exchangeable calcium of post-harvest soil

Appendix XLVII: Effect of pH, lime and varieties on available boron of post-harvest soil

SOV I	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
pН	1	0.13	0.13	156.71*	0.13	0.13	116.13*	0.25	0.13	133.53*	4.04
Lime (L)	3	0.01	0.00	5.73*	0.02	0.01	5.08*	0.03	0.01	5.36*	2.80
Variety(V)	2	0.001	0.0006	0.74	0.003	0.0016	1.53	0.004	0.0011	1.19	3.19
pH x L	3	0.001	0.0002	0.29	0.0055	0.0018	1.71	0.006	0.0010	1.10	2.80
L x V	6	0.0026	0.00043	0.53	0.0061	0.0010	0.95	0.0087	0.00072	0.77	2.29
pH x V	2	0.0019	0.0010	1.18	0.000	0.0002	0.19	0.002	0.0006	0.62	3.19
pHxLxV	6	0.0004	0.00007	0.09	0.004	0.0007	0.64	0.005	0.0004	0.41	2.29
Error	48	0.0388	0.0008		0.0517	0.0011		0.09	0.0009		
TSS	71	0.19	0.0027		0.21	0.003		0.41	0.0058		

Appendix XLVIII: Effect of pH, lime and varieties on exchangeable hydrogen of post-harvest soil

SOV I	DE	2016	2016			2017					Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
pН	1	0.26	0.26	11.26*	0.69	0.69	19.67*	0.95	0.48	16.30*	4.04
Lime (L)	3	3.42	1.14	48.60*	2.53	0.84	24.06*	5.95	0.99	33.89*	2.80
Variety(V)	2	0.02	0.01	0.48	0.00	0.00	0.06	0.03	0.01	0.23	3.19
pH x L	3	0.28	0.09	4.05*	0.33	0.11	3.12*	0.61	0.10	3.49*	2.80
L x V	6	0.03	0.01	0.25	0.00	0.00	0.02	0.04	0.00	0.11	2.29
pH x V	2	0.01	0.01	0.29	0.00	0.00	0.00	0.01	0.00	0.11	3.19
pHxLxV	6	0.03	0.00	0.19	0.01	0.00	0.02	0.03	0.00	0.09	2.29
Error	48	1.13	0.02		1.68	0.04		2.81	0.03		
TSS	71	5.19	0.073		5.25	0.074		10.46	0.15		

SOV I	DE	2016	2016			2017					Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
pН	1	0.19	0.19	19.53*	0.27	0.27	43.53*	0.46	0.23	28.94*	4.04
Lime (L)	3	4.26	1.42	148.14*	5.25	1.75	283.40*	9.51	1.58	201.18*	2.80
Variety(V)	2	0.02	0.01	1.05	0.00	0.00	0.03	0.02	0.01	0.65	3.19
pH x L	3	0.15	0.05	5.15*	0.15	0.05	8.32*	0.30	0.05	6.39*	2.80
L x V	6	0.03	0.01	0.55	0.01	0.00	0.14	0.04	0.00	0.39	2.29
pH x V	2	0.01	0.01	0.53	0.00	0.00	0.02	0.01	0.00	0.33	3.19
pHxLxV	6	0.03	0.00	0.45	0.00	0.00	0.03	0.03	0.00	0.28	2.29
Error	48	0.46	0.01		0.30	0.01		0.76	0.01		
TSS	71	5.14	0.072		5.98	0.084		11.13	0.16		

Appendix XLIX: Effect of pH, lime and varieties on exchangeable aluminium of post-harvest soil

Appendix L: Effect of pH, lime and varieties on total potential acidity of post-harvest soil

SOV	DE	2016	2016					Pooled			Etab
50V	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
pН	1	8.26	8.26	8.36*	16.18	16.18	16.40*	24.43	12.22	12.38*	4.04
Lime (L)	3	182.74	60.91	61.72*	180.99	60.33	61.17*	363.73	60.62	61.45*	2.80
Variety(V)	2	0.01	0.00	0.00	0.00	0.00	0.00	0.01	0.00	0.00	3.19
pH x L	3	1.32	0.44	0.45	1.52	0.51	0.51	2.84	0.47	0.48	2.80
L x V	6	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	2.29
pH x V	2	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	3.19
pHxLxV	6	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	2.29
Error	48	47.37	0.99		47.34	0.99		94.71	0.99		
TSS	71	239.71	3.38		246.03	3.47		486.71	3.40		

Appendix LI: Effect of lime, phosphorus and boron on plant height of maize at 30 DAS

SOM	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	223.31	223.31	17.77*	157.83	157.83	11.05*	381.14	190.57	14.20*	4.04
Phosphorus (P)	3	117.93	39.31	3.13*	441.33	147.11	10.30*	559.26	93.21	6.94*	2.80
Boron (B)	2	217.18	108.59	8.64*	361.38	180.69	12.66*	578.55	144.64	10.78*	3.19
L x P	3	64.72	21.57	1.72	45.87	15.29	1.07	110.59	18.43	1.37	2.80
РхВ	6	20.67	3.45	0.27	12.45	2.08	0.15	33.13	2.76	0.21	2.29
LxB	2	13.59	6.80	0.54	4.54	2.27	0.16	18.13	4.53	0.34	3.19
L x P x B	6	6.06	1.01	0.08	10.77	1.80	0.13	16.83	1.40	0.10	2.29
Error	48	603.11	12.56		685.32	14.28		1288.43	13.42		
TSS	71	1266.56	17.84		1719.49	24.22		3138.57	44.21		

SOV	DF	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
Lime (L)	1	8268.98	8268.98	452.73*	7539.92	7539.92	416.98*	15808.90	7904.45	434.95*	4.04
Phosphorus (P)	3	12600.35	4200.12	229.96*	11328.23	3776.08	208.83*	23928.57	3988.10	219.45*	2.80
Boron (B)	2	2119.37	1059.69	58.02*	1561.93	780.96	43.19*	3681.30	920.32	50.64*	3.19
LxP	3	183.07	61.02	3.34*	58.77	19.59	1.08	241.84	40.31	2.22	2.80
P x B	6	60.01	10.00	0.55	57.64	9.61	0.53	117.65	9.80	0.54	2.29
L x B	2	5.51	2.75	0.15	66.61	33.31	1.84	72.12	18.03	0.99	3.19
L x P x B	6	53.63	8.94	0.49	140.36	23.39	1.29	193.99	16.17	0.89	2.29
Error	48	876.70	18.26		867.94	18.08		1744.64	18.17		
TSS	71	24167.62	340.39		21621.39	304.53		45958.88	321.39		

Appendix LII: Effect of lime, phosphorus and boron on plant height of maize at 60 DAS

Appendix LIII: Effect of lime, phosphorus and boron on plant height of maize at harvest

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lao
Lime (L)	1	11917.11	11917.11	1206.37*	11730.01	11730.01	758.77*	23647.12	11823.56	933.28*	4.04
Phosphorus (P)	3	17427.53	5809.18	588.06*	19272.40	6424.13	415.56*	36699.93	6116.66	482.81*	2.80
Boron (B)	2	1571.78	785.89	79.56*	938.14	469.07	30.34*	2509.92	627.48	49.53*	3.19
LxP	3	24.53	8.18	0.83	40.81	13.60	0.88	65.34	10.89	0.86	2.80
РхВ	6	113.27	18.88	1.91	57.90	9.65	0.62	171.17	14.26	1.13	2.29
L x B	2	16.70	8.35	0.85	3.63	1.82	0.12	20.33	5.08	0.40	3.19
L x P x B	6	134.85	22.47	2.28	244.58	40.76	2.64*	379.42	31.62	2.50*	2.29
Error	48	474.17	9.88		742.04	15.46		1216.21	12.67		
TSS	71	31679.94	446.20		33029.51	465.20		64727.72	452.64		

Appendix LIV: Effect of lime, phosphorus and boron on number of leaves of maize at 30 DAS

SOM	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	1.39	1.39	5.56*	1.13	1.13	4.05*	2.51	1.26	4.76*	4.04
Phosphorus (P)	3	2.56	0.85	3.41*	2.82	0.94	3.38*	5.38	0.90	3.39*	2.80
Boron (B)	2	1.03	0.51	2.06	0.58	0.29	1.05	1.61	0.40	1.53	3.19
L x P	3	0.28	0.09	0.37	0.26	0.09	0.32	0.54	0.09	0.34	2.80
РхВ	6	0.19	0.03	0.13	0.31	0.05	0.18	0.50	0.04	0.16	2.29
LxB	2	0.19	0.10	0.39	0.08	0.04	0.15	0.28	0.07	0.26	3.19
L x P x B	6	0.14	0.02	0.09	0.36	0.06	0.22	0.50	0.04	0.16	2.29
Error	48	12.00	0.25		13.33	0.28		25.33	0.26		
TSS	71	17.78	0.25		18.88	0.27		36.83	0.52		

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	8.68	8.68	26.04*	8.68	8.68	31.25*	17.36	8.68	28.41*	4.04
Phosphorus (P)	3	19.49	6.50	19.49*	12.71	4.24	15.25*	32.19	5.37	17.56*	2.80
Boron (B)	2	8.36	4.18	12.54*	3.03	1.51	5.45*	11.39	2.85	9.32*	3.19
LxP	3	0.49	0.16	0.49	0.60	0.20	0.72	1.08	0.18	0.59	2.80
P x B	6	0.64	0.11	0.32	0.42	0.07	0.25	1.06	0.09	0.29	2.29
LxB	2	0.03	0.01	0.04	0.03	0.01	0.05	0.06	0.01	0.05	3.19
L x P x B	6	0.31	0.05	0.15	1.19	0.20	0.72	1.50	0.13	0.41	2.29
Error	48	16.00	0.33		13.33	0.28		29.33	0.31		
TSS	71	53.99	0.76		39.99	0.56		94.00	1.32		

Appendix LV: Effect of lime, phosphorus and boron on number of leaves of maize at 60 DAS

Appendix LVI: Effect of lime, phosphorus and boron on number of leaves of maize at harvest

SOM	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	33.35	33.35	82.79*	21.13	21.13	34.57*	54.47	27.24	53.73*	4.04
Phosphorus (P)	3	19.71	6.57	16.31*	21.15	7.05	11.54*	40.86	6.81	13.43*	2.80
Boron (B)	2	9.19	4.60	11.41*	6.78	3.39	5.55*	15.97	3.99	7.88*	3.19
LxP	3	0.26	0.09	0.22	0.26	0.09	0.14	0.53	0.09	0.17	2.80
РхВ	6	0.92	0.15	0.38	1.22	0.20	0.33	2.14	0.18	0.35	2.29
LxB	2	0.53	0.26	0.66	0.33	0.17	0.27	0.86	0.22	0.42	3.19
L x P x B	6	0.69	0.12	0.29	0.78	0.13	0.21	1.47	0.12	0.24	2.29
Error	48	19.33	0.40		29.33	0.61		48.67	0.51		
TSS	71	83.99	1.18		80.99	1.14		173.97	2.45		

Appendix LVII: Effect of lime, phosphorus and boron on leaf area index of maize at 30 DAS

SOM	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	0.04	0.04	7.37*	0.02	0.02	5.10*	0.06	0.03	6.47*	4.04
Phosphorus (P)	3	0.10	0.03	6.12*	0.16	0.05	14.40*	0.27	0.04	9.42*	2.80
Boron (B)	2	0.01	0.01	0.96	0.00	0.00	0.13	0.01	0.00	0.63	3.19
LxP	3	0.01	0.00	0.78	0.01	0.00	0.86	0.02	0.00	0.81	2.80
РхВ	6	0.01	0.00	0.20	0.01	0.00	0.35	0.01	0.00	0.26	2.29
LxB	2	0.02	0.01	1.49	0.00	0.00	0.33	0.02	0.00	1.03	3.19
L x P x B	6	0.03	0.00	0.81	0.00	0.00	0.11	0.03	0.00	0.53	2.29
Error	48	0.27	0.01		0.18	0.00		0.46	0.00		
TSS	71	0.50	0.007		0.39	0.0055		0.92	0.013		

SOV	DF	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
Lime (L)	1	0.05	0.05	47.57*	0.05	0.05	35.50*	0.10	0.05	40.45*	4.04
Phosphorus (P)	3	0.03	0.01	10.57*	0.06	0.02	13.16*	0.09	0.02	12.10*	2.80
Boron (B)	2	0.00	0.00	0.10	0.00	0.00	0.56	0.00	0.00	0.37	3.19
LxP	3	0.00	0.00	0.51	0.00	0.00	0.35	0.00	0.00	0.42	2.80
P x B	6	0.00	0.00	0.28	0.00	0.00	0.25	0.00	0.00	0.26	2.29
L x B	2	0.00	0.00	0.14	0.00	0.00	0.17	0.00	0.00	0.16	3.19
L x P x B	6	0.00	0.00	0.07	0.00	0.00	0.23	0.00	0.00	0.17	2.29
Error	48	0.05	0.00		0.07	0.00		0.12	0.00		
TSS	71	0.14	0.0020		0.19	0.0027		0.35	0.0049		

Appendix LVIII: Effect of lime, phosphorus and boron on leaf area index of maize at 60 DAS

Appendix LIX: Effect of lime, phosphorus and boron on leaf area index of maize at harvest

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
Lime (L)	1	0.01	0.01	11.08*	0.01	0.01	11.75*	0.01	0.01	11.47*	4.04
Phosphorus (P)	3	0.01	0.00	3.40*	0.02	0.01	8.61*	0.02	0.00	6.39*	2.80
Boron (B)	2	0.00	0.00	2.51	0.00	0.00	0.41	0.00	0.00	1.30	3.19
L x P	3	0.00	0.00	0.51	0.00	0.00	0.91	0.00	0.00	0.74	2.80
РхВ	6	0.00	0.00	1.18	0.00	0.00	0.25	0.00	0.00	0.64	2.29
L x B	2	0.00	0.00	0.09	0.00	0.00	2.86	0.00	0.00	1.68	3.19
L x P x B	6	0.00	0.00	0.73	0.00	0.00	0.44	0.00	0.00	0.56	2.29
Error	48	0.03	0.00		0.04	0.00		0.06	0.00		
TSS	71	0.05	0.00070		0.07	0.0010		0.13	0.0018		

Appendix LX: Effect of lime, phosphorus and boron on cob length of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
Lime (L)	1	4.45	4.45	50.62*	1.97	1.97	21.07*	6.42	3.21	35.40*	4.04
Phosphorus (P)	3	51.02	17.01	193.46*	38.35	12.78	136.98*	89.38	14.90	164.38*	2.80
Boron (B)	2	5.14	2.57	29.21*	3.40	1.70	18.20*	8.53	2.13	23.54*	3.19
LxP	3	0.33	0.11	1.26	0.24	0.08	0.86	0.57	0.10	1.05	2.80
РхВ	6	0.91	0.15	1.72	1.11	0.18	1.98	2.02	0.17	1.86	2.29
LxB	2	0.03	0.01	0.15	0.03	0.01	0.15	0.06	0.01	0.15	3.19
L x P x B	6	0.04	0.01	0.07	0.18	0.03	0.31	0.21	0.02	0.19	2.29
Error	48	4.22	0.09		4.48	0.09		8.70	0.09		
TSS	71	66.14	0.93		49.75	0.7		119.50	1.68		

SOV	DF	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
Lime (L)	1	3.87	3.87	28.09*	3.15	3.15	24.10*	7.02	3.51	26.15*	4.04
Phosphorus (P)	3	14.87	4.96	35.93*	15.24	5.08	38.87*	30.11	5.02	37.36*	2.80
Boron (B)	2	1.12	0.56	4.05*	1.33	0.67	5.11*	2.45	0.61	4.56*	3.19
LxP	3	0.22	0.07	0.54	0.17	0.06	0.43	0.39	0.07	0.49	2.80
P x B	6	0.17	0.03	0.20	0.29	0.05	0.37	0.46	0.04	0.28	2.29
LxB	2	0.04	0.02	0.16	0.00	0.00	0.01	0.05	0.01	0.09	3.19
L x P x B	6	0.05	0.01	0.06	0.11	0.02	0.13	0.15	0.01	0.09	2.29
Error	48	6.62	0.14		6.27	0.13		12.89	0.13		
TSS	71	26.96	0.38		26.57	0.37		53.56	0.37		

Appendix LXII: Effect of lime, phosphorus and boron on cob girth of maize

Appendix LXI: Effect of lime, phosphorus and boron on cob weight of maize

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	5270.22	5270.22	1425.29*	5573.92	5573.92	2294.84*	10844.14	5422.07	1770.03*	4.04
Phosphorus (P)	3	3767.48	1255.83	339.63*	3856.47	1285.49	529.25*	7623.95	1270.66	414.81*	2.80
Boron (B)	2	218.86	109.43	29.59*	316.80	158.40	65.22*	535.66	133.92	43.72*	3.19
LxP	3	131.83	43.94	11.88*	68.45	22.82	9.39*	200.27	33.38	10.90*	2.80
РхВ	6	21.35	3.56	0.96	9.33	1.55	0.64	30.68	2.56	0.83	2.29
LxB	2	1.50	0.75	0.20	1.47	0.73	0.30	2.97	0.74	0.24	3.19
L x P x B	6	8.05	1.34	0.36	11.50	1.92	0.79	19.55	1.63	0.53	2.29
Error	48	177.49	3.70		116.59	2.43		294.07	3.06		
TSS	71	9596.78	135.17		9954.52	140.20		19551.49	136.72		

Appendix LXIII: Effect of lime, phosphorus and boron on number of grains per row of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	136.13	136.13	544.50*	174.22	174.22	545.39*	310.35	155.17	545.00*	4.04
Phosphorus (P)	3	101.71	33.90	135.61*	152.50	50.83	159.13*	254.21	42.37	148.80*	2.80
Boron (B)	2	11.19	5.60	22.39*	11.03	5.51	17.26*	22.22	5.56	19.51*	3.19
LxP	3	5.82	1.94	7.76*	4.56	1.52	4.75*	10.38	1.73	6.07*	2.80
P x B	6	0.58	0.10	0.39	1.75	0.29	0.91	2.33	0.19	0.68	2.29
LxB	2	1.08	0.54	2.17	0.19	0.10	0.30	1.28	0.32	1.12	3.19
L x P x B	6	1.14	0.19	0.76	3.69	0.62	1.93	4.83	0.40	1.41	2.29
Error	48	12.00	0.25		15.33	0.32		27.33	0.28		
TSS	71	269.65	3.80		363.28	5.12		637.99	4.46		

SOV	DF -	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	45.13	45.13	58.02*	42.01	42.01	112.04*	87.14	43.57	75.59*	4.04
Phosphorus (P)	3	4.15	1.38	1.78	2.38	0.79	2.11	6.53	1.09	1.89	2.80
Boron (B)	2	1.75	0.88	1.12	1.44	0.72	1.93	3.19	0.80	1.39	3.19
L x P	3	0.60	0.20	0.26	0.15	0.05	0.14	0.75	0.13	0.22	2.80
P x B	6	0.47	0.08	0.10	0.67	0.11	0.30	1.14	0.09	0.16	2.29
LxB	2	0.25	0.13	0.16	0.11	0.06	0.15	0.36	0.09	0.16	3.19
L x P x B	6	0.19	0.03	0.04	0.22	0.04	0.10	0.42	0.03	0.06	2.29
Error	48	37.33	0.78		18.00	0.38		55.33	0.58		
TSS	71	89.88	1.27		64.99	0.92		155.31	1.09		

Appendix LXIV: Effect of lime, phosphorus and boron on number of rows per cob of maize

Appendix LXV: Effect of lime, phosphorus and boron on number of grains per cob of maize

SOV	DE	2016		2017			Pooled			Etab	
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	42389.01	42389.01	27.68*	51734.72	51734.72	118.16*	94123.74	47061.87	47.80*	4.04
Phosphorus (P)	3	43875.49	14625.16	9.55*	57347.00	19115.67	43.66*	101222.49	16870.41	17.14*	2.80
Boron (B)	2	12798.86	6399.43	4.18*	10527.08	5263.54	12.02*	23325.94	5831.49	5.92*	3.19
LxP	3	16932.93	5644.31	3.69*	42.28	14.09	0.03	16975.21	2829.20	2.87*	2.80
P x B	6	508.14	84.69	0.06	637.92	106.32	0.24	1146.06	95.50	0.10	2.29
LxB	2	42.36	21.18	0.01	340.19	170.10	0.39	382.56	95.64	0.10	3.19
L x P x B	6	1748.86	291.48	0.19	1975.47	329.25	0.75	3724.33	310.36	0.32	2.29
Error	48	73496.00	1531.17		21015.33	437.82		94511.33	984.49		
TSS	71	191791.65	2701.29		143620.00	2022.82		337045.16	2356.96		

Appendix LXVI: Effect of lime, phosphorus and boron on test weight of maize

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	114.61	114.61	113.56*	94.88	94.88	65.01*	209.49	104.74	84.86*	4.04
Phosphorus (P)	3	1.54	0.51	0.51	0.04	0.01	0.01	1.57	0.26	0.21	2.80
Boron (B)	2	0.36	0.18	0.18	0.21	0.11	0.07	0.58	0.14	0.12	3.19
L x P	3	4.25	1.42	1.40	7.71	2.57	1.76	11.96	1.99	1.61	2.80
P x B	6	1.13	0.19	0.19	2.14	0.36	0.24	3.27	0.27	0.22	2.29
L x B	2	0.09	0.04	0.04	0.25	0.13	0.09	0.34	0.09	0.07	3.19
L x P x B	6	0.57	0.09	0.09	0.27	0.05	0.03	0.84	0.07	0.06	2.29
Error	48	48.44	1.01		70.05	1.46		118.49	1.23		
TSS	71	170.98	2.41		175.55	2.47		346.53	2.42		

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	4439.96	4439.96	1882.81*	3987.25	3987.25	764.47*	8427.21	4213.60	1112.67*	4.04
Phosphorus (P)	3	2570.38	856.79	363.33*	2375.20	791.73	151.80*	4945.58	824.26	217.66*	2.80
Boron (B)	2	470.74	235.37	99.81*	968.01	484.00	92.80*	1438.74	359.69	94.98*	3.19
LxP	3	79.68	26.56	11.26*	45.76	15.25	2.92*	125.44	20.91	5.52*	2.80
P x B	6	4.60	0.77	0.32	6.70	1.12	0.21	11.30	0.94	0.25	2.29
LxB	2	1.19	0.59	0.25	0.56	0.28	0.05	1.75	0.44	0.12	3.19
L x P x B	6	4.50	0.75	0.32	5.85	0.97	0.19	10.35	0.86	0.23	2.29
Error	48	113.19	2.36		250.35	5.22		363.55	3.79		
TSS	71	7684.23	108.23		7639.67	107.60		15324.17	107.16		

Appendix LXVII: Effect of lime, phosphorus and boron on grain yield of maize

Appendix LXVIII: Effect of lime, phosphorus and boron on stover yield of maize

SOV	DE	2016			2017			Pooled			Etab
50V	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F tab
Lime (L)	1	10853.56	10853.56	303.70*	9078.33	9078.33	393.11*	19931.89	9965.94	338.80*	4.04
Phosphorus (P)	3	5357.51	1785.84	49.97*	6338.09	2112.70	91.48*	11695.60	1949.27	66.27*	2.80
Boron (B)	2	1393.30	696.65	19.49*	2217.16	1108.58	48.00*	3610.45	902.61	30.68*	3.19
LxP	3	45.04	15.01	0.42	5.52	1.84	0.08	50.56	8.43	0.29	2.80
РхВ	6	1.36	0.23	0.01	3.46	0.58	0.02	4.82	0.40	0.01	2.29
LxB	2	0.42	0.21	0.01	1.92	0.96	0.04	2.33	0.58	0.02	3.19
L x P x B	6	0.18	0.03	0.00	1.97	0.33	0.01	2.15	0.18	0.01	2.29
Error	48	1715.41	35.74		1108.49	23.09		2823.90	29.42		
TSS	71	19366.76	272.77		18754.95	264.15		38123.73	266.60		

Appendix LXIX: Effect of lime, phosphorus and boron on nitrogen content in grain of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	0.77	0.77	172.89*	0.72	0.72	188.53*	1.48	0.74	180.12*	4.04
Phosphorus (P)	3	0.13	0.04	9.53*	0.16	0.05	13.74*	0.28	0.05	11.47*	2.80
Boron (B)	2	0.14	0.07	15.99*	0.12	0.06	15.82*	0.26	0.07	15.91*	3.19
LxP	3	0.00	0.00	0.09	0.00	0.00	0.21	0.00	0.00	0.15	2.80
P x B	6	0.00	0.00	0.01	0.00	0.00	0.02	0.00	0.00	0.02	2.29
LxB	2	0.00	0.00	0.00	0.00	0.00	0.03	0.00	0.00	0.02	3.19
L x P x B	6	0.00	0.00	0.01	0.00	0.00	0.02	0.00	0.00	0.02	2.29
Error	48	0.21	0.00		0.18	0.00		0.40	0.00		
TSS	71	1.25	0.0176		1.18	0.0166		2.45	0.0171		

SOV	DF	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
Lime (L)	1	0.34	0.34	49.76*	0.22	0.22	36.60*	0.56	0.28	43.57*	4.04
Phosphorus (P)	3	0.07	0.02	3.57*	0.17	0.06	9.37*	0.24	0.04	6.30*	2.80
Boron (B)	2	0.08	0.04	5.56*	0.10	0.05	8.27*	0.18	0.04	6.84*	3.19
LxP	3	0.00	0.00	0.11	0.00	0.00	0.20	0.01	0.00	0.15	2.80
P x B	6	0.01	0.00	0.18	0.00	0.00	0.07	0.01	0.00	0.13	2.29
L x B	2	0.00	0.00	0.01	0.00	0.00	0.30	0.00	0.00	0.14	3.19
L x P x B	6	0.01	0.00	0.21	0.00	0.00	0.03	0.01	0.00	0.12	2.29
Error	48	0.33	0.01		0.29	0.01		0.62	0.01		
TSS	71	0.83	0.012		0.79	0.011		1.62	0.023		

Appendix LXX: Effect of lime, phosphorus and boron on nitrogen content in stover of maize

Appendix LXXI: Effect of lime, phosphorus and boron on phosphorus content in grain of maize

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	0.07	0.07	31.91*	0.08	0.08	29.13*	0.15	0.08	30.37*	4.04
Phosphorus (P)	3	0.10	0.03	15.22*	0.11	0.04	12.83*	0.21	0.04	13.89*	2.80
Boron (B)	2	0.03	0.02	6.81*	0.05	0.02	8.58*	0.08	0.02	7.79*	3.19
LxP	3	0.01	0.00	1.28	0.00	0.00	0.18	0.01	0.00	0.67	2.80
РхВ	6	0.00	0.00	0.12	0.00	0.00	0.24	0.01	0.00	0.19	2.29
LxB	2	0.00	0.00	0.12	0.00	0.00	0.03	0.00	0.00	0.07	3.19
L x P x B	6	0.00	0.00	0.05	0.00	0.00	0.02	0.00	0.00	0.04	2.29
Error	48	0.11	0.00		0.14	0.00		0.24	0.00		
TSS	71	0.33	0.0046		0.38	0.0054		0.73	0.0051		

Appendix LXXII: Effect of lime, phosphorus and boron on phosphorus content in stover of maize

SOV	DF	2016	2016			2017					Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	0.02	0.02	8.36*	0.01	0.01	5.28*	0.02	0.01	7.27*	4.04
Phosphorus (P)	3	0.02	0.01	3.34*	0.01	0.00	3.68*	0.03	0.01	3.46*	2.80
Boron (B)	2	0.01	0.01	3.65*	0.02	0.01	10.91*	0.04	0.01	6.22*	3.19
LxP	3	0.00	0.00	0.15	0.00	0.00	0.21	0.00	0.00	0.17	2.80
РхВ	6	0.00	0.00	0.11	0.00	0.00	0.24	0.00	0.00	0.16	2.29
LxB	2	0.00	0.00	0.03	0.00	0.00	0.37	0.00	0.00	0.15	3.19
L x P x B	6	0.00	0.00	0.14	0.00	0.00	0.22	0.00	0.00	0.17	2.29
Error	48	0.09	0.00		0.05	0.00		0.14	0.00		
TSS	71	0.14	0.0020		0.09	0.0013		0.25	0.0017		

SOV	DF	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	0.06	0.06	12.96*	0.06	0.06	10.97*	0.12	0.06	11.92*	4.04
Phosphorus (P)	3	0.06	0.02	3.83*	0.08	0.03	4.65*	0.13	0.02	4.26*	2.80
Boron (B)	2	0.00	0.00	0.34	0.00	0.00	0.09	0.00	0.00	0.21	3.19
LxP	3	0.00	0.00	0.07	0.00	0.00	0.01	0.00	0.00	0.04	2.80
P x B	6	0.00	0.00	0.02	0.00	0.00	0.01	0.00	0.00	0.01	2.29
LxB	2	0.00	0.00	0.04	0.00	0.00	0.05	0.00	0.00	0.04	3.19
L x P x B	6	0.00	0.00	0.01	0.00	0.00	0.01	0.00	0.00	0.01	2.29
Error	48	0.24	0.00		0.26	0.01		0.50	0.01		
TSS	71	0.36	0.0051		0.40	0.0056		0.80	0.0056		

Appendix LXXIII: Effect of lime, phosphorus and boron on potassium content in grain of maize

Appendix LXXIV: Effect of lime, phosphorus and boron on potassium content in stover of maize

SOV	DF	2016	2016			2017					Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	0.05	0.05	15.53*	0.06	0.06	21.71*	0.11	0.06	18.40*	4.04
Phosphorus (P)	3	0.09	0.03	8.90*	0.11	0.04	13.16*	0.20	0.03	10.89*	2.80
Boron (B)	2	0.00	0.00	0.29	0.00	0.00	0.30	0.00	0.00	0.29	3.19
LxP	3	0.00	0.00	0.05	0.00	0.00	0.56	0.01	0.00	0.29	2.80
РхВ	6	0.00	0.00	0.03	0.00	0.00	0.05	0.00	0.00	0.04	2.29
LxB	2	0.00	0.00	0.00	0.00	0.00	0.70	0.00	0.00	0.33	3.19
L x P x B	6	0.00	0.00	0.01	0.00	0.00	0.05	0.00	0.00	0.03	2.29
Error	48	0.16	0.00		0.14	0.00		0.30	0.00		
TSS	71	0.30	0.0042		0.33	0.0046		0.65	0.0045		

Appendix LXXV: Effect of lime, phosphorus and boron on calcium content in grain of maize

SOV	DF	2016	2016			2017					Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tau
Lime (L)	1	0.79	0.79	63.05*	0.76	0.76	58.27*	1.54	0.77	60.61*	4.04
Phosphorus (P)	3	0.15	0.05	4.05*	0.14	0.05	3.61*	0.29	0.05	3.82*	2.80
Boron (B)	2	0.01	0.00	0.30	0.01	0.00	0.20	0.01	0.00	0.25	3.19
L x P	3	0.02	0.01	0.61	0.02	0.01	0.41	0.04	0.01	0.51	2.80
P x B	6	0.01	0.00	0.07	0.00	0.00	0.04	0.01	0.00	0.05	2.29
LxB	2	0.00	0.00	0.15	0.00	0.00	0.01	0.00	0.00	0.08	3.19
LxPxB	6	0.00	0.00	0.07	0.00	0.00	0.01	0.01	0.00	0.04	2.29
Error	48	0.60	0.01		0.62	0.01		1.22	0.01		
TSS	71	1.58	0.022		1.55	0.022		3.13	0.022		

SOV	DF	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
Lime (L)	1	0.82	0.82	154.33*	0.87	0.87	143.70*	1.69	0.85	148.65*	4.04
Phosphorus (P)	3	0.12	0.04	7.82*	0.08	0.03	4.25*	0.20	0.03	5.91*	2.80
Boron (B)	2	0.00	0.00	0.28	0.00	0.00	0.13	0.00	0.00	0.20	3.19
LxP	3	0.00	0.00	0.08	0.00	0.00	0.08	0.00	0.00	0.08	2.80
P x B	6	0.00	0.00	0.07	0.00	0.00	0.06	0.00	0.00	0.07	2.29
LxB	2	0.00	0.00	0.06	0.00	0.00	0.03	0.00	0.00	0.04	3.19
L x P x B	6	0.00	0.00	0.03	0.00	0.00	0.07	0.00	0.00	0.05	2.29
Error	48	0.25	0.01		0.29	0.01		0.55	0.01		
TSS	71	1.20	0.017		1.25	0.018		2.45	0.017		

Appendix LXXVI: Effect of lime, phosphorus and boron on calcium content in stover of maize

Appendix LXXVII: Effect of lime, phosphorus and boron on boron content in grain of maize

SOV	DF	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	0.92	0.92	417.54*	0.92	0.92	424.38*	1.84	0.92	420.95*	4.04
Phosphorus (P)	3	0.00	0.00	0.58	0.00	0.00	0.72	0.01	0.00	0.65	2.80
Boron (B)	2	153.53	76.77	34916.01*	152.24	76.12	34931.43*	305.78	76.44	34923.68*	3.19
LxP	3	0.00	0.00	0.14	0.00	0.00	0.14	0.00	0.00	0.14	2.80
P x B	6	0.00	0.00	0.02	0.00	0.00	0.15	0.00	0.00	0.08	2.29
L x B	2	0.00	0.00	0.05	0.00	0.00	0.17	0.00	0.00	0.11	3.19
LxPxB	6	0.00	0.00	0.02	0.00	0.00	0.15	0.00	0.00	0.09	2.29
Error	48	0.11	0.00		0.10	0.00		0.21	0.00		
TSS	71	154.56	2.18		153.28	2.16		307.94	2.15		

Appendix LXXVIII: Effect of lime, phosphorus and boron on boron content in stover of maize

SOV	DF	2016	2016			2017					Etab
301	DI	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	0.72	0.72	197.58*	0.64	0.64	225.78*	1.36	0.68	209.88*	4.04
Phosphorus (P)	3	0.01	0.00	1.10	0.01	0.00	1.45	0.02	0.00	1.25	2.80
Boron (B)	2	16.10	8.05	2203.03*	15.84	7.92	2801.54*	31.94	7.99	2464.13*	3.19
LxP	3	0.00	0.00	0.10	0.00	0.00	0.43	0.00	0.00	0.24	2.80
РхВ	6	0.00	0.00	0.19	0.01	0.00	0.32	0.01	0.00	0.25	2.29
LxB	2	0.03	0.02	4.36*	0.04	0.02	7.68*	0.08	0.02	5.81*	3.19
L x P x B	6	0.00	0.00	0.03	0.00	0.00	0.27	0.01	0.00	0.13	2.29
Error	48	0.18	0.00		0.14	0.00		0.31	0.00		
TSS	71	17.05	0.24		16.69	0.24		33.76	0.24		

SOV J	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
Lime (L)	1	1791749.74	1791749.74	950.15*	1678643.32	1678643.32	878.71*	3470393.06	1735196.53	914.20*	4.04
Phosphorus (P)	3	602838.53	200946.18	106.56*	623138.00	207712.67	108.73*	1225976.53	204329.42	107.65*	2.80
Boron (B)	2	240969.70	120484.85	63.89*	336769.04	168384.52	88.14*	577738.74	144434.69	76.10*	3.19
LxP	3	18607.58	6202.53	3.29*	9978.64	3326.21	1.74	28586.22	4764.37	2.51	2.80
P x B	6	1048.31	174.72	0.09	889.59	148.27	0.08	1937.91	161.49	0.09	2.29
L x B	2	2012.04	1006.02	0.53	4325.69	2162.84	1.13	6337.72	1584.43	0.83	3.19
L x P x B	6	428.52	71.42	0.04	521.52	86.92	0.05	950.04	79.17	0.04	2.29
Error	48	90516.32	1885.76		91697.30	1910.36		182213.62	1898.06		
TSS	71	2748170.74	38706.63		2745963.09	38675.54		5505116.53	38497.32		

Appendix LXXIX: Effect of lime, phosphorus and boron on nitrogen uptake in grain of maize

Appendix LXXX: Effect of lime, phosphorus and boron on nitrogen uptake in stover of maize

SOV I	DE	2016		2017			Pooled			Etab	
50V	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	1724357.59	1724357.59	230.39*	1286207.51	1286207.51	163.06*	3010565.10	1505282.55	195.84*	4.04
Phosphorus (P)	3	478575.38	159525.13	21.31*	639972.64	213324.21	27.04*	1118548.02	186424.67	24.25*	2.80
Boron (B)	2	291804.05	145902.02	19.49*	401182.00	200591.00	25.43*	692986.05	173246.51	22.54*	3.19
L x P	3	25917.41	8639.14	1.15	11714.65	3904.88	0.50	37632.05	6272.01	0.82	2.80
PxB	6	6261.76	1043.63	0.14	7162.20	1193.70	0.15	13423.96	1118.66	0.15	2.29
L x B	2	5934.72	2967.36	0.40	17077.11	8538.55	1.08	23011.83	5752.96	0.75	3.19
LxPxB	6	9539.31	1589.88	0.21	622.22	103.70	0.01	10161.53	846.79	0.11	2.29
Error	48	359263.38	7484.65		378625.76	7888.04		737889.14	7686.35		
TSS	71	2901653.60	40868.36		2742564.08	38627.66		5644272.98	79496.80		

Appendix LXXXI: Effect of lime, phosphorus and boron on phosphorus uptake in grain of maize

SOV	DE	2016	2016		2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
Lime (L)	1	211646.69	211646.69	210.14*	222167.78	222167.78	179.72*	433814.47	216907.23	193.38*	4.04
Phosphorus (P)	3	173111.18	57703.73	57.29*	177517.44	59172.48	47.87*	350628.62	58438.10	52.10*	2.80
Boron (B)	2	42149.68	21074.84	20.92*	82896.85	41448.42	33.53*	125046.52	31261.63	27.87*	3.19
LxP	3	11784.01	3928.00	3.90*	4312.09	1437.36	1.16	16096.09	2682.68	2.39	2.80
P x B	6	2132.74	355.46	0.35	3503.25	583.88	0.47	5635.99	469.67	0.42	2.29
LxB	2	1144.97	572.48	0.57	1237.28	618.64	0.50	2382.25	595.56	0.53	3.19
LxPxB	6	992.13	165.36	0.16	900.50	150.08	0.12	1892.63	157.72	0.14	2.29
Error	48	48344.52	1007.18		59335.53	1236.16		107680.05	1121.67		
TSS	71	491305.91	6919.80		551870.72	7772.83		1056193.72	7385.97		

SOV D	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	81712.40	81712.40	34.22*	55053.36	55053.36	43.26*	136765.76	68382.88	37.37*	4.04
Phosphorus (P)	3	63026.14	21008.71	8.80*	52387.21	17462.40	13.72*	115413.35	19235.56	10.51*	2.80
Boron (B)	2	35748.19	17874.09	7.49*	57211.63	28605.82	22.48*	92959.82	23239.96	12.70*	3.19
LxP	3	3367.24	1122.41	0.47	1503.07	501.02	0.39	4870.31	811.72	0.44	2.80
P x B	6	1861.52	310.25	0.13	1335.92	222.65	0.17	3197.44	266.45	0.15	2.29
LxB	2	756.17	378.09	0.16	2712.49	1356.25	1.07	3468.66	867.17	0.47	3.19
L x P x B	6	1603.42	267.24	0.11	1720.55	286.76	0.23	3323.97	277.00	0.15	2.29
Error	48	114604.40	2387.59		61079.99	1272.50		175684.39	1830.05		
TSS	71	302679.48	4263.09		233004.22	3281.75		555622.79	3885.47		

Appendix LXXXII: Effect of lime, phosphorus and boron on phosphorus uptake in stover of maize

Appendix LXXXIII: Effect of lime, phosphorus and boron on potassium uptake in grain of maize

SOV D	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	303227.08	303227.08	156.14*	296237.00	296237.00	120.52*	599464.08	299732.04	136.24*	4.04
Phosphorus (P)	3	188188.95	62729.65	32.30*	210758.70	70252.90	28.58*	398947.65	66491.27	30.22*	2.80
Boron (B)	2	25586.81	12793.40	6.59*	38617.49	19308.74	7.86*	64204.30	16051.07	7.30*	3.19
LxP	3	5781.28	1927.09	0.99	1993.43	664.48	0.27	7774.71	1295.78	0.59	2.80
PxB	6	647.32	107.89	0.06	148.50	24.75	0.01	795.82	66.32	0.03	2.29
L x B	2	172.57	86.29	0.04	514.53	257.27	0.10	687.10	171.78	0.08	3.19
L x P x B	6	169.30	28.22	0.01	345.42	57.57	0.02	514.72	42.89	0.02	2.29
Error	48	93215.62	1941.99		117979.81	2457.91		211195.43	2199.95		
TSS	71	616988.92	8689.98		666594.89	9388.66		1306805.25	9138.50		

Appendix LXXXIV: Effect of lime, phosphorus and boron on potassium uptake in stover of maize

SOM	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
Lime (L)	1	2060700.37	2060700.37	239.74*	1822554.14	1822554.14	266.61*	3883254.51	1941627.26	251.64*	4.04
Phosphorus (P)	3	1347862.54	449287.51	52.27*	1590816.47	530272.16	77.57*	2938679.01	489779.83	63.48*	2.80
Boron (B)	2	214677.35	107338.67	12.49*	304021.57	152010.78	22.24*	518698.92	129674.73	16.81*	3.19
L x P	3	19512.89	6504.30	0.76	13762.85	4587.62	0.67	33275.75	5545.96	0.72	2.80
P x B	6	1186.91	197.82	0.02	3122.89	520.48	0.08	4309.80	359.15	0.05	2.29
L x B	2	501.21	250.61	0.03	9516.46	4758.23	0.70	10017.67	2504.42	0.32	3.19
L x P x B	6	465.92	77.65	0.01	2713.01	452.17	0.07	3178.93	264.91	0.03	2.29
Error	48	412589.48	8595.61		328126.25	6835.96		740715.73	7715.79		
TSS	71	4057496.68	57147.84		4074633.64	57389.21		8154990.75	57027.91		

SOV I	DE	2016	2016		2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
Lime (L)	1	867459.62	867459.62	139.24*	802965.51	802965.51	120.41*	1670425.13	835212.56	129.51*	4.04
Phosphorus (P)	3	241000.63	80333.54	12.89*	217455.14	72485.05	10.87*	458455.77	76409.29	11.85*	2.80
Boron (B)	2	1714.78	857.39	0.14	10317.46	5158.73	0.77	12032.24	3008.06	0.47	3.19
L x P	3	35415.57	11805.19	1.89	24004.70	8001.57	1.20	59420.26	9903.38	1.54	2.80
PxB	6	2416.92	402.82	0.06	1505.79	250.97	0.04	3922.71	326.89	0.05	2.29
L x B	2	1635.91	817.95	0.13	715.01	357.51	0.05	2350.92	587.73	0.09	3.19
L x P x B	6	1927.02	321.17	0.05	1092.38	182.06	0.03	3019.40	251.62	0.04	2.29
Error	48	299032.54	6229.84		320081.98	6668.37		619114.52	6449.11		
TSS	71	1450602.99	20431.03		1378137.97	19410.39		2831528.44	19800.90		

Appendix LXXXV: Effect of lime, phosphorus and boron on calcium uptake in grain of maize

Appendix LXXXVI: Effect of lime, phosphorus and boron on calcium uptake in stover of maize

SOV D	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	2290242.29	2290242.29	288.65*	2273554.22	2273554.22	328.64*	4563796.51	2281898.25	307.28*	4.04
Phosphorus (P)	3	503545.02	167848.34	21.15*	401366.29	133788.76	19.34*	904911.31	150818.55	20.31*	2.80
Boron (B)	2	17809.43	8904.71	1.12	39052.49	19526.25	2.82	56861.92	14215.48	1.91	3.19
LxP	3	20578.82	6859.61	0.86	7132.14	2377.38	0.34	27710.96	4618.49	0.62	2.80
P x B	6	2286.97	381.16	0.05	3061.32	510.22	0.07	5348.29	445.69	0.06	2.29
L x B	2	3561.46	1780.73	0.22	5349.45	2674.73	0.39	8910.91	2227.73	0.30	3.19
L x P x B	6	1168.46	194.74	0.02	3816.46	636.08	0.09	4984.92	415.41	0.06	2.29
Error	48	380853.01	7934.44		332064.23	6918.00		712917.24	7426.22		
TSS	71	3220045.45	45352.75		3065396.61	43174.60		6285769.0	43956.43		

Appendix LXXXVII: Effect of lime, phosphorus and boron on boron uptake in grain of maize

SOV I	DE	2016		2017			Pooled			Etab	
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	628750.29	628750.29	2170.35*	578161.93	578161.93	1009.21*	1206912.22	603456.11	1399.18*	4.04
Phosphorus (P)	3	305461.68	101820.56	351.47*	283972.06	94657.35	165.23*	589433.74	98238.96	227.78*	2.80
Boron (B)	2	1166180.58	583090.29	2012.73*	1392789.42	696394.71	1215.59*	2558970.00	639742.50	1483.31*	3.19
LxP	3	10051.76	3350.59	11.57*	5184.62	1728.21	3.02*	15236.39	2539.40	5.89*	2.80
P x B	6	5153.00	858.83	2.96*	5418.02	903.00	1.58	10571.01	880.92	2.04	2.29
LxB	2	8089.11	4044.55	13.96*	9959.18	4979.59	8.69*	18048.29	4512.07	10.46*	3.19
L x P x B	6	1089.20	181.53	0.63	901.03	150.17	0.26	1990.23	165.85	0.38	2.29
Error	48	13905.62	289.70		27498.53	572.89		41404.15	431.29		
TSS	71	2138681.25	30122.27		2303884.79	32449.08		4443892.40	31076.17		

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
Lime (L)	1	247011.95	247011.95	427.55*	211495.99	211495.99	608.50*	458507.94	229253.97	495.52*	4.04
Phosphorus (P)	3	85271.58	28423.86	49.20*	101774.50	33924.83	97.61*	187046.08	31174.35	67.38*	2.80
Boron (B)	2	334895.70	167447.85	289.83*	385713.52	192856.76	554.87*	720609.23	180152.31	389.39*	3.19
LxP	3	1201.30	400.43	0.69	63.23	21.08	0.06	1264.53	210.76	0.46	2.80
РхВ	6	1364.30	227.38	0.39	2101.61	350.27	1.01	3465.91	288.83	0.62	2.29
L x B	2	5572.14	2786.07	4.82*	5796.28	2898.14	8.34*	11368.42	2842.10	6.14*	3.19
L x P x B	6	13.83	2.31	0.00	47.72	7.95	0.02	61.55	5.13	0.01	2.29
Error	48	27731.43	577.74		16683.36	347.57		44414.79	462.65		
TSS	71	703062.24	9902.29		723676.22	10192.62		1426976.23	9978.85		

Appendix LXXXVIII: Effect of lime, phosphorus and boron on boron uptake in stover of maize

Appendix LXXXIX: Effect of lime, phosphorus and boron on pH of post-harvest soil

SOV	DF -	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lab
Lime (L)	1	3.43	3.43	162.66*	4.37	4.37	276.47*	7.80	3.90	211.44*	4.04
Phosphorus (P)	3	0.08	0.03	1.33	0.05	0.02	1.10	0.14	0.02	1.23	2.80
Boron (B)	2	0.00	0.00	0.03	0.00	0.00	0.05	0.00	0.00	0.04	3.19
LxP	3	0.01	0.00	0.15	0.02	0.01	0.36	0.03	0.00	0.24	2.80
РхВ	6	0.00	0.00	0.01	0.01	0.00	0.07	0.01	0.00	0.03	2.29
LxB	2	0.01	0.00	0.17	0.00	0.00	0.02	0.01	0.00	0.10	3.19
L x P x B	6	0.00	0.00	0.02	0.01	0.00	0.06	0.01	0.00	0.04	2.29
Error	48	1.01	0.02		0.76	0.02		1.77	0.02		
TSS	71	4.54	0.064		5.21	0.073		9.76	0.068		

Appendix XC: Effect of lime, phosphorus and boron on electrical conductivity of post-harvest soil

SOV I	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	0.01	0.01	3.30	0.00	0.00	1.07	0.01	0.01	2.09	4.04
Phosphorus (P)	3	0.01	0.00	1.33	0.00	0.00	0.36	0.01	0.00	0.80	2.80
Boron (B)	2	0.00	0.00	0.18	0.00	0.00	0.12	0.00	0.00	0.15	3.19
LxP	3	0.01	0.00	0.97	0.00	0.00	0.29	0.01	0.00	0.60	2.80
P x B	6	0.00	0.00	0.34	0.01	0.00	0.50	0.01	0.00	0.42	2.29
L x B	2	0.01	0.00	1.54	0.00	0.00	0.42	0.01	0.00	0.93	3.19
L x P x B	6	0.00	0.00	0.29	0.00	0.00	0.07	0.01	0.00	0.17	2.29
Error	48	0.11	0.00		0.13	0.00		0.24	0.00		
TSS	71	0.15	0.0021		0.15	0.0021		0.30	0.0042		

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r lau
Lime (L)	1	441.34	441.34	140.81*	449.25	449.25	137.45*	890.59	445.30	139.10*	4.04
Phosphorus (P)	3	2.01	0.67	0.21	2.44	0.81	0.25	4.45	0.74	0.23	2.80
Boron (B)	2	0.03	0.02	0.01	0.04	0.02	0.01	0.08	0.02	0.01	3.19
LxP	3	0.36	0.12	0.04	0.26	0.09	0.03	0.63	0.10	0.03	2.80
P x B	6	0.28	0.05	0.01	0.32	0.05	0.02	0.60	0.05	0.02	2.29
LxB	2	0.00	0.00	0.00	0.05	0.03	0.01	0.05	0.01	0.00	3.19
L x P x B	6	0.08	0.01	0.00	0.24	0.04	0.01	0.32	0.03	0.01	2.29
Error	48	150.45	3.13		156.89	3.27		307.33	3.20		
TSS	71	594.56	8.37		609.49	8.58		1220.42	17.19		

Appendix XCI: Effect of lime, phosphorus and boron on percent base saturation of post-harvest soil

Appendix XCII: Effect of lime, phosphorus and boron on organic carbon of post-harvest soil

SOV	DE	2016	2016					Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	F lao
Lime (L)	1	0.00	0.00	0.24	0.00	0.00	0.20	0.00	0.00	0.22	4.04
Phosphorus (P)	3	0.01	0.00	0.30	0.01	0.00	0.20	0.01	0.00	0.25	2.80
Boron (B)	2	0.01	0.00	0.36	0.00	0.00	0.01	0.01	0.00	0.18	3.19
LxP	3	0.00	0.00	0.01	0.00	0.00	0.01	0.00	0.00	0.01	2.80
РхВ	6	0.00	0.00	0.01	0.00	0.00	0.01	0.00	0.00	0.01	2.29
L x B	2	0.00	0.00	0.00	0.00	0.00	0.02	0.00	0.00	0.01	3.19
L x P x B	6	0.00	0.00	0.03	0.00	0.00	0.01	0.00	0.00	0.02	2.29
Error	48	0.46	0.01		0.50	0.01		0.95	0.01		
TSS	71	0.48	0.0068		0.51	0.0072		1.00	0.014		

Appendix XCIII: Effect of lime, phosphorus and boron on soil available nitrogen of post-harvest soil

SOV 1	DE	2016	2016		2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	1829.62	1829.62	533.05*	1504.35	1504.35	740.40*	3333.97	1666.99	610.16*	4.04
Phosphorus (P)	3	17.07	5.69	1.66	12.49	4.16	2.05	29.56	4.93	1.80	2.80
Boron (B)	2	4.46	2.23	0.65	7.40	3.70	1.82	11.85	2.96	1.08	3.19
L x P	3	20.11	6.70	1.95	4.60	1.53	0.75	24.71	4.12	1.51	2.80
РхВ	6	11.61	1.93	0.56	2.00	0.33	0.16	13.61	1.13	0.42	2.29
L x B	2	0.31	0.16	0.05	0.56	0.28	0.14	0.87	0.22	0.08	3.19
L x P x B	6	8.89	1.48	0.43	2.16	0.36	0.18	11.05	0.92	0.34	2.29
Error	48	164.75	3.43		97.53	2.03		262.28	2.73		
TSS	71	2056.82	28.97		1631.08	22.97		3707.44	25.93		

SOV	DE	2016			2017			Pooled			Etab
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	T tab
Lime (L)	1	47.26	47.26	62.19*	76.18	76.18	58.40*	123.43	61.72	59.79*	4.04
Phosphorus (P)	3	56.25	18.75	24.68*	63.18	21.06	16.14*	119.43	19.90	19.28*	2.80
Boron (B)	2	0.21	0.10	0.14	0.93	0.46	0.36	1.14	0.28	0.28	3.19
L x P	3	3.51	1.17	1.54	1.25	0.42	0.32	4.76	0.79	0.77	2.80
P x B	6	2.14	0.36	0.47	0.55	0.09	0.07	2.69	0.22	0.22	2.29
LxB	2	0.04	0.02	0.03	0.10	0.05	0.04	0.14	0.03	0.03	3.19
L x P x B	6	1.32	0.22	0.29	1.00	0.17	0.13	2.32	0.19	0.19	2.29
Error	48	36.47	0.76		62.62	1.30		99.09	1.03		
TSS	71	147.20	2.07		205.81	2.90		353.01	2.47		

Appendix XCIV: Effect of lime, phosphorus and boron on soil available phosphorus of post-harvest soil

Appendix XCV: Effect of lime, phosphorus and boron on soil available potassium of post-harvest soil

SOM	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	21.59	21.59	10.63*	46.29	46.29	16.91*	67.88	33.94	14.23*	4.04
Phosphorus (P)	3	2.27	0.76	0.37	5.37	1.79	0.65	7.63	1.27	0.53	2.80
Boron (B)	2	2.25	1.12	0.55	2.93	1.46	0.53	5.17	1.29	0.54	3.19
L x P	3	13.84	4.61	2.27	13.27	4.42	1.62	27.11	4.52	1.90	2.80
РхВ	6	14.30	2.38	1.17	12.37	2.06	0.75	26.67	2.22	0.93	2.29
L x B	2	0.89	0.45	0.22	9.59	4.80	1.75	10.49	2.62	1.10	3.19
L x P x B	6	4.99	0.83	0.41	13.27	2.21	0.81	18.26	1.52	0.64	2.29
Error	48	97.50	2.03		131.42	2.74		228.92	2.38		
TSS	71	157.64	2.22		234.50	3.30		397.82	5.60		

Appendix XCVI: Effect of lime, phosphorus and boron on soil exchangeable calcium of post-harvest soil

SOV	DE	2016			2017			Pooled			Etab
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	T tab
Lime (L)	1	25.32	25.32	3162.69*	26.15	26.15	4373.27*	51.47	25.74	3680.22*	4.04
Phosphorus (P)	3	13.36	4.45	556.21*	13.12	4.37	731.61*	26.48	4.41	631.19*	2.80
Boron (B)	2	0.00	0.00	0.07	0.00	0.00	0.09	0.00	0.00	0.08	3.19
L x P	3	0.00	0.00	0.02	0.01	0.00	0.34	0.01	0.00	0.15	2.80
РхВ	6	0.00	0.00	0.01	0.00	0.00	0.06	0.00	0.00	0.03	2.29
L x B	2	0.00	0.00	0.00	0.00	0.00	0.13	0.00	0.00	0.06	3.19
L x P x B	6	0.00	0.00	0.02	0.01	0.00	0.17	0.01	0.00	0.08	2.29
Error	48	0.38	0.01		0.29	0.01		0.67	0.01		
TSS	71	39.07	0.55		39.58	0.56		78.68	0.55		

SOV	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	I tab
Lime (L)	1	0.05	0.05	5.97*	0.03	0.03	4.54*	0.08	0.04	5.36*	4.04
Phosphorus (P)	3	0.00	0.00	0.05	0.01	0.00	0.26	0.01	0.00	0.14	2.80
Boron (B)	2	0.17	0.09	10.04*	0.10	0.05	7.93*	0.27	0.07	9.14*	3.19
LxP	3	0.00	0.00	0.15	0.00	0.00	0.06	0.01	0.00	0.11	2.80
P x B	6	0.00	0.00	0.08	0.01	0.00	0.25	0.01	0.00	0.15	2.29
LxB	2	0.03	0.01	1.70	0.01	0.00	0.53	0.04	0.01	1.20	3.19
L x P x B	6	0.07	0.01	1.40	0.02	0.00	0.52	0.09	0.01	1.02	2.29
Error	48	0.41	0.01		0.31	0.01		0.72	0.01		
TSS	71	0.75	0.011		0.48	0.0068		1.26	0.018		

Appendix XCVII: Effect of lime, phosphorus and boron on soil available boron of post-harvest soil

Appendix XCVIII: Effect of lime, phosphorus and boron on soil exchangeable hydrogen of post-harvest soil

SOM	DE	2016			2017			Pooled			Etab
301	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	3.56	3.56	671.70*	1.35	1.35	529.81*	4.91	2.46	625.62*	4.04
Phosphorus (P)	3	0.01	0.00	0.72	0.01	0.00	0.68	0.02	0.00	0.71	2.80
Boron (B)	2	0.00	0.00	0.02	0.00	0.00	0.18	0.00	0.00	0.07	3.19
LxP	3	0.03	0.01	1.78	0.01	0.00	1.84	0.04	0.01	1.80	2.80
РхВ	6	0.00	0.00	0.02	0.00	0.00	0.19	0.00	0.00	0.08	2.29
L x B	2	0.00	0.00	0.21	0.00	0.00	0.34	0.00	0.00	0.25	3.19
L x P x B	6	0.00	0.00	0.03	0.00	0.00	0.14	0.00	0.00	0.07	2.29
Error	48	0.25	0.01		0.12	0.00		0.38	0.00		
TSS	71	3.86	0.054		1.50	0.021		5.36	0.075		

Appendix XCIX: Effect of lime, phosphorus and boron on soil exchangeable aluminium of post-harvest soil

SOV	DE	2016			2017			Pooled	Pooled		
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tab
Lime (L)	1	0.17	0.17	42.58*	0.30	0.30	109.59*	0.47	0.23	70.45*	4.04
Phosphorus (P)	3	0.01	0.00	0.55	0.00	0.00	0.27	0.01	0.00	0.43	2.80
Boron (B)	2	0.00	0.00	0.46	0.00	0.00	0.73	0.01	0.00	0.57	3.19
LxP	3	0.02	0.01	1.91	0.02	0.01	1.84	0.04	0.01	1.88	2.80
P x B	6	0.00	0.00	0.01	0.00	0.00	0.05	0.00	0.00	0.03	2.29
LxB	2	0.00	0.00	0.04	0.00	0.00	0.02	0.00	0.00	0.03	3.19
L x P x B	6	0.00	0.00	0.04	0.00	0.00	0.01	0.00	0.00	0.03	2.29
Error	48	0.19	0.00		0.13	0.00		0.32	0.00		
TSS	71	0.39	0.0055		0.46	0.0065		0.85	0.012		

SOV	DE	2016			2017			Pooled			Etab
	DF	SS	MSS	F cal	SS	MSS	F cal	SS	MSS	F cal	r tao
Lime (L)	1	52.14	52.14	5618.10*	49.78	49.78	9520.36*	101.92	50.96	7024.44*	4.04
Phosphorus (P)	3	0.01	0.00	0.51	0.02	0.01	1.20	0.03	0.01	0.76	2.80
Boron (B)	2	0.01	0.00	0.36	0.01	0.00	0.68	0.01	0.00	0.47	3.19
L x P	3	0.02	0.01	0.85	0.02	0.01	1.20	0.04	0.01	0.97	2.80
РхВ	6	0.00	0.00	0.01	0.00	0.00	0.04	0.00	0.00	0.02	2.29
L x B	2	0.00	0.00	0.01	0.00	0.00	0.16	0.00	0.00	0.06	3.19
L x P x B	6	0.00	0.00	0.02	0.00	0.00	0.03	0.00	0.00	0.03	2.29
Error	48	0.45	0.01		0.25	0.01		0.70	0.01		
TSS	71	52.63	0.74		50.08	0.71		102.81	0.72		

Appendix C: Effect of lime, phosphorus and boron on total potential acidity of post-harvest soil